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(REV 10-95)

U.S. DEPARTMENT OF COMMERCE PATENT AND TRADEMARK OFFICE

ATTORNEY'S DOCKET NUMBER

BB-1170

**TRANSMITTAL LETTER TO THE UNITED STATES
DESIGNATED/ELECTED OFFICE (DO/EO/US)
CONCERNING A FILING UNDER 35 U.S.C. 371**

U.S. APPLICATION NO. (IF KNOWN, SEE 37 CFR)

09/720383

INTERNATIONAL APPLICATION NO.

DEC 21 2000 PCT/US99/15871

INTERNATIONAL FILING DATE

13 JULY 1999 (13.07.99)

PRIORITY DATE CLAIMED

14 JULY 1998 (14.07.98)

TITLE OF INVENTION

PLANNING CELLULOSE SYNTHASES

APPLICANT(S) FOR DO/EO/US

ALLEN, Stephen M. et al.

Applicant herewith submits to the United States Designated/Elected Office (DO/EO/US) the following items and other information

1. ☒ This is a **FIRST** submission of items concerning a filing under 35 U.S.C. 371.
2. ☐ This is a **SECOND** or **SUBSEQUENT** submission of items concerning a filing under 35 U.S.C. 371.
3. ☒ This is an express request to being national examination procedures (35 U.S.C. 371(f)) at any time rather than delay examination until the expiration of the applicable time limit set in 35 U.S.C. 371(b)) and PCT Articles 22 and 39(1).
4. ☒ A proper Demand for International Preliminary Examination was made by the 19th month from the earliest claimed priority date.
5. ☒ A copy of the International Application was filed (35 U.S.C. 371 (c) (2))
 - a. ☒ is transmitted herewith (required only if not transmitted by the International Bureau.
 - b. ☐ has been transmitted by the International Bureau.
 - c. ☐ is not required, as the application was filed in the United States Receiving Office (RO/US)
6. ☐ A translation of the International Application into English (35 U.S.C. 371 (c) (2)).
7. ☒ A copy of the International Search Report (PCT/ISA/210).
8. ☒ Amendments to the claims of the International Application under PCT Article 19 (35 U.S.C. 371 (c) (3))
 - a. ☐ are transmitted herewith (required only if not transmitted by the International Bureau).
 - b. ☐ have been transmitted by the International Bureau.
 - c. ☐ have not been made; however, the time limit for making such amendments has NOT expired.
 - d. ☒ have not been made and will not be made.
9. ☐ A translation of the amendments to the claims under PCT Article 19 (35 U.S.C. 371 (c)(3)).
10. ☒ An oath or declaration of the inventor(s) (35 U.S.C. 371 (c)(4)).
11. ☐ A copy of the International Preliminary Examination Report (PCT/IPEA/409)
12. ☐ A translation of the annexes to the International Preliminary Examination Report under PCT Article 36 (35 U.S.C. 371 (c)(5)).

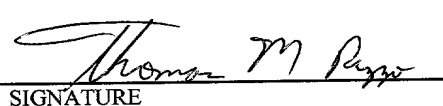
Items 13 to 18 below concern document(s) or information included :

13. ☐ An Information Disclosure Statement under 37 CFR 1.97 and 1.98.
14. ☒ An assignment document for recording. A separate cover sheet in compliance with 37 CFR 3.28 and 3.31 is included.
15. ☐ A **FIRST** preliminary amendment.
A **SECOND** or **SUBSEQUENT** preliminary amendment.
16. ☐ A substitute specification.
17. ☒ A change of power of attorney and/or address letter.
18. ☒ Certificate of Mailing by Express Mail.
19. ☐ Other items or information:

17. General Power of Attorney

18. Express Mailing Label No.: EL031052215US

Marked
12/21/2000

APPLICATION NO. (IF KNOWN, SEE 37 CFR) 09/720383		INTERNATIONAL APPLICATION NO. PCT/US99/15871		ATTORNEY'S DOCKET NUMBER BB-1170	
20. The following fees are submitted				CALCULATIONS PTO USE ONLY	
BASIC NATIONAL FEE (37 CFR 1.492 (a) (1) - (5)) :					
<input checked="" type="checkbox"/> Search Report has been prepared by the EPO or JPO \$840.00					
<input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) \$690.00					
<input type="checkbox"/> No international preliminary examination fee paid to USPTO (37 CFR 1.482) but international search fee paid to USPTO (37 CFR 1.445(a)(2)) \$760.00					
<input type="checkbox"/> Neither international preliminary examination fee paid to USPTO (37 CFR 1.482) nor international search fee (37 CFR 1.445(a)(2)) paid to USPTO \$1000.00					
<input type="checkbox"/> International preliminary examination fee paid to USPTO (37 CFR 1.482) And all claims satisfied provisions of PCT Article 33(2)-(4) \$ 100.00					
ENTER APPROPRIATE BASIC FEE AMOUNT =				\$840.00	
Surcharge of \$130.00 for furnishing the oath or declaration later than months from the earliest claimed priority date (37 CFR 1.492 (e)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				\$0.00	
CLAIMS	NUMBER FILED	NUMBER EXTRA	RATE		
Total Claims	21 - 20 =	1 x	\$18.00	\$18.00	
Independent Claims	6 - 3 =	1 x	\$80.00	\$240.00	
Multiple Dependent Claims (check if applicable)			<input checked="" type="checkbox"/>	\$260.00	
TOTAL OF ABOVE CALCULATIONS =				\$518.00	
Reduction of 1/2 for filing by small entity, if applicable. Verified Small Entity Statement must also be filed (Note 37 CFR 1.9, 1.27, 1.28) (check if applicable). <input type="checkbox"/>				\$0.00	
SUBTOTAL =				\$518.00	
Processing Fee of \$130.00 for furnishing the English translation later than months from the earliest claimed priority date (37 CFR 1.492 (f)). <input type="checkbox"/> 20 <input type="checkbox"/> 30				\$0.00	
TOTAL NATIONAL FEE =				\$1,358.00	
Fee for recording the enclosed assignment (37 CFR 1.21(h)). The assignment must be accompanied by an appropriate cover sheet (37 CFR 3.28, 3.31) (check if applicable). <input checked="" type="checkbox"/>				\$40.00	
TOTAL FEES ENCLOSED =				\$1,398.00	
				Amount to be : refunded	\$
				Charged	\$
<input type="checkbox"/> A check in the amount of _____ to cover the above fees enclosed.					
<input checked="" type="checkbox"/> Please charge my Deposit Account No. 04-1928 in the amount of \$1,398.00 to cover the above fees.					
<input checked="" type="checkbox"/> The Commissioner is hereby authorized to charge any fees which may be required, or credit any overpayment to Deposit Account No. 04-1928 a duplicate copy of this sheet is enclosed.					
NOTE : Where an appropriate time limit under 37 CFR 1.494 or 1.495 has not been met, a petition to revive (CFR 1.37(a) or (b)) must be filed and granted to restore the application to pending status.					
SEND ALL CORRESPONDENCE TO:					
RIZZO, THOMAS M. E. I. DU PONT DE NEMOURS AND COMPANY LEGAL PATENT RECORDS CENTER 1007 MARKET STREET WILMINGTON, DELAWARE 19898 UNITED STATES OF AMERICA			<div> SIGNATURE</div> <div>RIZZO, THOMAS M. NAME</div> <div>41,272 REGISTRATION NUMBER</div> <div>December 14, 2000 DATE</div>		

TITLEPLANT CELLULOSE SYNTHASES

This application claims the benefit of U.S. Provisional Application No. 60/092,844, filed July 14, 1998.

FIELD OF THE INVENTION

This invention is in the field of plant molecular biology. More specifically, this invention pertains to nucleic acid fragments encoding cellulose biosynthetic enzymes in plants and seeds.

BACKGROUND OF THE INVENTION

Cellulose is a major component of plant fiber, e.g. cotton fiber. Cellulose is composed of crystalline beta-1,4-glucan microfibrils (see World Patent Publication No. WO 98/00549). These microfibrils are strong and can resist enzymatic and mechanical degradation and are important in determining nutritional quality of animal and human foodstuffs. Hence, modification of the biosynthetic pathway responsible for cellulose synthesis through modification of cellulose synthase activity could potentially alter fiber quantity, either by producing more or less fiber in a particular plant species or in a specific organ or tissue of a particular plant. Modification of cellulose synthase activity could increase the value of the fiber to the end-user and may improve the structural integrity of the plant cell wall. Lastly, because cellulose is a major cell wall component, inhibition of cellulose synthesis would probably be lethal. Thus, cellulose synthase may serve as the target for a novel class of herbicides. Plant cellulose synthase genes, homologs of the bacterial *celA* genes encoding the catalytic subunit of cellulose synthase, have been reported from cotton, *Arabidopsis*, rice and alfalfa (World Patent Publication Nos. WO 98/00549 and WO 98/18949).

There is a great deal of interest in identifying the genes that encode proteins involved in cellulose synthesis. These genes may be used in plant cells to control the synthesis of cellulose. Accordingly, the availability of nucleic acid sequences encoding all or a portion of a cellulose synthase would facilitate studies to better understand cellulose synthesis in plants and provide genetic tools to alter cellulose production.

SUMMARY OF THE INVENTION

The instant invention relates to isolated nucleic acid fragments encoding cellulose biosynthesis enzymes. Specifically, this invention concerns an isolated nucleic acid fragment encoding a cellulose synthase and an isolated nucleic acid fragment that is substantially similar to an isolated nucleic acid fragment encoding a cellulose synthase. In addition, this invention relates to a nucleic acid fragment that is complementary to the nucleic acid fragment encoding cellulose synthase. An additional embodiment of the instant invention pertains to a polypeptide encoding all or a substantial portion of a cellulose synthase.

In another embodiment, the instant invention relates to a chimeric gene encoding a cellulose synthase, or to a chimeric gene that comprises a nucleic acid fragment that is complementary to a nucleic acid fragment encoding a cellulose synthase, operably linked to suitable regulatory sequences, wherein expression of the chimeric gene results in production of levels of the encoded protein in a transformed host cell that is altered (i.e., increased or decreased) from the level produced in an untransformed host cell.

In a further embodiment, the instant invention concerns a transformed host cell comprising in its genome a chimeric gene encoding a cellulose synthase, operably linked to suitable regulatory sequences. Expression of the chimeric gene results in production of altered levels of the encoded protein in the transformed host cell. The transformed host cell can be of eukaryotic or prokaryotic origin, and include cells derived from higher plants and microorganisms. The invention also includes transformed plants that arise from transformed host cells of higher plants, and seeds derived from such transformed plants.

An additional embodiment of the instant invention concerns a method of altering the level of expression of a cellulose synthase in a transformed host cell comprising:
a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a cellulose synthase; and b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of altered levels of cellulose synthase in the transformed host cell.

An addition embodiment of the instant invention concerns a method for obtaining a nucleic acid fragment encoding all or a substantial portion of an amino acid sequence encoding a cellulose synthase.

A further embodiment of the instant invention is a method for evaluating at least one compound for its ability to inhibit the activity of a cellulose synthase, the method comprising the steps of: (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a cellulose synthase, operably linked to suitable regulatory sequences; (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of cellulose synthase in the transformed host cell; (c) optionally purifying the cellulose synthase expressed by the transformed host cell; (d) treating the cellulose synthase with a compound to be tested; and (e) comparing the activity of the cellulose synthase that has been treated with a test compound to the activity of an untreated cellulose synthase, thereby selecting compounds with potential for inhibitory activity.

BRIEF DESCRIPTION OF THE DRAWINGS AND SEQUENCE DESCRIPTIONS

The invention can be more fully understood from the following detailed description and the accompanying drawings and Sequence Listing which form a part of this application.

Figure 1 shows a comparison of the amino acid sequences set forth in SEQ ID NOs:2, 4, 8, 10, 12, 14, 16, 18, 20 and 22 and the *Arabidopsis thaliana* sequences (SEQ ID NOs:23 (gi 2827139), 24 (gi 2827141), 26 (gi 4467125), 27 (gi 4886756) and 29 (gi 3135611)) and *Gossypium hirsutum* sequences (SEQ ID NOs:25 (gi 1706958) and 28 (gi 5081779)).

5 Table 1 lists the polypeptides that are described herein, the designation of the cDNA clones that comprise the nucleic acid fragments encoding polypeptides representing all or a substantial portion of these polypeptides, and the corresponding identifier (SEQ ID NO:) as used in the attached Sequence Listing. The sequence descriptions and Sequence Listing attached hereto comply with the rules governing nucleotide and/or amino acid sequence
10 disclosures in patent applications as set forth in 37 C.F.R. §1.821-1.825.

TABLE 1
Cellulose Biosynthetic Enzymes

Protein	Clone Designation	SEQ ID NO:	
		(Nucleotide)	(Amino Acid)
Cellulose Synthase	bsh1.pk0002.f6	1	2
Cellulose Synthase	Contig composed of: cco1n.pk0005.g3 cdt2c.pk002.g1 cdt2c.pk002.l16 csc1c.pk002.i1 p0031.ccmar05rb p0110.cgsma57r	3	4
Cellulose Synthase	cr1n.pk0135.e10	5	6
Cellulose Synthase	p0097.cqrad17rc	7	8
Cellulose Synthase	p0122.ckamh70rc	9	10
Cellulose Synthase	rlr24.pk0073.g1	11	12
Cellulose Synthase	sdp2c.pk005.o22	13	14
Cellulose Synthase	ses8w.pk0028.f3	15	16
Cellulose Synthase	ssl.pk0036.c10	17	18
Cellulose Synthase	Contig composed of: wl1.pk0009.c9 wr1.pk0160.d11 wre1n.pk0043.f9 wre1n.pk0043.h8 wre1n.pk0131.g10	19	20
Cellulose Synthase	wl1n.pk0044.b1	21	22

15 The Sequence Listing contains the one letter code for nucleotide sequence characters and the three letter codes for amino acids as defined in conformity with the IUPAC-IUBMB standards described in *Nucleic Acids Research* 13:3021-3030 (1985) and in the *Biochemical Journal* 219 (No. 2):345-373 (1984) which are herein incorporated by reference. The

symbols and format used for nucleotide and amino acid sequence data comply with the rules set forth in 37 C.F.R. §1.822.

DETAILED DESCRIPTION OF THE INVENTION

In the context of this disclosure, a number of terms shall be utilized. As used herein, a
5 "nucleic acid fragment" is a polymer of RNA or DNA that is single- or double-stranded, optionally containing synthetic, non-natural or altered nucleotide bases. A nucleic acid fragment in the form of a polymer of DNA may be comprised of one or more segments of cDNA, genomic DNA or synthetic DNA.

As used herein, "contig" refers to a nucleotide sequence that is assembled from two or
10 more constituent nucleotide sequences that share common or overlapping regions of sequence homology. For example, the nucleotide sequences of two or more nucleic acid fragments can be compared and aligned in order to identify common or overlapping sequences. Where common or overlapping sequences exist between two or more nucleic acid fragments, the sequences (and thus their corresponding nucleic acid fragments) can be
15 assembled into a single contiguous nucleotide sequence.

As used herein, "substantially similar" refers to nucleic acid fragments wherein changes in one or more nucleotide bases results in substitution of one or more amino acids, but do not affect the functional properties of the polypeptide encoded by the nucleotide sequence. "Substantially similar" also refers to nucleic acid fragments wherein changes in
20 one or more nucleotide bases does not affect the ability of the nucleic acid fragment to mediate alteration of gene expression by gene silencing through for example antisense or co-suppression technology. "Substantially similar" also refers to modifications of the nucleic acid fragments of the instant invention such as deletion or insertion of one or more nucleotides that do not substantially affect the functional properties of the resulting
25 transcript vis-à-vis the ability to mediate gene silencing or alteration of the functional properties of the resulting protein molecule. It is therefore understood that the invention encompasses more than the specific exemplary nucleotide or amino acid sequences and includes functional equivalents thereof.

For example, it is well known in the art that antisense suppression and co-suppression
30 of gene expression may be accomplished using nucleic acid fragments representing less than the entire coding region of a gene, and by nucleic acid fragments that do not share 100% sequence identity with the gene to be suppressed. Moreover, alterations in a nucleic acid fragment which result in the production of a chemically equivalent amino acid at a given site, but do not effect the functional properties of the encoded polypeptide, are well known in
35 the art. Thus, a codon for the amino acid alanine, a hydrophobic amino acid, may be substituted by a codon encoding another less hydrophobic residue, such as glycine, or a more hydrophobic residue, such as valine, leucine, or isoleucine. Similarly, changes which result in substitution of one negatively charged residue for another, such as aspartic acid for

glutamic acid, or one positively charged residue for another, such as lysine for arginine, can also be expected to produce a functionally equivalent product. Nucleotide changes which result in alteration of the N-terminal and C-terminal portions of the polypeptide molecule would also not be expected to alter the activity of the polypeptide. Each of the proposed
5 modifications is well within the routine skill in the art, as is determination of retention of biological activity of the encoded products.

Moreover, substantially similar nucleic acid fragments may also be characterized by their ability to hybridize, under stringent conditions (0.1X SSC, 0.1% SDS, 65°C), with the nucleic acid fragments disclosed herein.

10 Substantially similar nucleic acid fragments of the instant invention may also be characterized by the percent identity of the amino acid sequences that they encode to the amino acid sequences disclosed herein, as determined by algorithms commonly employed by those skilled in this art. Preferred are those nucleic acid fragments whose nucleotide
15 sequences encode amino acid sequences that are 80% identical to the amino acid sequences reported herein. More preferred nucleic acid fragments encode amino acid sequences that are 90% identical to the amino acid sequences reported herein. Most preferred are nucleic acid fragments that encode amino acid sequences that are 95% identical to the amino acid sequences reported herein. Sequence alignments and percent identity calculations were performed using the Megalign program of the LASARGENE bioinformatics computing suite
20 (DNASTAR Inc., Madison, WI). Multiple alignment of the sequences was performed using the Clustal method of alignment (Higgins and Sharp (1989) *CABIOS* 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH PENALTY=10). Default parameters for pairwise alignments using the Clustal method were KTUPLE 1, GAP PENALTY=3, WINDOW=5 and DIAGONALS SAVED=5.

25 A "substantial portion" of an amino acid or nucleotide sequence comprises an amino acid or a nucleotide sequence that is sufficient to afford putative identification of the protein or gene that the amino acid or nucleotide sequence comprises. Amino acid and nucleotide sequences can be evaluated either manually by one skilled in the art, or by using computer-based sequence comparison and identification tools that employ algorithms such as BLAST
30 (Basic Local Alignment Search Tool; Altschul et al. (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/). In general, a sequence of ten or more contiguous amino acids or thirty or more contiguous nucleotides is necessary in order to putatively identify a polypeptide or nucleic acid sequence as homologous to a known protein or gene. Moreover, with respect to nucleotide sequences, gene-specific oligonucleotide probes
35 comprising 30 or more contiguous nucleotides may be used in sequence-dependent methods of gene identification (e.g., Southern hybridization) and isolation (e.g., *in situ* hybridization of bacterial colonies or bacteriophage plaques). In addition, short oligonucleotides of 12 or more nucleotides may be used as amplification primers in PCR in order to obtain a particular

nucleic acid fragment comprising the primers. Accordingly, a "substantial portion" of a nucleotide sequence comprises a nucleotide sequence that will afford specific identification and/or isolation of a nucleic acid fragment comprising the sequence. The instant specification teaches amino acid and nucleotide sequences encoding polypeptides that

5 comprise one or more particular plant proteins. The skilled artisan, having the benefit of the sequences as reported herein, may now use all or a substantial portion of the disclosed sequences for purposes known to those skilled in this art. Accordingly, the instant invention comprises the complete sequences as reported in the accompanying Sequence Listing, as well as substantial portions of those sequences as defined above.

10 "Codon degeneracy" refers to divergence in the genetic code permitting variation of the nucleotide sequence without effecting the amino acid sequence of an encoded polypeptide. Accordingly, the instant invention relates to any nucleic acid fragment comprising a nucleotide sequence that encodes all or a substantial portion of the amino acid sequences set forth herein. The skilled artisan is well aware of the "codon-bias" exhibited

15 by a specific host cell in usage of nucleotide codons to specify a given amino acid. Therefore, when synthesizing a nucleic acid fragment for improved expression in a host cell, it is desirable to design the nucleic acid fragment such that its frequency of codon usage approaches the frequency of preferred codon usage of the host cell.

"Synthetic nucleic acid fragments" can be assembled from oligonucleotide building

20 blocks that are chemically synthesized using procedures known to those skilled in the art. These building blocks are ligated and annealed to form larger nucleic acid fragments which may then be enzymatically assembled to construct the entire desired nucleic acid fragment. "Chemically synthesized", as related to nucleic acid fragment, means that the component nucleotides were assembled *in vitro*. Manual chemical synthesis of nucleic acid fragments

25 may be accomplished using well established procedures, or automated chemical synthesis can be performed using one of a number of commercially available machines. Accordingly, the nucleic acid fragments can be tailored for optimal gene expression based on optimization of nucleotide sequence to reflect the codon bias of the host cell. The skilled artisan appreciates the likelihood of successful gene expression if codon usage is biased towards

30 those codons favored by the host. Determination of preferred codons can be based on a survey of genes derived from the host cell where sequence information is available.

"Gene" refers to a nucleic acid fragment that expresses a specific protein, including regulatory sequences preceding (5' non-coding sequences) and following (3' non-coding sequences) the coding sequence. "Native gene" refers to a gene as found in nature with its

35 own regulatory sequences. "Chimeric gene" refers any gene that is not a native gene, comprising regulatory and coding sequences that are not found together in nature. Accordingly, a chimeric gene may comprise regulatory sequences and coding sequences that are derived from different sources, or regulatory sequences and coding sequences derived

from the same source, but arranged in a manner different than that found in nature.

“Endogenous gene” refers to a native gene in its natural location in the genome of an organism. A “foreign” gene refers to a gene not normally found in the host organism, but that is introduced into the host organism by gene transfer. Foreign genes can comprise native genes inserted into a non-native organism, or chimeric genes. A “transgene” is a gene that has been introduced into the genome by a transformation procedure.

“Coding sequence” refers to a nucleotide sequence that codes for a specific amino acid sequence. “Regulatory sequences” refer to nucleotide sequences located upstream (5' non-coding sequences), within, or downstream (3' non-coding sequences) of a coding sequence, and which influence the transcription, RNA processing or stability, or translation of the associated coding sequence. Regulatory sequences may include promoters, translation leader sequences, introns, and polyadenylation recognition sequences.

“Promoter” refers to a nucleotide sequence capable of controlling the expression of a coding sequence or functional RNA. In general, a coding sequence is located 3' to a promoter sequence. The promoter sequence consists of proximal and more distal upstream elements, the latter elements often referred to as enhancers. Accordingly, an “enhancer” is a nucleotide sequence which can stimulate promoter activity and may be an innate element of the promoter or a heterologous element inserted to enhance the level or tissue-specificity of a promoter. Promoters may be derived in their entirety from a native gene, or be composed of different elements derived from different promoters found in nature, or even comprise synthetic nucleotide segments. It is understood by those skilled in the art that different promoters may direct the expression of a gene in different tissues or cell types, or at different stages of development, or in response to different environmental conditions. Promoters which cause a nucleic acid fragment to be expressed in most cell types at most times are commonly referred to as “constitutive promoters”. New promoters of various types useful in plant cells are constantly being discovered; numerous examples may be found in the compilation by Okamuro and Goldberg (1989) *Biochemistry of Plants* 15:1-82. It is further recognized that since in most cases the exact boundaries of regulatory sequences have not been completely defined, nucleic acid fragments of different lengths may have identical promoter activity.

The “translation leader sequence” refers to a nucleotide sequence located between the promoter sequence of a gene and the coding sequence. The translation leader sequence is present in the fully processed mRNA upstream of the translation start sequence. The translation leader sequence may affect processing of the primary transcript to mRNA, mRNA stability or translation efficiency. Examples of translation leader sequences have been described (Turner and Foster (1995) *Molecular Biotechnology* 3:225).

The “3' non-coding sequences” refer to nucleotide sequences located downstream of a coding sequence and include polyadenylation recognition sequences and other sequences

encoding regulatory signals capable of affecting mRNA processing or gene expression. The polyadenylation signal is usually characterized by affecting the addition of polyadenylic acid tracts to the 3' end of the mRNA precursor. The use of different 3' non-coding sequences is exemplified by Ingelbrecht et al. (1989) *Plant Cell* 1:671-680.

5 "RNA transcript" refers to the product resulting from RNA polymerase-catalyzed transcription of a DNA sequence. When the RNA transcript is a perfect complementary copy of the DNA sequence, it is referred to as the primary transcript or it may be a RNA sequence derived from posttranscriptional processing of the primary transcript and is referred to as the mature RNA. "Messenger RNA (mRNA)" refers to the RNA that is
10 without introns and that can be translated into polypeptide by the cell. "cDNA" refers to a double-stranded DNA that is complementary to and derived from mRNA. "Sense" RNA refers to an RNA transcript that includes the mRNA and so can be translated into a polypeptide by the cell. "Antisense RNA" refers to an RNA transcript that is complementary to all or part of a target primary transcript or mRNA and that blocks the
15 expression of a target gene (see U.S. Patent No. 5,107,065, incorporated herein by reference). The complementarity of an antisense RNA may be with any part of the specific nucleotide sequence, i.e., at the 5' non-coding sequence, 3' non-coding sequence, introns, or the coding sequence. "Functional RNA" refers to sense RNA, antisense RNA, ribozyme RNA, or other RNA that may not be translated but yet has an effect on cellular processes.

20 The term "operably linked" refers to the association of two or more nucleic acid fragments on a single nucleic acid fragment so that the function of one is affected by the other. For example, a promoter is operably linked with a coding sequence when it is capable of affecting the expression of that coding sequence (i.e., that the coding sequence is under the transcriptional control of the promoter). Coding sequences can be operably linked to
25 regulatory sequences in sense or antisense orientation.

The term "expression", as used herein, refers to the transcription and stable accumulation of sense (mRNA) or antisense RNA derived from the nucleic acid fragment of the invention. Expression may also refer to translation of mRNA into a polypeptide.

30 "Antisense inhibition" refers to the production of antisense RNA transcripts capable of suppressing the expression of the target protein. "Overexpression" refers to the production of a gene product in transgenic organisms that exceeds levels of production in normal or non-transformed organisms. "Co-suppression" refers to the production of sense RNA transcripts capable of suppressing the expression of identical or substantially similar foreign or endogenous genes (U.S. Patent No. 5,231,020, incorporated herein by reference).

35 "Altered levels" refers to the production of gene product(s) in transgenic organisms in amounts or proportions that differ from that of normal or non-transformed organisms.

"Mature" protein refers to a post-translationally processed polypeptide; i.e., one from which any pre- or propeptides present in the primary translation product have been removed.

“Precursor” protein refers to the primary product of translation of mRNA; i.e., with pre- and propeptides still present. Pre- and propeptides may be but are not limited to intracellular localization signals.

A “chloroplast transit peptide” is an amino acid sequence which is translated in conjunction with a protein and directs the protein to the chloroplast or other plastid types present in the cell in which the protein is made. “Chloroplast transit sequence” refers to a nucleotide sequence that encodes a chloroplast transit peptide. A “signal peptide” is an amino acid sequence which is translated in conjunction with a protein and directs the protein to the secretory system (Chrispeels (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53). If the protein is to be directed to a vacuole, a vacuolar targeting signal (*supra*) can further be added, or if to the endoplasmic reticulum, an endoplasmic reticulum retention signal (*supra*) may be added. If the protein is to be directed to the nucleus, any signal peptide present should be removed and instead a nuclear localization signal included (Raikhel (1992) *Plant Phys.* 100:1627-1632).

“Transformation” refers to the transfer of a nucleic acid fragment into the genome of a host organism, resulting in genetically stable inheritance. Host organisms containing the transformed nucleic acid fragments are referred to as “transgenic” organisms. Examples of methods of plant transformation include *Agrobacterium*-mediated transformation (De Blaere et al. (1987) *Meth. Enzymol.* 143:277) and particle-accelerated or “gene gun” transformation technology (Klein et al. (1987) *Nature (London)* 327:70-73; U.S. Patent No. 4,945,050, incorporated herein by reference).

Standard recombinant DNA and molecular cloning techniques used herein are well known in the art and are described more fully in Sambrook et al. *Molecular Cloning: A Laboratory Manual*; Cold Spring Harbor Laboratory Press: Cold Spring Harbor, 1989 (hereinafter “Maniatis”).

Nucleic acid fragments encoding at least a portion of a cellulose synthase enzyme have been isolated and identified by comparison of random plant cDNA sequences to public databases containing nucleotide and protein sequences using the BLAST algorithms well known to those skilled in the art. The nucleic acid fragments of the instant invention may be used to isolate cDNAs and genes encoding homologous proteins from the same or other plant species. Isolation of homologous genes using sequence-dependent protocols is well known in the art. Examples of sequence-dependent protocols include, but are not limited to, methods of nucleic acid hybridization, and methods of DNA and RNA amplification as exemplified by various uses of nucleic acid amplification technologies (e.g., polymerase chain reaction, ligase chain reaction).

For example, genes encoding other cellulose synthase enzymes, either as cDNAs or genomic DNAs, could be isolated directly by using all or a portion of the instant nucleic acid fragments as DNA hybridization probes to screen libraries from any desired plant employing

methodology well known to those skilled in the art. Specific oligonucleotide probes based upon the instant nucleic acid sequences can be designed and synthesized by methods known in the art (Maniatis). Moreover, the entire sequences can be used directly to synthesize DNA probes by methods known to the skilled artisan such as random primer DNA labeling, nick translation, or end-labeling techniques, or RNA probes using available *in vitro* transcription systems. In addition, specific primers can be designed and used to amplify a part or all of the instant sequences. The resulting amplification products can be labeled directly during amplification reactions or labeled after amplification reactions, and used as probes to isolate full length cDNA or genomic fragments under conditions of appropriate stringency.

In addition, two short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols to amplify longer nucleic acid fragments encoding homologous genes from DNA or RNA. The polymerase chain reaction may also be performed on a library of cloned nucleic acid fragments wherein the sequence of one primer is derived from the instant nucleic acid fragments, and the sequence of the other primer takes advantage of the presence of the polyadenylic acid tracts to the 3' end of the mRNA precursor encoding plant genes. Alternatively, the second primer sequence may be based upon sequences derived from the cloning vector. For example, the skilled artisan can follow the RACE protocol (Frohman et al. (1988) *Proc. Natl. Acad. Sci. USA* 85:8998) to generate cDNAs by using PCR to amplify copies of the region between a single point in the transcript and the 3' or 5' end. Primers oriented in the 3' and 5' directions can be designed from the instant sequences. Using commercially available 3' RACE or 5' RACE systems (BRL), specific 3' or 5' cDNA fragments can be isolated (Ohara et al. (1989) *Proc. Natl. Acad. Sci. USA* 86:5673; Loh et al. (1989) *Science* 243:217). Products generated by the 3' and 5' RACE procedures can be combined to generate full-length cDNAs (Frohman and Martin (1989) *Techniques* 1:165).

Availability of the instant nucleotide and deduced amino acid sequences facilitates immunological screening of cDNA expression libraries. Synthetic peptides representing portions of the instant amino acid sequences may be synthesized. These peptides can be used to immunize animals to produce polyclonal or monoclonal antibodies with specificity for peptides or proteins comprising the amino acid sequences. These antibodies can be then be used to screen cDNA expression libraries to isolate full-length cDNA clones of interest (Lerner (1984) *Adv. Immunol.* 36:1; Maniatis).

The nucleic acid fragments of the instant invention may be used to create transgenic plants in which the disclosed polypeptides are present at higher or lower levels than normal or in cell types or developmental stages in which they are not normally found. This would have the effect of altering the level of cellulose synthase in those cells.

Overexpression of the proteins of the instant invention may be accomplished by first constructing a chimeric gene in which the coding region is operably linked to a promoter capable of directing expression of a gene in the desired tissues at the desired stage of development. For reasons of convenience, the chimeric gene may comprise promoter sequences and translation leader sequences derived from the same genes. 3' Non-coding sequences encoding transcription termination signals may also be provided. The instant chimeric gene may also comprise one or more introns in order to facilitate gene expression.

Plasmid vectors comprising the instant chimeric gene can then be constructed. The choice of plasmid vector is dependent upon the method that will be used to transform host plants. The skilled artisan is well aware of the genetic elements that must be present on the plasmid vector in order to successfully transform, select and propagate host cells containing the chimeric gene. The skilled artisan will also recognize that different independent transformation events will result in different levels and patterns of expression (Jones et al. (1985) *EMBO J.* 4:2411-2418; De Almeida et al. (1989) *Mol. Gen. Genetics* 218:78-86), and thus that multiple events must be screened in order to obtain lines displaying the desired expression level and pattern. Such screening may be accomplished by Southern analysis of DNA, Northern analysis of mRNA expression, Western analysis of protein expression, or phenotypic analysis.

For some applications it may be useful to direct the instant polypeptides to different cellular compartments, or to facilitate its secretion from the cell. It is thus envisioned that the chimeric gene described above may be further supplemented by altering the coding sequence to encode the instant polypeptides with appropriate intracellular targeting sequences such as transit sequences (Keegstra (1989) *Cell* 56:247-253), signal sequences or sequences encoding endoplasmic reticulum localization (Chrispeels (1991) *Ann. Rev. Plant Phys. Plant Mol. Biol.* 42:21-53), or nuclear localization signals (Raikhel (1992) *Plant Phys.* 100:1627-1632) added and/or with targeting sequences that are already present removed. While the references cited give examples of each of these, the list is not exhaustive and more targeting signals of utility may be discovered in the future.

It may also be desirable to reduce or eliminate expression of genes encoding the instant polypeptides in plants for some applications. In order to accomplish this, a chimeric gene designed for co-suppression of the instant polypeptide can be constructed by linking a gene or gene fragment encoding that polypeptide to plant promoter sequences. Alternatively, a chimeric gene designed to express antisense RNA for all or part of the instant nucleic acid fragment can be constructed by linking the gene or gene fragment in reverse orientation to plant promoter sequences. Either the co-suppression or antisense chimeric genes could be introduced into plants via transformation wherein expression of the corresponding endogenous genes are reduced or eliminated.

Molecular genetic solutions to the generation of plants with altered gene expression have a decided advantage over more traditional plant breeding approaches. Changes in plant phenotypes can be produced by specifically inhibiting expression of one or more genes by antisense inhibition or cosuppression (U. S. Patent Nos. 5,190,931, 5,107,065 and 5,283,323). An antisense or cosuppression construct would act as a dominant negative regulator of gene activity. While conventional mutations can yield negative regulation of gene activity these effects are most likely recessive. The dominant negative regulation available with a transgenic approach may be advantageous from a breeding perspective. In addition, the ability to restrict the expression of specific phenotype to the reproductive tissues of the plant by the use of tissue specific promoters may confer agronomic advantages relative to conventional mutations which may have an effect in all tissues in which a mutant gene is ordinarily expressed.

The person skilled in the art will know that special considerations are associated with the use of antisense or cosuppression technologies in order to reduce expression of particular genes. For example, the proper level of expression of sense or antisense genes may require the use of different chimeric genes utilizing different regulatory elements known to the skilled artisan. Once transgenic plants are obtained by one of the methods described above, it will be necessary to screen individual transgenics for those that most effectively display the desired phenotype. Accordingly, the skilled artisan will develop methods for screening large numbers of transformants. The nature of these screens will generally be chosen on practical grounds, and is not an inherent part of the invention. For example, one can screen by looking for changes in gene expression by using antibodies specific for the protein encoded by the gene being suppressed, or one could establish assays that specifically measure enzyme activity. A preferred method will be one which allows large numbers of samples to be processed rapidly, since it will be expected that a large number of transformants will be negative for the desired phenotype.

The instant polypeptides (or portions thereof) may be produced in heterologous host cells, particularly in the cells of microbial hosts, and can be used to prepare antibodies to the these proteins by methods well known to those skilled in the art. The antibodies are useful for detecting the polypeptides of the instant invention *in situ* in cells or *in vitro* in cell extracts. Preferred heterologous host cells for production of the instant polypeptides are microbial hosts. Microbial expression systems and expression vectors containing regulatory sequences that direct high level expression of foreign proteins are well known to those skilled in the art. Any of these could be used to construct a chimeric gene for production of the instant polypeptides. This chimeric gene could then be introduced into appropriate microorganisms via transformation to provide high level expression of the encoded cellulose synthase. An example of a vector for high level expression of the instant polypeptides in a bacterial host is provided (Example 6).

Additionally, the instant polypeptides can be used as a targets to facilitate design and/or identification of inhibitors of those enzymes that may be useful as herbicides. This is desirable because the polypeptides described herein catalyze a step in the synthesis of cellulose. Accordingly, inhibition of the activity of one or more of the enzymes described
5 herein could lead to inhibition plant growth. Thus, the instant polypeptides could be appropriate for new herbicide discovery and design.

All or a substantial portion of the nucleic acid fragments of the instant invention may also be used as probes for genetically and physically mapping the genes that they are a part of, and as markers for traits linked to those genes. Such information may be useful in plant
10 breeding in order to develop lines with desired phenotypes. For example, the instant nucleic acid fragments may be used as restriction fragment length polymorphism (RFLP) markers. Southern blots (Maniatis) of restriction-digested plant genomic DNA may be probed with the nucleic acid fragments of the instant invention. The resulting banding patterns may then be subjected to genetic analyses using computer programs such as MapMaker (Lander et al.
15 (1987) *Genomics* 1:174-181) in order to construct a genetic map. In addition, the nucleic acid fragments of the instant invention may be used to probe Southern blots containing restriction endonuclease-treated genomic DNAs of a set of individuals representing parent and progeny of a defined genetic cross. Segregation of the DNA polymorphisms is noted and used to calculate the position of the instant nucleic acid sequence in the genetic map
20 previously obtained using this population (Botstein et al. (1980) *Am. J. Hum. Genet.* 32:314-331).

The production and use of plant gene-derived probes for use in genetic mapping is described in Bernatzky and Tanksley (1986) *Plant Mol. Biol. Reporter* 4(1):37-41. Numerous publications describe genetic mapping of specific cDNA clones using the
25 methodology outlined above or variations thereof. For example, F2 intercross populations, backcross populations, randomly mated populations, near isogenic lines, and other sets of individuals may be used for mapping. Such methodologies are well known to those skilled in the art.

Nucleic acid probes derived from the instant nucleic acid sequences may also be used
30 for physical mapping (i.e., placement of sequences on physical maps; see Hoheisel et al. In: *Nonmammalian Genomic Analysis: A Practical Guide*, Academic press 1996, pp. 319-346, and references cited therein).

In another embodiment, nucleic acid probes derived from the instant nucleic acid sequences may be used in direct fluorescence *in situ* hybridization (FISH) mapping (Trask
35 (1991) *Trends Genet.* 7:149-154). Although current methods of FISH mapping favor use of large clones (several to several hundred KB; see Laan et al. (1995) *Genome Research* 5:13-20), improvements in sensitivity may allow performance of FISH mapping using shorter probes.

A variety of nucleic acid amplification-based methods of genetic and physical mapping may be carried out using the instant nucleic acid sequences. Examples include allele-specific amplification (Kazazian (1989) *J. Lab. Clin. Med.* 114(2):95-96), polymorphism of PCR-amplified fragments (CAPS; Sheffield et al. (1993) *Genomics* 16:325-332), allele-specific ligation (Landegren et al. (1988) *Science* 241:1077-1080), nucleotide extension reactions (Sokolov (1990) *Nucleic Acid Res.* 18:3671), Radiation Hybrid Mapping (Walter et al. (1997) *Nature Genetics* 7:22-28) and Happy Mapping (Dear and Cook (1989) *Nucleic Acid Res.* 17:6795-6807). For these methods, the sequence of a nucleic acid fragment is used to design and produce primer pairs for use in the amplification reaction or in primer extension reactions. The design of such primers is well known to those skilled in the art. In methods employing PCR-based genetic mapping, it may be necessary to identify DNA sequence differences between the parents of the mapping cross in the region corresponding to the instant nucleic acid sequence. This, however, is generally not necessary for mapping methods.

Loss of function mutant phenotypes may be identified for the instant cDNA clones either by targeted gene disruption protocols or by identifying specific mutants for these genes contained in a maize population carrying mutations in all possible genes (Ballinger and Benzer (1989) *Proc. Natl. Acad. Sci USA* 86:9402; Koes et al. (1995) *Proc. Natl. Acad. Sci USA* 92:8149; Bensen et al. (1995) *Plant Cell* 7:75). The latter approach may be accomplished in two ways. First, short segments of the instant nucleic acid fragments may be used in polymerase chain reaction protocols in conjunction with a mutation tag sequence primer on DNAs prepared from a population of plants in which Mutator transposons or some other mutation-causing DNA element has been introduced (see Bensen, *supra*). The amplification of a specific DNA fragment with these primers indicates the insertion of the mutation tag element in or near the plant gene encoding the instant polypeptides. Alternatively, the instant nucleic acid fragment may be used as a hybridization probe against PCR amplification products generated from the mutation population using the mutation tag sequence primer in conjunction with an arbitrary genomic site primer, such as that for a restriction enzyme site-anchored synthetic adaptor. With either method, a plant containing a mutation in the endogenous gene encoding the instant polypeptides can be identified and obtained. This mutant plant can then be used to determine or confirm the natural function of the instant polypeptides disclosed herein.

EXAMPLES

The present invention is further defined in the following Examples, in which all parts and percentages are by weight and degrees are Celsius, unless otherwise stated. It should be understood that these Examples, while indicating preferred embodiments of the invention, are given by way of illustration only. From the above discussion and these Examples, one skilled in the art can ascertain the essential characteristics of this invention, and without

departing from the spirit and scope thereof, can make various changes and modifications of the invention to adapt it to various usages and conditions.

EXAMPLE 1

Composition of cDNA Libraries: Isolation and Sequencing of cDNA Clones

- 5 cDNA libraries representing mRNAs from various barley, corn, rice, soybean and wheat tissues were prepared. The characteristics of the libraries are described below.

TABLE 2

cDNA Libraries from Barley, Corn, Rice, Soybean and Wheat

Library	Tissue	Clone
bsh1	Barley (<i>Hordeum vulgare</i>) sheath, developing seedling	bsh1.pk0002.f6
cco1n	Corn (<i>Zea mays</i>) cob of 67 day old plants grown in green house*	cco1n.pk0005.g3
cdt2c	Corn (<i>Zea mays</i>) developing tassel 2	cdt2c.pk002.g1 cdt2c.pk002.i16
cr1n	Corn (<i>Zea mays</i>) root from 7 day seedlings grown in light*	cr1n.pk0135.e10
csc1c	Corn (<i>Zea mays</i>) 20 day seedling (germination under cold stress)	csc1c.pk002.i1
p0031	Corn (<i>Zea mays</i>) shoot culture, initiated from seed derived meristems culture was maintained on 273N medium.	p0031.ccmr05rb
p0110	Corn (<i>Zea mays</i>) stages V3/V4** leaf tissue minus midrib harvested 4 hours, 24 hours and 7 days after infiltration with salicylic acid, tissues pooled*	p0110.cgsma57r
p0097	Corn (<i>Zea mays</i>) stage V9** whorl section (7 cm) from plant infected four times with european corn borer	p0097.cqrad17rc
p0122	Corn (<i>Zea mays</i>) pith tissue collected from internode subtending ear node 5 days after pollination	p0122.ckamh70rc
rlr24	Rice (<i>Oryza sativa</i>) leaf (15 days after germination) 24 hours after infection of <i>Magaporthe grisea</i> strain 4360-R-62 (AVR2-YAMO); Resistant	rlr24.pk0073.g1
sdp2c	Soybean (<i>Glycine max</i>) developing pods 6-7 mm	sdp2c.pk005.o22
ses8w	Soybean (<i>Glycine max</i>) mature embryo 8 weeks after subculture	ses8w.pk0028.f3
ssl	Soybean (<i>Glycine max</i>) seedling 5-10 day	ssl.pk0036.c10
wl1	Wheat (<i>Triticum aestivum</i>) leaf 7 day old seedling, light grown	wl1.pk0009.c9
wl1n	Wheat (<i>Triticum aestivum</i>) leaf 7 day old seedling, light grown*	wl1n.pk0044.b1
wr1	Wheat (<i>Triticum aestivum</i>) root; 7 day old seedling, light grown	wr1.pk0160.d11

Library	Tissue	Clone
wre1n	Wheat (<i>Triticum aestivum</i>) root: 7 day old etiolated seedling*	wre1n.pk0043.f9
		wre1n.pk0043.h8
		wre1n.pk0131.g10

*These libraries were normalized essentially as described in U.S. Patent No. 5,482,845, incorporated herein by reference.

5 **V3, V4 and V9 refer to stages of corn growth. The descriptions can be found in "How a Corn Plant Develops" Special Report No. 48, Iowa State University of Science and Technology Cooperative Extension Service Ames, Iowa, Reprinted February 1993.

cDNA libraries may be prepared by any one of many methods available. For example, the cDNAs may be introduced into plasmid vectors by first preparing the cDNA
10 libraries in Uni-ZAP™ XR vectors according to the manufacturer's protocol (Stratagene Cloning Systems, La Jolla, CA). The Uni-ZAP™ XR libraries are converted into plasmid libraries according to the protocol provided by Stratagene. Upon conversion, cDNA inserts will be contained in the plasmid vector pBluescript. In addition, the cDNAs may be introduced directly into precut Bluescript II SK(+) vectors (Stratagene) using T4 DNA
15 ligase (New England Biolabs), followed by transfection into DH10B cells according to the manufacturer's protocol (GIBCO BRL Products). Once the cDNA inserts are in plasmid vectors, plasmid DNAs are prepared from randomly picked bacterial colonies containing recombinant pBluescript plasmids, or the insert cDNA sequences are amplified via polymerase chain reaction using primers specific for vector sequences flanking the inserted
20 cDNA sequences. Amplified insert DNAs or plasmid DNAs are sequenced in dye-primer sequencing reactions to generate partial cDNA sequences (expressed sequence tags or "ESTs"; see Adams et al., (1991) *Science* 252:1651). The resulting ESTs are analyzed using a Perkin Elmer Model 377 fluorescent sequencer.

EXAMPLE 2

25 Identification of cDNA Clones

cDNA clones encoding cellulose synthase enzymes were identified by conducting BLAST (Basic Local Alignment Search Tool; Altschul et al. (1993) *J. Mol. Biol.* 215:403-410; see also www.ncbi.nlm.nih.gov/BLAST/) searches for similarity to sequences contained in the BLAST "nr" database (comprising all non-redundant GenBank CDS
30 translations, sequences derived from the 3-dimensional structure Brookhaven Protein Data Bank, the last major release of the SWISS-PROT protein sequence database, EMBL, and DDBJ databases). The cDNA sequences obtained in Example 1 were analyzed for similarity to all publicly available DNA sequences contained in the "nr" database using the BLASTN algorithm provided by the National Center for Biotechnology Information (NCBI). The

DNA sequences were translated in all reading frames and compared for similarity to all publicly available protein sequences contained in the "nr" database using the BLASTX algorithm (Gish and States (1993) *Nature Genetics* 3:266-272) provided by the NCBI. For convenience, the P-value (probability) of observing a match of a cDNA sequence to a sequence contained in the searched databases merely by chance as calculated by BLAST are reported herein as "pLog" values, which represent the negative of the logarithm of the reported P-value. Accordingly, the greater the pLog value, the greater the likelihood that the cDNA sequence and the BLAST "hit" represent homologous proteins.

EXAMPLE 3

Characterization of cDNA Clones Encoding Cellulose Synthase

The BLASTX search using the EST sequences from clones listed in Table 3 revealed similarity of the polypeptides encoded by the cDNAs to cellulose synthase from *Arabidopsis thaliana* (NCBI Identifier No. gi 2827139, gi 2827141, gi 4467125, gi 4886756 and gi 3135611) and *Gossypium hirsutum* (NCBI Identifier No. gi 1706958 and 5081779). Shown in Table 3 are the BLAST results for individual ESTs ("EST"), the sequences of the entire cDNA inserts comprising the indicated cDNA clones ("FIS"), complete gene sequences ("CGS") or contigs assembled from two or more ESTs ("Contig"):

TABLE 3

BLAST Results for Sequences Encoding Polypeptides Homologous to *Arabidopsis thaliana* and *Gossypium hirsutum* Cellulose Synthase

Clone	Status	BLAST pLog Score
bsh1.pk0002.f6	FIS	154.00 (gi 2827139)
Contig composed of: ccoln.pk0005.g3 cdt2c.pk002.g1 cdt2c.pk002.l16 csc1c.pk002.i1 p0031.ccmr05rb p0110.cgsm57r	Contig	>254.00 (gi 2827141)
cr1n.pk0135.e10	FIS	176.00 (gi 1706958)
p0097.cqrad17rc	CGS	>254.00 (gi 2827141)
p0122.ckamh70rc	CGS	>254.00 (gi 2827141)
rlr24.pk0073.g1	EST	77.70 (gi 4467125)
sdp2c.pk005.o22	FIS	>254.00 (gi 4886756)
ses8w.pk0028.f3	EST	>254.00 (gi 2827139)
ssl.pk0036.c10	EST	>254.00 (gi 2827141)
Contig composed of: wl1.pk0009.c9 wr1.pk0160.d11 wre1n.pk0043.f9	Contig	>254.00 (gi 5081779)

wreln.pk0043.h8		
wreln.pk0131.g10		
wl1n.pk0044.b1	EST	166.00 (gi 3135611)

Figure 1 presents an alignment of the amino acid sequences set forth in SEQ ID NOs:2, 4, 8, 10, 12, 14, 16, 18, 20 and 22 and the *Arabidopsis thaliana* (SEQ ID NOs:23 (gi 2827139), 24 (gi 2827141), 26 (gi 4467125), 27 (gi 4886756) and 29 (gi 3135611)) and *Gossypium hirsutum* (SEQ ID NOs:25 (gi 1706958) and 28 (gi 5081779)) sequences. The data in Table 4 represents a calculation of the percent identity of the amino acid sequences set forth in SEQ ID NOs:2, 4, 8, 10, 12, 14, 16, 18, 20 and 22 and the *Arabidopsis thaliana* (SEQ ID NOs:23, 24, 26, 27 and 29) and *Gossypium hirsutum* (SEQ ID NOs:25 and 28) sequences.

TABLE 4

Percent Identity of Amino Acid Sequences Deduced From the Nucleotide Sequences of cDNA Clones Encoding Polypeptides Homologous to *Arabidopsis thaliana* and *Gossypium hirsutum* Cellulose Synthase

SEQ ID NO.	Percent Identity to
2	82% (gi 2827139)
4	69% (gi 2827141)
6	89% (gi 1706958)
8	70% (gi 2827141)
10	70% (gi 2827141)
12	36% (gi 4467125)
14	86% (gi 4886756)
16	88% (gi 2827139)
18	86% (gi 2827141)
20	87% (gi 5081779)
22	70% (gi 3135611)

Sequence alignments and percent identity calculations were performed using the Megalign program of the LASARGENE bioinformatics computing suite (DNASTAR Inc., Madison, WI). Multiple alignment of the sequences was performed using the Clustal method of alignment (Higgins and Sharp (1989) *CABIOS*. 5:151-153) with the default parameters (GAP PENALTY=10, GAP LENGTH PENALTY=10). Default parameters for pairwise alignments using the Clustal method were KTUPLE 1, GAP PENALTY=3, WINDOW=5 and DIAGONALS SAVED=5. Sequence alignments and BLAST scores and probabilities indicate that the nucleic acid fragments comprising the instant cDNA clones

encode a substantial portion of a cellulose synthase. These sequences represent the first barley, corn, rice, soybean and wheat sequences encoding cellulose synthase.

EXAMPLE 4

Expression of Chimeric Genes in Monocot Cells

5 A chimeric gene comprising a cDNA encoding the instant polypeptides in sense orientation with respect to the maize 27 kD zein promoter that is located 5' to the cDNA fragment, and the 10 kD zein 3' end that is located 3' to the cDNA fragment, can be constructed. The cDNA fragment of this gene may be generated by polymerase chain reaction (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites
10 (NcoI or SmaI) can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the digested vector pML103 as described below. Amplification is then performed in a standard PCR. The amplified DNA is then digested with restriction enzymes NcoI and SmaI and fractionated on an agarose gel. The appropriate band can be isolated from the gel and combined with a 4.9 kb NcoI-SmaI fragment of the
15 plasmid pML103. Plasmid pML103 has been deposited under the terms of the Budapest Treaty at ATCC (American Type Culture Collection, 10801 University Blvd., Manassas, VA 20110-2209), and bears accession number ATCC 97366. The DNA segment from pML103 contains a 1.05 kb SalI-NcoI promoter fragment of the maize 27 kD zein gene and a 0.96 kb SmaI-SalI fragment from the 3' end of the maize 10 kD zein gene in the vector
20 pGem9Zf(+) (Promega). Vector and insert DNA can be ligated at 15°C overnight, essentially as described (Maniatis). The ligated DNA may then be used to transform *E. coli* XL1-Blue (Epicurian Coli XL-1 Blue™; Stratagene). Bacterial transformants can be screened by restriction enzyme digestion of plasmid DNA and limited nucleotide sequence analysis using the dideoxy chain termination method (Sequenase™ DNA Sequencing Kit:
25 U.S. Biochemical). The resulting plasmid construct would comprise a chimeric gene encoding, in the 5' to 3' direction, the maize 27 kD zein promoter, a cDNA fragment encoding the instant polypeptides, and the 10 kD zein 3' region.

The chimeric gene described above can then be introduced into corn cells by the following procedure. Immature corn embryos can be dissected from developing caryopses
30 derived from crosses of the inbred corn lines H99 and LH132. The embryos are isolated 10 to 11 days after pollination when they are 1.0 to 1.5 mm long. The embryos are then placed with the axis-side facing down and in contact with agarose-solidified N6 medium (Chu et al. (1975) *Sci. Sin. Peking* 18:659-668). The embryos are kept in the dark at 27°C. Friable embryogenic callus consisting of undifferentiated masses of cells with somatic
35 proembryoids and embryoids borne on suspensor structures proliferates from the scutellum of these immature embryos. The embryogenic callus isolated from the primary explant can be cultured on N6 medium and sub-cultured on this medium every 2 to 3 weeks.

The plasmid, p35S/Ac (obtained from Dr. Peter Eckes, Hoechst Ag, Frankfurt, Germany) may be used in transformation experiments in order to provide for a selectable marker. This plasmid contains the *Pat* gene (see European Patent Publication 0 242 236) which encodes phosphinothricin acetyl transferase (PAT). The enzyme PAT confers

5 resistance to herbicidal glutamine synthetase inhibitors such as phosphinothricin. The *pat* gene in p35S/Ac is under the control of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*.

The particle bombardment method (Klein et al. (1987) *Nature* 327:70-73) may be used

10 to transfer genes to the callus culture cells. According to this method, gold particles (1 μ m in diameter) are coated with DNA using the following technique. Ten μ g of plasmid DNAs are added to 50 μ L of a suspension of gold particles (60 mg per mL). Calcium chloride (50 μ L of a 2.5 M solution) and spermidine free base (20 μ L of a 1.0 M solution) are added to the particles. The suspension is vortexed during the addition of these solutions. After

15 10 minutes, the tubes are briefly centrifuged (5 sec at 15,000 rpm) and the supernatant removed. The particles are resuspended in 200 μ L of absolute ethanol, centrifuged again and the supernatant removed. The ethanol rinse is performed again and the particles resuspended in a final volume of 30 μ L of ethanol. An aliquot (5 μ L) of the DNA-coated gold particles can be placed in the center of a Kapton™ flying disc (Bio-Rad Labs). The

20 particles are then accelerated into the corn tissue with a Biolistic™ PDS-1000/He (Bio-Rad Instruments, Hercules CA), using a helium pressure of 1000 psi, a gap distance of 0.5 cm and a flying distance of 1.0 cm.

For bombardment, the embryogenic tissue is placed on filter paper over agarose-solidified N6 medium. The tissue is arranged as a thin lawn and covered a circular area of

25 about 5 cm in diameter. The petri dish containing the tissue can be placed in the chamber of the PDS-1000/He approximately 8 cm from the stopping screen. The air in the chamber is then evacuated to a vacuum of 28 inches of Hg. The macrocarrier is accelerated with a helium shock wave using a rupture membrane that bursts when the He pressure in the shock tube reaches 1000 psi.

30 Seven days after bombardment the tissue can be transferred to N6 medium that contains glufosinate (2 mg per liter) and lacks casein or proline. The tissue continues to grow slowly on this medium. After an additional 2 weeks the tissue can be transferred to fresh N6 medium containing glufosinate. After 6 weeks, areas of about 1 cm in diameter of actively growing callus can be identified on some of the plates containing the glufosinate-supplemented medium. These calli may continue to grow when sub-cultured on the

35 selective medium.

Plants can be regenerated from the transgenic callus by first transferring clusters of tissue to N6 medium supplemented with 0.2 mg per liter of 2,4-D. After two weeks the

tissue can be transferred to regeneration medium (Fromm et al. (1990) *Bio/Technology* 8:833-839).

EXAMPLE 5

Expression of Chimeric Genes in Dicot Cells

5 A seed-specific expression cassette composed of the promoter and transcription terminator from the gene encoding the β subunit of the seed storage protein phaseolin from the bean *Phaseolus vulgaris* (Doyle et al. (1986) *J. Biol. Chem.* 261:9228-9238) can be used for expression of the instant polypeptides in transformed soybean. The phaseolin cassette includes about 500 nucleotides upstream (5') from the translation initiation codon and about 10 1650 nucleotides downstream (3') from the translation stop codon of phaseolin. Between the 5' and 3' regions are the unique restriction endonuclease sites Nco I (which includes the ATG translation initiation codon), Sma I, Kpn I and Xba I. The entire cassette is flanked by Hind III sites.

The cDNA fragment of this gene may be generated by polymerase chain reaction 15 (PCR) of the cDNA clone using appropriate oligonucleotide primers. Cloning sites can be incorporated into the oligonucleotides to provide proper orientation of the DNA fragment when inserted into the expression vector. Amplification is then performed as described above, and the isolated fragment is inserted into a pUC18 vector carrying the seed expression cassette.

20 Soybean embryos may then be transformed with the expression vector comprising sequences encoding the instant polypeptides. To induce somatic embryos, cotyledons, 3-5 mm in length dissected from surface sterilized, immature seeds of the soybean cultivar A2872, can be cultured in the light or dark at 26°C on an appropriate agar medium for 6-10 weeks. Somatic embryos which produce secondary embryos are then excised and 25 placed into a suitable liquid medium. After repeated selection for clusters of somatic embryos which multiplied as early, globular staged embryos, the suspensions are maintained as described below.

Soybean embryogenic suspension cultures can maintained in 35 mL liquid media on a rotary shaker, 150 rpm, at 26°C with florescent lights on a 16:8 hour day/night schedule.

30 Cultures are subcultured every two weeks by inoculating approximately 35 mg of tissue into 35 mL of liquid medium.

Soybean embryogenic suspension cultures may then be transformed by the method of particle gun bombardment (Klein et al. (1987) *Nature* (London) 327:70, U.S. Patent No. 4,945,050). A DuPont Biolistic™ PDS1000/HE instrument (helium retrofit) can be used 35 for these transformations.

A selectable marker gene which can be used to facilitate soybean transformation is a chimeric gene composed of the 35S promoter from Cauliflower Mosaic Virus (Odell et al. (1985) *Nature* 313:810-812), the hygromycin phosphotransferase gene from plasmid pJR225

(from *E. coli*: Gritz et al.(1983) *Gene* 25:179-188) and the 3' region of the nopaline synthase gene from the T-DNA of the Ti plasmid of *Agrobacterium tumefaciens*. The seed expression cassette comprising the phaseolin 5' region, the fragment encoding the instant polypeptides and the phaseolin 3' region can be isolated as a restriction fragment. This fragment can then

To 50 μ L of a 60 mg/mL 1 μ m gold particle suspension is added (in order): 5 μ L DNA (1 μ g/ μ L), 20 μ L spermidine (0.1 M), and 50 μ L CaCl_2 (2.5 M). The particle preparation is then agitated for three minutes, spun in a microfuge for 10 seconds and the supernatant removed. The DNA-coated particles are then washed once in 400 μ L 70% ethanol and resuspended in 40 μ L of anhydrous ethanol. The DNA/particle suspension can be sonicated three times for one second each. Five μ L of the DNA-coated gold particles are then loaded on each macro carrier disk.

Approximately 300-400 mg of a two-week-old suspension culture is placed in an empty 60x15 mm petri dish and the residual liquid removed from the tissue with a pipette. For each transformation experiment, approximately 5-10 plates of tissue are normally bombarded. Membrane rupture pressure is set at 1100 psi and the chamber is evacuated to a vacuum of 28 inches mercury. The tissue is placed approximately 3.5 inches away from the retaining screen and bombarded three times. Following bombardment, the tissue can be divided in half and placed back into liquid and cultured as described above.

Five to seven days post bombardment, the liquid media may be exchanged with fresh media, and eleven to twelve days post bombardment with fresh media containing 50 mg/mL hygromycin. This selective media can be refreshed weekly. Seven to eight weeks post bombardment, green, transformed tissue may be observed growing from untransformed, necrotic embryogenic clusters. Isolated green tissue is removed and inoculated into individual flasks to generate new, clonally propagated, transformed embryogenic suspension cultures. Each new line may be treated as an independent transformation event. These suspensions can then be subcultured and maintained as clusters of immature embryos or regenerated into whole plants by maturation and germination of individual somatic embryos.

EXAMPLE 6

Expression of Chimeric Genes in Microbial Cells

The cDNAs encoding the instant polypeptides can be inserted into the T7 *E. coli* expression vector pBT430. This vector is a derivative of pET-3a (Rosenberg et al. (1987) *Gene* 56:125-135) which employs the bacteriophage T7 RNA polymerase/T7 promoter system. Plasmid pBT430 was constructed by first destroying the EcoR I and Hind III sites in pET-3a at their original positions. An oligonucleotide adaptor containing EcoR I and Hind III sites was inserted at the BamH I site of pET-3a. This created pET-3aM with additional unique cloning sites for insertion of genes into the expression vector. Then, the Nde I site at the position of translation initiation was converted to an Nco I site using

oligonucleotide-directed mutagenesis. The DNA sequence of pET-3aM in this region, 5'-CATATGG, was converted to 5'-CCCATGG in pBT430.

Plasmid DNA containing a cDNA may be appropriately digested to release a nucleic acid fragment encoding the protein. This fragment may then be purified on a 1% NuSieve GTG™ low melting agarose gel (FMC). Buffer and agarose contain 10 µg/ml ethidium bromide for visualization of the DNA fragment. The fragment can then be purified from the agarose gel by digestion with GELase™ (Epicentre Technologies) according to the manufacturer's instructions, ethanol precipitated, dried and resuspended in 20 µL of water. Appropriate oligonucleotide adapters may be ligated to the fragment using T4 DNA ligase (New England Biolabs, Beverly, MA). The fragment containing the ligated adapters can be purified from the excess adapters using low melting agarose as described above. The vector pBT430 is digested, dephosphorylated with alkaline phosphatase (NEB) and deproteinized with phenol/chloroform as described above. The prepared vector pBT430 and fragment can then be ligated at 16°C for 15 hours followed by transformation into DH5 electrocompetent cells (GIBCO BRL). Transformants can be selected on agar plates containing LB media and 100 µg/mL ampicillin. Transformants containing the gene encoding the instant polypeptides are then screened for the correct orientation with respect to the T7 promoter by restriction enzyme analysis.

For high level expression, a plasmid clone with the cDNA insert in the correct orientation relative to the T7 promoter can be transformed into *E. coli* strain BL21(DE3) (Studier et al. (1986) *J. Mol. Biol.* 189:113-130). Cultures are grown in LB medium containing ampicillin (100 mg/L) at 25°C. At an optical density at 600 nm of approximately 1, IPTG (isopropylthio-β-galactoside, the inducer) can be added to a final concentration of 0.4 mM and incubation can be continued for 3 h at 25°. Cells are then harvested by centrifugation and re-suspended in 50 µL of 50 mM Tris-HCl at pH 8.0 containing 0.1 mM DTT and 0.2 mM phenyl methylsulfonyl fluoride. A small amount of 1 mm glass beads can be added and the mixture sonicated 3 times for about 5 seconds each time with a microprobe sonicator. The mixture is centrifuged and the protein concentration of the supernatant determined. One µg of protein from the soluble fraction of the culture can be separated by SDS-polyacrylamide gel electrophoresis. Gels can be observed for protein bands migrating at the expected molecular weight.

EXAMPLE 7

Evaluating Compounds for Their Ability to Inhibit the Activity of Cellulose Synthase

The polypeptides described herein may be produced using any number of methods known to those skilled in the art. Such methods include, but are not limited to, expression in bacteria as described in Example 6, or expression in eukaryotic cell culture, *in planta*, and using viral expression systems in suitably infected organisms or cell lines. The instant

polypeptides may be expressed either as mature forms of the proteins as observed *in vivo* or as fusion proteins by covalent attachment to a variety of enzymes, proteins or affinity tags. Common fusion protein partners include glutathione S-transferase ("GST"), thioredoxin ("Trx"), maltose binding protein, and C- and/or N-terminal hexahistidine polypeptide ("His)₆"). The fusion proteins may be engineered with a protease recognition site at the fusion point so that fusion partners can be separated by protease digestion to yield intact mature enzyme. Examples of such proteases include thrombin, enterokinase and factor Xa. However, any protease can be used which specifically cleaves the peptide connecting the fusion protein and the enzyme.

Purification of the instant polypeptides, if desired, may utilize any number of separation technologies familiar to those skilled in the art of protein purification. Examples of such methods include, but are not limited to, homogenization, filtration, centrifugation, heat denaturation, ammonium sulfate precipitation, desalting, pH precipitation, ion exchange chromatography, hydrophobic interaction chromatography and affinity chromatography, wherein the affinity ligand represents a substrate, substrate analog or inhibitor. When the instant polypeptides are expressed as fusion proteins, the purification protocol may include the use of an affinity resin which is specific for the fusion protein tag attached to the expressed enzyme or an affinity resin containing ligands which are specific for the enzyme. For example, the instant polypeptides may be expressed as a fusion protein coupled to the C-terminus of thioredoxin. In addition, a (His)₆ peptide may be engineered into the N-terminus of the fused thioredoxin moiety to afford additional opportunities for affinity purification. Other suitable affinity resins could be synthesized by linking the appropriate ligands to any suitable resin such as Sepharose-4B. In an alternate embodiment, a thioredoxin fusion protein may be eluted using dithiothreitol; however, elution may be accomplished using other reagents which interact to displace the thioredoxin from the resin. These reagents include β -mercaptoethanol or other reduced thiol. The eluted fusion protein may be subjected to further purification by traditional means as stated above, if desired. Proteolytic cleavage of the thioredoxin fusion protein and the enzyme may be accomplished after the fusion protein is purified or while the protein is still bound to the ThioBond™ affinity resin or other resin.

Crude, partially purified or purified enzyme, either alone or as a fusion protein, may be utilized in assays for the evaluation of compounds for their ability to inhibit enzymatic activation of the instant polypeptides disclosed herein. Assays may be conducted under well known experimental conditions which permit optimal enzymatic activity. For example, assays for cellulose synthase activity are presented in WO 98/18949 and WO 98/00549.

CLAIMS

What is claimed is:

1. An isolated nucleic acid fragment comprising at least 900 nucleotides, wherein the nucleic acid fragment encodes a cellulose synthase comprising a member selected from the group consisting of:
 - (a) an isolated nucleic acid fragment encoding an amino acid sequence that is at least 90% identical to the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2, 6, 12, 14, 16, 18, 20 and 22;
 - (b) an isolated nucleic acid fragment that is complementary to (a).
2. The isolated nucleic acid fragment of Claim 1 wherein nucleic acid fragment is a functional RNA.
3. The isolated nucleic acid fragment of Claim 1 wherein the nucleotide sequence of the fragment comprises the sequence set forth in a member selected from the group consisting of SEQ ID NO:1, 5, 11, 13, 15, 17, 19 and 21.
4. A chimeric gene comprising the nucleic acid fragment of Claim 1 operably linked to suitable regulatory sequences.
5. A transformed host cell comprising the chimeric gene of Claim 4.
6. A cellulose synthase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:2, 6, 12, 14, 16, 18, 20 and 22.
7. An isolated nucleic acid fragment encoding a cellulose synthase comprising a member selected from the group consisting of:
 - (a) an isolated nucleic acid fragment encoding an amino acid sequence that is functionally active polypeptide and at least 80% identical to the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:4, 8 and 10 ;
 - (b) an isolated nucleic acid fragment that is complementary to (a).
8. The isolated nucleic acid fragment of Claim 7 wherein nucleic acid fragment is a functional RNA.
9. The isolated nucleic acid fragment of Claim 7 wherein the nucleotide sequence of the fragment comprises the sequence set forth in a member selected from the group consisting of SEQ ID NO:3, 7 and 9.
10. A chimeric gene comprising the nucleic acid fragment of Claim 7 operably linked to suitable regulatory sequences.
11. A transformed host cell comprising the chimeric gene of Claim 10.

12. A cellulose synthase polypeptide comprising all or a substantial portion of the amino acid sequence set forth in a member selected from the group consisting of SEQ ID NO:4, 8, 10.

13. A method of altering the level of expression of a cellulose synthase in a host cell comprising:

- (a) transforming a host cell with the chimeric gene of any of Claims 4 and 10; and
- (b) growing the transformed host cell produced in step (a) under conditions that are suitable for expression of the chimeric gene

wherein expression of the chimeric gene results in production of altered levels of a cellulose synthase in the transformed host cell.

14. A method of obtaining a nucleic acid fragment encoding all or a substantial portion of the amino acid sequence encoding a cellulose synthase comprising:

- (a) probing a cDNA or genomic library with the nucleic acid fragment of any of Claims 1 and 7;
- (b) identifying a DNA clone that hybridizes with the nucleic acid fragment any of of Claims 1 and 7;
- (c) isolating the DNA clone identified in step (b); and
- (d) sequencing the cDNA or genomic fragment that comprises the clone isolated in step (c)

wherein the sequenced nucleic acid fragment encodes all or a substantial portion of the amino acid sequence encoding a cellulose synthase.

15. A method of obtaining a nucleic acid fragment encoding a substantial portion of an amino acid sequence encoding a cellulose synthase comprising:

- (a) synthesizing an oligonucleotide primer corresponding to a portion of the sequence set forth in any of SEQ ID NOs:1, 3, 5, 7, 9, 11, 13, 15, 17, 19 and 21; and
- (b) amplifying a cDNA insert present in a cloning vector using the oligonucleotide primer of step (a) and a primer representing sequences of the cloning vector

wherein the amplified nucleic acid fragment encodes a substantial portion of an amino acid sequence encoding a cellulose synthase.

16. The product of the method of Claim 14.

17. The product of the method of Claim 15.

18. A method for evaluating at least one compound for its ability to inhibit the activity of a cellulose synthase, the method comprising the steps of:

- 5
- (a) transforming a host cell with a chimeric gene comprising a nucleic acid fragment encoding a cellulose synthase, operably linked to suitable regulatory sequences;
- (b) growing the transformed host cell under conditions that are suitable for expression of the chimeric gene wherein expression of the chimeric gene results in production of the cellulose synthase encoded by the operably linked nucleic acid fragment in the transformed host cell;
- (c) optionally purifying the cellulose synthase expressed by the transformed host cell;
- 10 (d) treating the cellulose synthase with a compound to be tested; and
- (e) comparing the activity of the cellulose synthase that has been treated with a test compound to the activity of an untreated cellulose synthase, thereby selecting compounds with potential for inhibitory activity.

Figure 1

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1
60
SEQ ID NO:2 -----
SEQ ID NO:4 RAAQAQRNKGKQPQEEQKLASVSLP--LPHSRFIPFPFRRRYRRR---RTHACPG----I
SEQ ID NO:6 -----
SEQ ID NO:8 HSSYTKSRSSLAQPRAAPRQAQPPP--ATAACACERSPRPGDQRRGGLRAFRCAAAAGFV
SEQ ID NO:10 -----
SEQ ID NO:12 RCS---RRWTCSSPPPTPTRSRRSPR RTP-----
SEQ ID NO:14 -----
SEQ ID NO:16 -----
SEQ ID NO:18 -----
SEQ ID NO:20 -----
SEQ ID NO:22 -----
SEQ ID NO:23 -----
SEQ ID NO:24 MNTGGR-----
SEQ ID NO:25 -----
SEQ ID NO:26 MASTPPQTSKKVRNNSGSGQTVKFARRTSSGRYVSLR-RDNIELSGELSGDYSNYTVHIP
SEQ ID NO:27 -----
SEQ ID NO:28 -----
SEQ ID NO:29 R---PR-----

61
120
SEQ ID NO:2 -----
SEQ ID NO:4 W-RSGSARG---ME-ASAGLVAGSHNRNELV-VIRRDGEPGPKP--MDQRNGQVCQI--
SEQ ID NO:6 -----
SEQ ID NO:8 RERDPAGRGGGPEME-ASAGLVAGSHNRNELV-VIRRDRESGAAGGGAARRAEAPCQI--
SEQ ID NO:10 -----ME-ASAGLVAGSHNRNELV-VIRRDGDPGPKP--PREQNGQVCQI--
SEQ ID NO:12 -----C-----
SEQ ID NO:14 -----ME-ASAGLVAGSHNRNELV-VIHGHEEP--KA--LKNLDGQVCEI--
SEQ ID NO:16 -----
SEQ ID NO:18 -----
SEQ ID NO:20 -----
SEQ ID NO:22 -----
SEQ ID NO:23 -----ME-ASAGLVAGSYRRNELV-RIRHESDGGTKP--LKNMNGQICQI--
SEQ ID NO:24 -----LIAGSHNRNEFV-LI--NADESARIRSVQELSGQTCQI--
SEQ ID NO:25 -----
SEQ ID NO:26 PTPDNQPMATKAEEQYVNSLFTGGFNSVTRAHLMDKVIDSDVTHPQAGAKGSSCAMP
SEQ ID NO:27 -----ME-ASAGLVAGSHNRNELV-VIHGHEEP--KP--LKNLDGQVCEI--
SEQ ID NO:28 -----
SEQ ID NO:29 -----LIAGSHNRNEFV-LI--NADENARIRSVQELSGQTCQI--

121
180
SEQ ID NO:2 -----
SEQ ID NO:4 CGDDVGRNPDGEPFVACNECAFPICRDCYERREGTQNCPOCKTRFKRLKGCARVPGD-
SEQ ID NO:6 -----
SEQ ID NO:8 CGDEVGVGFDGEPFVACNECAFPVCRACYERREGSQACPQCRTRYKRLKGCPRVAGD-
SEQ ID NO:10 CGDDVGLAPGGDPFVACNECAFPVCRDCYERREGTQNCPOCKTRYKRLKGCQRTVTD-
SEQ ID NO:12 -----CPY-----
SEQ ID NO:14 CGDGVGLTVDGLFVACNECGFPVCRPCYERREGSHLCPQCKTRYKRLKGSPRVEGDD
SEQ ID NO:16 -----
SEQ ID NO:18 -----
SEQ ID NO:20 -----
SEQ ID NO:22 -----
SEQ ID NO:23 CGDDVGLAETGDFVACNECAFPVCRPCYERKDGTOCCPQCKTRFRHRGSPRVEGDE
SEQ ID NO:24 CGDEIELTVSSELFVACNECAFPVCRPCYERREGNQACPQCKTRYKRIKGSPRVDGDD
SEQ ID NO:25 -----
SEQ ID NO:26 CDGNVMKDERGKDVMPCECRFKICRDCFMDAQKE-TGLCPGCKEQYK-----

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Figure 1 (cont'd.)

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SEQ ID NO:27 CGDQIGLTVEGDLFVACNECGFPACRCPYEYERREGTQNCPCQCKTRYKRLRGSPRVEGDE
SEQ ID NO:28 -----
SEQ ID NO:29 CRDEIELTVDGEPFVACNECAFPVCRPCYEYERREGNQACPQCKTRFKRLKGSPPRVEGD-

181 240
SEQ ID NO:2 -----
SEQ ID NO:4 EEEDGVDDLENEFNWSDK----HDSQYLAESMLHAHMSYG-RGADLDGVPQPFHPINVP
SEQ ID NO:6 -----
SEQ ID NO:8 EEEDGVDDLEGEFGLQDGAHEDDPQYVAESMLRAQMSYG-RGGDAH---PGFSPVPNVP
SEQ ID NO:10 EEEDGVDDLDNEFNW-DG----HDSQSVAESMLYGHMSYG-RGGDPNGAPQAFQLNPNVP
SEQ ID NO:12 -----
SEQ ID NO:14 DEEDV-DDIEHEFNIDEQKNKHGQ---VAEAMLHGRMSYG--RGPEDDDNSQFPTPIVAG
SEQ ID NO:16 -----
SEQ ID NO:18 -----
SEQ ID NO:20 -----
SEQ ID NO:22 -----P
SEQ ID NO:23 DEDDV-DDIENEFNYAQGANKARH---QRHGE---EFSSS--SRHESQPIPLLTHGHTVS
SEQ ID NO:24 EEEEDIDDLLEYEFD-----HGMDPEHAAEAALSSRLNTG--RGGLDSAPPG----SQIP
SEQ ID NO:25 -----
SEQ ID NO:26 -----IGDLDD-----TPDYSSGALPLPAPG-----
SEQ ID NO:27 DEEDI-DDIEYEFNIEHEQDKHKH---SAEAMLYGKMSYG--RGPEDDENGFRP-PVIAG
SEQ ID NO:28 -----
SEQ ID NO:29 EEEDDIDDLNEFEYGN---NGIGFDQVSEGMSISRRNSGFPQSDLSAPPG----SQIP

241 300
SEQ ID NO:2 -----
SEQ ID NO:4 LLTNGQMVDIPPDQHALVPSFV---GGGGKRIHPLPYADPNLPVQPRSMPSKDLAAYG
SEQ ID NO:6 -----
SEQ ID NO:8 LLTNGQMVDIPPEQHALVPSYMSGGGGGGKRIHPLPFADPNLPVQPRSMPSKDLAAYG
SEQ ID NO:10 LLTNGQMVDIPPEQHALVPSFM---GGGGKRIHPLPYADPSLPVQPRSMPSKDLAAYG
SEQ ID NO:12 -----
SEQ ID NO:14 GRSR-----PVSGEFPISSNAYGDQMLSSSLHKRVHPYPVSEPGSARW-----DEKKXDG
SEQ ID NO:16 -----
SEQ ID NO:18 -----
SEQ ID NO:20 -----
SEQ ID NO:22 LLTNGQMVDIPPEQHALVPSYMSGGGGGGKRIHPLPFADPNLPVQPRSMPSKDLAAYG
SEQ ID NO:23 GEIRTPDTQSVRTTSGPLGPSDRNAISSPYIDPR-QPVPVRIVDPSK-----DLNSYG
SEQ ID NO:24 LLTYCDEDADMYSRHALIVP--PS-TGYGNRVYPAPFTDSSAPPQARSMVPQKDIAEYG
SEQ ID NO:25 -----
SEQ ID NO:26 -----KDQRGNNNNNMSMMKRNQNGEFDHNRWLF-----ETQGTYG
SEQ ID NO:27 GHS-----GEFPVGG-GYGNG--EHGLHKRVHPYPSEAGS-----EGG
SEQ ID NO:28 -----
SEQ ID NO:29 LLTYGDEDVEISSDRHALIVP--PSLGGHGNRVHPVSLSDPTVAHRRLMVPQKDLAVYG

301 360
SEQ ID NO:2 -----
SEQ ID NO:4 YGSVAWKERMESWKQKQ-ERMHQTNRNDGGGD-----DGDDADLPLM-DEARQPLSR
SEQ ID NO:6 -----
SEQ ID NO:8 YGSVAWKERMEGWKQKQ-ERLQHVRSSEGGGDW-----DGDDADLPLM-DEARQPLSR
SEQ ID NO:10 YGSVAWKERMENWKQRQ-ERMHQTNRNDGGGD-----DGDDADLPLM-DEARQPLSR
SEQ ID NO:12 -----
SEQ ID NO:14 ----WKDRMDDWKLOQG-----NLGPEPDEDPDAAAML-DEARQPLSR
SEQ ID NO:16 -----
SEQ ID NO:18 -----HE-----
SEQ ID NO:20 -----
SEQ ID NO:22 YGSVAWKERMEGWKQKQ-ERLQHVRSSEGGGDW-----DGDDADLPLM-DEARQPLSR
SEQ ID NO:23 LGNVDWKERVEGWKLKQEKNNLQMTGKYHEGKGG-EIEGTGSNGEELQM-ADDTRLPMSR

```

Figure 1 (cont'd.)

SEQ ID NO:24 YGSVAWKDRMEVWKRROGEKLQVIKHEGGNNGRGSN-DDDELDDPDMPPM-DEGRQPLSR
 SEQ ID NO:25 -----
 SEQ ID NO:26 YGNAYWP-----QDEMYGD-----DMDEGMRGGMVETADKPWRPLSR
 SEQ ID NO:27 -----WRERMDDWKLQHG-----NLGPEPDDDPPEGLI-DEARQPLSR
 SEQ ID NO:28 -----
 SEQ ID NO:29 YGSVAWKDRMEEWKRKQNEKLQVVRHEGDP-----DFEDGDDADFPMM-DEGRQPLSM

361 420
 SEQ ID NO:2 -----
 SEQ ID NO:4 KIPLSSQINPYRMIIRLVVLGFFFHYRVMHPVDAFALWLISVICEIWFAMSWILDQ
 SEQ ID NO:6 -----
 SEQ ID NO:8 KVPISSSRINPYRMIIVIRLVVLGFFFHYRVMHPAKDAFALWLISVICEIWFAMSWILDQ
 SEQ ID NO:10 KIPLSSQINPYRMIIRLVVLGFFFHYRVMHPVNDALFALWLISVICEIWFAMSWILDQ
 SEQ ID NO:12 -----
 SEQ ID NO:14 KVPIASSKINPYRMVIVARLVILAFFLRYRLMNPVHDALGLWLTSIICEIWFASFILDQ
 SEQ ID NO:16 -----
 SEQ ID NO:18 -----LHPVNDAYGLWLTSVICEIWFASWIMDQ
 SEQ ID NO:20 -----
 SEQ ID NO:22 KVPISSSRINPYRMIIVIRLVVLGFFFHYRVMHPAKDAFALWLISVICEIWFAMSCILDQ
 SEQ ID NO:23 VVIPSSRLTPYRVVILRLIILCFFLQYRTHPVKNAYPLWLTSVICEIWFASFILDQ
 SEQ ID NO:24 KLPIRSSRINPYRMLILCRLAILGLFFHYRILHVPVNDAYGLWLTSVICEIWFASWILDQ
 SEQ ID NO:25 -----
 SEQ ID NO:26 RIPIPAAIISPYRLIVIRFVVLGFFLTWRIRNPNEAIWLWLMSIICELWFGFSWILDQ
 SEQ ID NO:27 KVPIASSKINPYRMVIVARLVILAVFLRYRLMNPVHDALGLWLTSVICEIWFASWILDQ
 SEQ ID NO:28 -----
 SEQ ID NO:29 KIPKSSKINPYRMLIVLRLVILGLFFHYRILHVPKDAYALWLISVICEIWFASWILDQ

421 480
 SEQ ID NO:2 -----
 SEQ ID NO:4 FPKWFPIERETYLDRSLRFDKEGHPS-----QLAPVDFVSTVDPLKEPPLVTANTVLS
 SEQ ID NO:6 -----
 SEQ ID NO:8 FPKWLPPIERETYLDRSLRFDKEGQPS-----QLAPIDFFVSTVDPTKEPPLVTANTVLS
 SEQ ID NO:10 FPKWFPIERETYLDRSLRFDKEGQPS-----QLAPIDFFVSTVDPLKEPPLVTANTVLS
 SEQ ID NO:12 -----
 SEQ ID NO:14 FPKWFPIDRETYLDRSLRIRYEREGEPN-----MLAPVDVVFVSTVDPMKEPPLVTANTVLS
 SEQ ID NO:16 -----
 SEQ ID NO:18 FPKWYPIQRETYLDRSLRYEKEGKPS-----ELSSVDVVFVSTVDPMKEPPLITANTVLS
 SEQ ID NO:20 -----
 SEQ ID NO:22 FPKWFPIERETYLDRSLRFDKEGQPS-----QLAPIDFFVSTVDPTKEPPLVTANTVLS
 SEQ ID NO:23 FPKWYPINRETYLDRSLAIRYDRDGEPS-----QLVPVDVVFVSTVDPLKEPPLVTANTVLS
 SEQ ID NO:24 FPKWYPIERETYLDRSLRYEKEGKPS-----GLAPVDVVFVSTVDPLKEPPLITANTVLS
 SEQ ID NO:25 -----
 SEQ ID NO:26 IPKLCPINRSTDLEVLRLDKFDMPSNPTGRSDLPGIDLFVSTADPEKEPPLVTANTVLS
 SEQ ID NO:27 FPKWFPIERETYLDRSLRYEREGEPN-----MLAPVDVVFVSTVDPLKEPPLVTANTVLS
 SEQ ID NO:28 -----
 SEQ ID NO:29 FPKWYPIERETYLDRSLRYEKEGKPS-----GLSPVDVVFVSTVDPLKEPPLITANTVLS

481 540
 SEQ ID NO:2 -----
 SEQ ID NO:4 ILSVDYPVDKVSVCYVSDDGAAMLTFEALSETSEFAKKWVPFCKRYSLEPRAPWEYFQ--
 SEQ ID NO:6 -----H-----
 SEQ ID NO:8 ILSVDYPVEKVSVCYVSDDGAAMLTFEALSETSEFAKKWVPFCKRYSLEPRAPWEYFQ--
 SEQ ID NO:10 ILSVDYPVDKVSVCYVSDDGAAMLTFEALSETSEFAKKWVPFCKRYSLEPRAPWEYFQ--
 SEQ ID NO:12 ILAAGYPAGKVTCYISDDAGAEVTRNAVVEAARFAALWVSFCRKHGVEPRNLEAYFNAGE
 SEQ ID NO:14 ILAMDYPVDKISVCYISDDGASMCTFESLSETAEFARKWVPFCKKFSIEPRAPEMYFSE--
 SEQ ID NO:16 -----
 SEQ ID NO:18 ILAVDYPVDKVACYSVCYVSDDGAAMLTFEALSETSEFARRWVPFCKKYSLEPRAPWEYFQ--

Figure 1 (cont'd.)

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SEQ ID NO:20 -----
SEQ ID NO:22 ILSVDYPVEKVS CYVSDDG AAMLT FEALSETSEFAKKWVPFSKKFNIEPRAPEWYFQ--
SEQ ID NO:23 ILSVDYPVDK VACYVSDDG SAMLTFESLSETAEFAKKWVPFCKKFNIEPRAPEFYFAQ--
SEQ ID NO:24 ILAVDYPVDK VACYVSDDG AAMLT FEALSDTAEFARKWVPFCKKFNIEPRAPEWYFSQ--
SEQ ID NO:25 -----RRWVPFCKKHNV EPRAP EYFNE--
SEQ ID NO:26 ILAVDYPVEKVS CYLSDDG GALLSFEAMAEASFADLWVPFCRKHNIEPRNPDSYFSL--
SEQ ID NO:27 ILAMDYPVEKIS CYVSDDG ASMLTFESLSETAEFARKWVPFCKKFSIEPRAPEMYFTL--
SEQ ID NO:28 ----DYPVEKVS CYVSDDG AAMLT FEALSETSEFARKWVPFCKKYNIEPRAPEWYFAQ--
SEQ ID NO:29 ILAVDYPVDK VACYVSDDG AAMLT FEALSETAEFARKWVPFCKKYCIEPRAPEWYFCH--

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541

600

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SEQ ID NO:2 -----
SEQ ID NO:4 -----KIDY LKDKVAPNFVRERRAMKREYEEFKVRINALVAKAQ-----
SEQ ID NO:6 -----
SEQ ID NO:8 -----KIDY LKDKVAASFVRERRAMKREYEEFKVRINALVAKAQ-----
SEQ ID NO:10 -----KIDY LKDKVAANFVRERRAMKREYEEFKVRINALVAKAQ-----
SEQ ID NO:12 GGGGKAKVVARGSY-RGMAWPELVDRRRVRREYEEMRLRIDALQAADARRR-----
SEQ ID NO:14 -----KIDY LKDKVQPTFVKERRAMKREYEEFKVRINALVAKAQ-----
SEQ ID NO:16 -----AKAQ-----
SEQ ID NO:18 -----KMDY LKNKVHPAFVRERRAMKRDYEEFKVRINSLVATAQ-----
SEQ ID NO:20 -----
SEQ ID NO:22 -----KIDY LKDKVAASFVRERRAMKREYEEFKVRINALVAKAQ-----
SEQ ID NO:23 -----KIDY LKDKIQPSFVKERRAMKREYEEFKVRINALVAKAQ-----
SEQ ID NO:24 -----KMDY LKNKVHPAFVRERRAMKRDYEEFKVKINALVATAQ-----
SEQ ID NO:25 -----KIDY LKDKVHPSFVKERRAMKREYEEFKVRINALVAKAQ-----
SEQ ID NO:26 -----KIDPTKNKSRI DFKDRRKIKREYDEFKVRINGLPDSIRRRSDAFNAREE
SEQ ID NO:27 -----KVDY LQDKVHPTFVKERRAMKREYEEFKVRINAQVAKAS-----
SEQ ID NO:28 -----KIDY LKDKVQTSFVKERRAMKREYEEFKVRVNLVAKAQ-----
SEQ ID NO:29 -----KMDY LKNKVHPAFVRERRAMKRDYEEFKVKINALVATAQ-----

```

601

660

```

SEQ ID NO:2 -----
SEQ ID NO:4 -----KVPEEGWTMQDGTWPWG-----NNVRDHPGMIQVFL---
SEQ ID NO:6 -----
SEQ ID NO:8 -----KVPEEGWTMQDGSPWPG-----NNVRDHPGMIQVFL---
SEQ ID NO:10 -----KVPEEGWTMQDGTWPWG-----NNVRDHPGMIQVFL---
SEQ ID NO:12 -----RRGAADHDHAGVVQVLIDFA
SEQ ID NO:14 -----KVPQGGWIMQDGTWPWG-----NNTKDHPGMIQVFL---
SEQ ID NO:16 -----KMPEEGWTMQDGTWPWG-----NNPRDHPGMIQVFL---
SEQ ID NO:18 -----KVPEDGWTMQDGTWPWG-----NNVRDHPGMIQVFL---
SEQ ID NO:20 -----
SEQ ID NO:22 -----KVPEEGWTMQDGSPWPG-----
SEQ ID NO:23 -----KIPEEGWTMQDGTWPWG-----NNTRDHPGMIQVFL---
SEQ ID NO:24 -----KVPEEGWTMQDGTWPWG-----NNVRDHPGMIQVFL---
SEQ ID NO:25 -----KKPEEGWVMQDGTWPWG-----NNTRDHPGMIQVYL---
SEQ ID NO:26 MKALKQMRESGGDPTEPVKPKATW-MADGTHWPGTWAASTREHSGDGHAGILQVMLKPP
SEQ ID NO:27 -----KVPLEGWIMQDGTWPWG-----NNTKDHPGMIQVFL---
SEQ ID NO:28 -----KVPEEGWIMQDGTWPWG-----NNTRDHPGMIQVFL---
SEQ ID NO:29 -----KVPEDGWTMQDGTWPWG-----NSVRDHPGMIQVFL---

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661

720

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SEQ ID NO:2 -----
SEQ ID NO:4 -----G-QSGGHDVE----GNELPRLVYVSREKRPGYNHHKKAGAMNALVRVSAVLTNA
SEQ ID NO:6 -----
SEQ ID NO:8 -----G-QSGGRDVE----GNELPRLVYVSREKRPGYNHHKKAGAMNALVRVSAVLSNA
SEQ ID NO:10 -----G-QSGGLDCE----GNELPRLVYVSREKRPGYNHHKKAGAMNALVRVSAVLTNA
SEQ ID NO:12 GSV PQ LGVANGSKLIDVASVDVCLPALVYVCREKRRGHAAHHRKAGAMNA-----

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Figure 1 (cont'd.)

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SEQ ID NO:14 -----G-SSGGLDTE----GNQLPRLVYVSREKRPGFQHHKKAGAMNALVRVSAVLTNA
SEQ ID NO:16 -----G-HSGGLDTD----GNELPRLVYVSREKRPGFQHHKKAGAMNALIRVSAVLTNG
SEQ ID NO:18 -----G-QDGV RDVE----GNELPRLVYVSREKRPGFQHHKKAGAMNALVRASAIITNA
SEQ ID NO:20 -----
SEQ ID NO:22 -----
SEQ ID NO:23 -----G-HSGGLDTD----GNELPRLIYVSREKRPGFQHHKKAGAMNALIRVSAVLTNG
SEQ ID NO:24 -----G-HSGV RDTD----GNELPRLVYVSREKRPGFQHHKKAGAMNSLIRVSAVLSNA
SEQ ID NO:25 -----G-SAGALDVD----GKELPRLVYVSREKRPQYQHHKKAGAENALVRVSAVLTNA
SEQ ID NO:26 SSDPLIG-NSDDKVIDFSDDTDLPLMFVYVSREKRPQYDHNKKAGAMNALVRASAILSNG
SEQ ID NO:27 -----G-HSGGFDVE----GHELPLRVYVSREKRPGFQHHKKAGAMNALVRVAGVLTNA
SEQ ID NO:28 -----G-QSGGLDAE----GNELPRLVYVSREKRPGFQHHKKAGAMNALVRVSAVLTNG
SEQ ID NO:29 -----G-SDGV RDVE----NNELPRLVYVSREKRPGFQHHKKAGAMNSLIRVSGVLSNA

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721
SEQ ID NO:2 -----
SEQ ID NO:4 PYLLNLDCDHYINNSKAIKEAMCFMMDPLLGGK-----VCYVQFPQRFDGIDRHDRYAN
SEQ ID NO:6 -----
SEQ ID NO:8 AYLLNLDCDHYINNSKAIKEAMCFMMDPLVGKK-----VCYVQFPQRFDGIDKNDRYAN
SEQ ID NO:10 PYLLNLDCDHYINNSKAIKEAMCFMMDPLLGGK-----VCYVQFPQRFDGIDRHDRYAN
SEQ ID NO:12 PFILDLDCDHYVNNNSQALRAGICFMIERGGGGAEDAGAVAFVQFPQRFVGDVDPGDRYAN
SEQ ID NO:14 PFMLNLDCDHYVNNNSKAAREAMCFMMDPQTGGK-----VCYVQFPQRFDGIDTHDRYAN
SEQ ID NO:16 AYLLNVDCDHYFNNSKALKEAMCFMMDPVLGGK-----TCYVQFPQRFDGIDLHDRYAN
SEQ ID NO:18 PYLLNVDCDHYINNSKALREAMCFMMDPQLGGK-----VCYVQFPQRFDGIDRHDRYSN
SEQ ID NO:20 -----EAMCFLMDPNLGPQ-----VCYVQFPQRFDGIDRNDRYAN
SEQ ID NO:22 -----
SEQ ID NO:23 AYLLNVDCDHYFNNSKAIKEAMCFMMDPAIGKK-----CCYVQFPQRFDGIDLHDRYAN
SEQ ID NO:24 PYLLNVDCDHYINNSKAIRESMCFMMDPQSGKK-----VCYVQFPQRFDGIDRHDRYSN
SEQ ID NO:25 PFILNLDCDHYINNSKAMREAMCFMMDPQFGKK-----LCYVQFPQRFDGIDRHDRYAN
SEQ ID NO:26 PFILNLDCDHYIYNCKAVREGMCFMMDRG-GED-----ICYIQFPQRFEGIDPSDRYAN
SEQ ID NO:27 PFMLNLDCDHYVNNNSKAVREAMCFMMDPQIGKK-----VCYVQFPQRFDGIDTNDRYAN
SEQ ID NO:28 AFLNLDCDHYINNSKALREAMCFMMDPNLGGKQ-----VCYVQFPQRFDGIDRNDRYAN
SEQ ID NO:29 PYLLNVDCDHYINNSKALREAMCFMMDPQSGKK-----ICYVQFPQRFDGIDRHDRYSN

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781
SEQ ID NO:2 -----
SEQ ID NO:4 RNVVFFDINMKGLDGIQGPIYVGTGCVFRRQALYGYDAP---KTKKPPSRTCNCWPKWCI
SEQ ID NO:6 -----
SEQ ID NO:8 RNVVFFDINMKGLDGIQGPIYVGTGCVFRRQALYGYDAP---KTKKPPSRTCNCWPKWCL
SEQ ID NO:10 RNVVFFDINMKGLDGIQGPIYVGTGCVFRRQALYGYDAP---KTKKPPSRTCNCWPKWCF
SEQ ID NO:12 HNRVLFDCDELGLDGLQGPIYVGTGCLFRRVALYSVDLPR-----
SEQ ID NO:14 RNTVFFDINMKGLDGIQGPVYVGTGCVFRRQALYGYNPPKGP--RPMVSCDC-----
SEQ ID NO:16 RNIVFFDINMKGQDGVQGPVYVGTGCCFNQALYGYDPVLTEEDLE-----PNIIV
SEQ ID NO:18 RNVVFFDINMKGLDGIQGPIYVGTGCVFRRYALYGYDAP---AKKKPPSKTCNCWPKWCC
SEQ ID NO:20 RNTVFFDINLRGLDGIQGPVYVGTGCVFNRTAIYGYEPPKAK--K-----PGFLA
SEQ ID NO:22 -----
SEQ ID NO:23 RNIVFFDINMKGLDGIQGPVYVGTGCCFNQALYGYDPVLTEEDLE-----PNIIV
SEQ ID NO:24 RNVVFFDINMKGLDGIQGPIYVGTGCVFRRQALYGFADAP---KKKKPPGKTCNCWPKWCC
SEQ ID NO:25 RNVVFFDINMLGLDGLQGPIYVGTGCVFNQALYGYDPPVSEKRPK---MTCDCWPSWCC
SEQ ID NO:26 NNTVFFDGNMRALDGVQGPVYVGTGTMFRRFALYGFDP-----
SEQ ID NO:27 RNTVFFDINMKGLDGIQGPVYVGTGCVFKRQALYGYEPPKGP--RPMISCGC-----
SEQ ID NO:28 RNTVFFDINLRGLDGIQGPVYVGTGCVFNRTALYGYEPPKPKHRK-----TGILS
SEQ ID NO:29 RNVVFFDINMKGLDGLQGPIYVGTGCVFRRQALYGFADAP---KKKKGPRKTCNCWPKWCL

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Figure 1 (cont'd.)

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841
SEQ ID NO:2 -----
SEQ ID NO:4 CCCCFCNRKTKKKTKTSKP-----KFEKIKKLF-KKKENQAPAYALGEIDEA--APG-
SEQ ID NO:6 -----
SEQ ID NO:8 SCCC---SRNKNKKKTTKP-----KTEKKKRLFFKKAENPSPAYALGEIDEG--APG-
SEQ ID NO:10 CCCCFCNRKQKK---TTKP-----KTEKKKLLFFKKEENQSPAYALGEIDEA--APG-
SEQ ID NO:12 -----
SEQ ID NO:14 -CPCFCGRKKKYKE-----KNDANGEAASLKG
SEQ ID NO:16 -KSCCGSRKKKGKGNKK-----YS-DKKKAMGR--TESTVPIFNMEDIEEGVEG--Y
SEQ ID NO:18 LCC--GSRKKKN---ANS-----KKEKKRKV--KHSEASKQIHALENIEAGN--EG-
SEQ ID NO:20 -SLCXG-KKKASKSKKR-----SSDKKSNKH--VDSSVPVFNLEDIEEGVEGAGF
SEQ ID NO:22 -----
SEQ ID NO:23 -KSCCGSRKKKGKSS-KK-----YNYEKRRGINR--SDSNAPLFNMEDIDEGFEG--Y
SEQ ID NO:24 LCC--GLRKKSK---T-----KAKDKKT--NTKETSQIHALENVDEGVIVPV-
SEQ ID NO:25 -CCCGSRKKSKKKGEKKGLGGLLYGKKKKMMGKNYVKKGSAPVFDLEEIEEGLEG--Y
SEQ ID NO:26 -----NPDKLLLEKKESETEALTTSDFDPLDVTQLPKRFGNSTLL-----AESIPI
SEQ ID NO:27 -CPCFCGRRRKNNK-----FSKNDMNGDVAAALGG
SEQ ID NO:28 -SLCGSRKKSSSKSSK-----GSDKKKSGKH--VDSTVPVFNLEDIEEGVEGAGF
SEQ ID NO:29 LCF--GSRKNRK---AKT-----VAADKKK--KNREASKQIHALENIEEGRGHKV-

901
SEQ ID NO:2 -----
SEQ ID NO:4 AENEKASIVNQKLEKKFGQSSVFVASTLLENGGTLKSASPASLLKEAIHVISCYEDKT
SEQ ID NO:6 -----ET
SEQ ID NO:8 ADIEKAGIVNQKLEKKFGQSSVFVASTLLENGGTLKSASPASLLKEAIHVISCYEDKT
SEQ ID NO:10 AENEKAGIVNQKLEKKFGQSSVFVASTLLENGGTLKSASPASLLKEAIHVISCYEDKT
SEQ ID NO:12 -----
SEQ ID NO:14 MDDDKEVLMSQMNFEKKFGQSSIFVTSTLMEEGGVPPSSSPAALLKEAIHVISCYEDKT
SEQ ID NO:16 DD-ERTLLMSQKSLEKRFQSPVFIAATFMEQGGIPPTNPATLLKEAIHVISCYEDKT
SEQ ID NO:18 TNNEKTSNLTQTKLEKRFQSPVFVASTLLDDGGVPHGVSPASLLKEAIHVISCYEDKT
SEQ ID NO:20 DD-EKSVLMSQMSLEKRFQSSAAFVASTLMEYGGVPPQSSTPESLLKEAIHVISCYEDKS
SEQ ID NO:22 -----
SEQ ID NO:23 DD-ERSILMSQRSVEKRFQSPVFIAATFMEQGGIPPTNPATLLKEAIHVISCYEDKT
SEQ ID NO:24 SNVEKRSEATQLKLEKKFGQSPVFVASAVLQNGGVPRNASPACLLREAIHVISCYEDKT
SEQ ID NO:25 EELEKSTLMSQKNFEKRFQSPVFIASTLMENGGLPEGTNSTSLIKEAIHVISCYEEKT
SEQ ID NO:26 AEFQGRPLADHPAV--KYGRPP---GALR---VPRDPLDATTVAESVSVISCWYEDKT
SEQ ID NO:27 AEGDKHELMFEMNFEKTFGQSSIFVTSTLMEEGGVPPSSSPAALLKEAIHVISCYEDKT
SEQ ID NO:28 DD-EKSLMSQMSLEKRFQSSAAFVASTLMENGGVPPQSATPETLLKEAIHVISCYEDKT
SEQ ID NO:29 LNVEQSTEAMQMKLQKKYGQSPVFVASARLENGGMARNASPACLLKEAIHVISCYEDKT

961
SEQ ID NO:2 -----HEDILTGFKMHWISYICMPPRPCFKGSAPINLSDRLNQVLRWAL
SEQ ID NO:4 GWGKDIGWIYGSVTEDILTGFKMCHGWRSIYICPKRAAFKGSAPINLSDRLHQVLRWAL
SEQ ID NO:6 EWGKEIGWIYGSVTEDILTGFKMCHGWKSVYCTPTRPAFKGSAPINLSDRLHQVLRWAL
SEQ ID NO:8 DWGKEIGWIYGSITEDILTGFKMCHGWRSIYICPKRPAFKGSAPINLSDRLHQVLRWAL
SEQ ID NO:10 DWGKEIGWIYGSVTEDILTGFKMCHGWRSIYICPKRVAFKGSAPINLSDRLHQVLRWAL
SEQ ID NO:12 -----WRP-----RRSL
SEQ ID NO:14 EWGLELGWIYGSITEDILTGFKMCHGWRSIYICMPKRAAFKGTAPINLSDRLNQVLRWAL
SEQ ID NO:16 EWGKEIGWIYGSVTEDILTGFKMHWISYICMPPRPAFKGSAPINLSDRLNQVLRWAL
SEQ ID NO:18 EWGKEVGWIYGSVTEDILTGFKMCHGWRSVYICPKRPAFKGSAPINLSDRLHQVLRWAL
SEQ ID NO:20 EWGTEIGWIYGSVTEDILTGFKMHWISYICMPKRAAFKGSAPINLSDRLNQVLRWAL
SEQ ID NO:22 -----
SEQ ID NO:23 EWGKEIGWIYGSVTEDILTGFKMHWISYICNPPRPAFKGSAPINLSDRLNQVLRWAL
SEQ ID NO:24 EWGKEIGWIYGSVTEDILTGFKMCHGWRSVYICMPKRAAFKGSAPINLSDRLHQVLRWAL
SEQ ID NO:25 EWGKEIGWIYGSVTEDILTGFKMCHGWKSVYICPKRPAFKGSAPINLSDRLHQVLRWAL
SEQ ID NO:26 EWGDRVGWIYGSVTEDVVTGYRMHNRGWRSVYICITKRDSFRGSAPINLTDRLHQVLRWAT

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Figure 1 (cont'd.)

SEQ ID NO:27 EWGTELGWIIYGSITEDILTGFKMHCGRWRSIYCMKRPAPFKGSAPINLSDRLNQVLRWAL
 SEQ ID NO:28 DWGSEIGWIIYGSVTEDILTGFKMARGWRSIYCMKRPAPFKGSAPINLSDRLNQVLRWAL
 SEQ ID NO:29 EWGKEIGWIIYGSVTEDILTGSKMHSWGWRHVYCTPKLAAPFKGSAPINLSDRLHQVLRWAL

1021 1080
 SEQ ID NO:2 GSVEILFSRHCPWIWYNYGG-RLKLLERMAYINTIVYPITSLPLIAYCVLPALCLLTNKF
 SEQ ID NO:4 GSIEIFFSNHCPWYGYGGG-LKFLERFSYINSIVYPWTSIPLLAYCTLPALCLLTGKFI
 SEQ ID NO:6 GSVEIFMSRHCPWYAYGG-RLKWLERFAYTNTIVYPFTSIPLLAYCTIPAVCLLTGKFI
 SEQ ID NO:8 GSVEIFFSKHCPWYGYGGG-LKFLERFSYINSIVYPWTSIPLLAYCTLPALCLLTGKFI
 SEQ ID NO:10 GSIEIFFSNHCPWYGYGGG-LKFLERFSYINSIVYPWTSIPLLAYCTLPALCLLTGKFI
 SEQ ID NO:12 G-----CRL-----
 SEQ ID NO:14 GSIEIFFSHHCPWYGFKEKKLKWLERFAYANTTVYPFTSIPLVAYCILPAVCLLTGKFI
 SEQ ID NO:16 GSIEIFLSRHCPWYGYNG-KLKPLMRLAYINTIVYPFTSIPLIAYCTLPALCLLTNKF
 SEQ ID NO:18 GSVEIFFSRHCPWIWYGYGGG-LKLLERFSYINSVVPWTSIPLLVYCTLPALCLLTGKFI
 SEQ ID NO:20 GSVEILFSRHCPWYGYGG-RLKFLERFAYINTTIYPLTSLPLLVYCILPAICLLTGKFI
 SEQ ID NO:22 -----
 SEQ ID NO:23 GSIEILLSRHCPWIWYGYHG-RLRLLERIAYINTIVYPITSIPLIAYCILPAFCLITDRFI
 SEQ ID NO:24 GSVEIFLSRHCPWIWYGYGGG-LKWLERFSYINSVVPWTSIPLIVYCSLPAVCLLTGKFI
 SEQ ID NO:25 GSVEIFLSRHCPWYGYGG-KLKWLERLAYINTIVYPFTSIPLLAYCTIPAVCLLTGKFI
 SEQ ID NO:26 GSVEIFFSRNNAI---LASKRLKFLQRLAYLNVGIYPFTSLFLILYCFLPASFSLFGQFI
 SEQ ID NO:27 GSVEIFFSRHSPWYGYKGGKLKWLERFAYANTTIYPFTSIPLLAYCILPAICLLTDKFI
 SEQ ID NO:28 GSVEILFSRHCPWIWYGYSG-RLKWLERFAYVNTTIYPVTAIPLMYCTLPVAVCLLTNKF
 SEQ ID NO:29 GSVEIFLSRHCPWIWYGYGGG-LKWLERLSYINSVVPWTSIPLIVYCSLPAICLLTGKFI

1081 1140
 SEQ ID NO:2 IPEISNYAGMFFILMFASIFATGILELRWSGVGIEDWWRNEQFWVIGGSAHLFAVFQGL
 SEQ ID NO:4 TPELNNVASLWFMFLFICIFATSILEMRWSGVGIDDWWRNEQFWVIGGVSSHFAVFQGL
 SEQ ID NO:6 IPTLNNLASIWFIALFLSIIATSVLELRWSGVSIEDWWRNEQFWVIGGVSAHLFAVFQGL
 SEQ ID NO:8 TPELTNVAISWFMALFICISVTGILEMRWSGVAIDDWWRNEQFWVIGGVSAHLFAVFQGL
 SEQ ID NO:10 TPELNNVASLWFMFLFICIFATSILEMRWSGVGIDDWWRNEQFWVIGGVSSHFAVFQGL
 SEQ ID NO:12 -----
 SEQ ID NO:14 MPPISTFAGLYFVALFSSIIATGILELKWSGVSIEEWWRNEQFWVIGGVSAHLFAVQGL
 SEQ ID NO:16 IPEISNFASMWFIILFVSIFTTSILELRWSGVSIEDWWRNEQFWVIGGSAHLFAVFQGL
 SEQ ID NO:18 VPEISNYASLVFMALFISIAATGILEMQWGGVSIDDWWRNEQFWVIGGVSSHFAVFQGL
 SEQ ID NO:20 MPEISNLASIWFIALFLSIFATGILEMRWSGVGIEDWWRNEQFWVIGGISAHFAVFQGL
 SEQ ID NO:22 -----
 SEQ ID NO:23 IPEISNYASIWFIILFISIAVTGILELRWSGVSIEDWWRNEQFWVIGGSAHLFAVFQGL
 SEQ ID NO:24 VPEISNYAGILFMLMFISIAVTGILEMQWGGVGIDDWWRNEQFWVIGGASSHFAVFQGL
 SEQ ID NO:25 IPTLSNLTSVWFLALFLSIIATGILELRWSGVSIQDWRNEQFWVIGGVSAHLFAVFQGL
 SEQ ID NO:26 VRTLSISFLVYLLMITICILGLAVLEVKWSGIGLEEWWRNEQWLLISGTSSHLAYVQGV
 SEQ ID NO:27 MPPISTFASLFFISLFMSIIVTGILELRWSGVSIEEWWRNEQFWVIGGISAHFAVFQGL
 SEQ ID NO:28 IPQISNLASIWFIISLFLSIFATGILKMKWNGVGIDQWWRNEQFWVIGGVSAHLFAVFQGL
 SEQ ID NO:29 VPEISNYASILFMALFSSIAITGILEMQWGVGIDDWWRNEQFWVIGGVSAHLFAVFQGL

1141 1200
 SEQ ID NO:2 LKVLAGIDTNFTVTSKANDEDGD--FAELYVFKWTSLLIPPTTVLVINLVGMVAGISYAI
 SEQ ID NO:4 LKVIAGVDTSTFTVTSKGGDD--EE-FSELYTFKWTLLIPPTLTLNFIGVAGISNAI
 SEQ ID NO:6 LKVLGGVDTSTFTVTSKAAGDEADA-FGDLYLFKWTLLVPPTTLIIINMVGIVAGVSDAV
 SEQ ID NO:8 LKVFAIDTSTFTVTSKAGDD--EE-FSELYTFKWTLLIPPTLTLNFIGVAGISNAI
 SEQ ID NO:10 LKVIAGVDTSTFTVTSKGGDD--EE-FSELYTFKWTLLIPPTLTLNFIGVAGVSNAI
 SEQ ID NO:12 -----
 SEQ ID NO:14 LKVLAGIDTNFTVTSKATDDE-E--FGELYTFKWTLLIPPTTLIIINIVGVVAGISDAI
 SEQ ID NO:16 LKVLAGIDTNFTVTSKASDEDGD--FAELYVFKWTSLLIPPTTVLVINLVGIVAGVSYAI
 SEQ ID NO:18 LKVLAGVNTNFTVTSKAADD--GE-FSELYIFKWTLLIPPTTLIMNIVGVVAGISDAI
 SEQ ID NO:20 LKVLAGIDTNFTVTSKANDEEGD--FAELYMFKWTLLIPPTTLIIINMVGIVAGTSYAI
 SEQ ID NO:22 -----
 SEQ ID NO:23 LKVLAGIDTNFTVTSKATDEDGD--FAELYIFKWTALLIPPTTVLLVNLIGIVAGVSYAV

Figure 1 (cont'd.)

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SEQ ID NO:24 LKVLAVNTNFTVTSKAADD--GA-FSELYIFKWTLLIPPTLLIINIIGVIVGVSDAI
SEQ ID NO:25 LKVLAVDTNFTVTAKAADDTE---FGELYLFKWTLLIPPTLLIILNMVGVVAGVSDAI
SEQ ID NO:26 LKVIAGIEISFTLTTKSGDDNEDIYADLYIVKWSSLMIPPIVIAMVNIIAIVVAFIRTI
SEQ ID NO:27 LKILAGIDTNFTVTSKATDDD-D--FGELYAFKWTLLIPPTTVLIINIVGVVAGISDAI
SEQ ID NO:28 LKVLAVDTNFTVTSKASDEGD--FAELYMFKWTLLIPPTLLIINLVGVVAGISYVI
SEQ ID NO:29 LKVLAVDTNFTVTSKAADD--GE-FSDLYLFKWTSLIPPMPTLLIINIVGVIVGVSDAI

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1201 1260
SEQ ID NO:2 NSGYQSWGPLEFGKLFFSIWVILHLYPFLKGLMGRQNRTPPTIVIVWSILLASIFSLWVKI
SEQ ID NO:4 NNGYESWGPLEFGKLFFAFWVIVHLYPFLKGLVGRQNRTPPTIVIVWSILLASIFSLWVRI
SEQ ID NO:6 NNGYGSWGPLEFGKLFFSFWVIVHLYPFLKGLMGRQNRTPPTIVVLSILLASIFSLWVRI
SEQ ID NO:8 NNGYESWGPLEFGKLFFAFWVIVHLYPFLKGLVGRQNRTPPTIVIVWSILLASIFSLWVRI
SEQ ID NO:10 NNGYESWGPLEFGKLFFAFWVIVHLYPFLKGLVGRQNRTPPTIVIVWSILLASIFSLWVRI
SEQ ID NO:12 -----LGEDER-----LWSRM
SEQ ID NO:14 NNGYQSWGPLEFGKLFFSFWVIVHLYPFLKGLMGRQNRTPPTIVIVWSVLLASIFSLWVRI
SEQ ID NO:16 NSGYQSWGPLEFGKLFFAIWVIAHLYPFLKGLLGRQNRTPPTIVIVWSVLLASIFSLWVRI
SEQ ID NO:18 NNGYDSWGPLEFGKLFFALWVILHLYPFLKGLLGRQNRTPPTIVIVWSVLLASIFSLWVRI
SEQ ID NO:20 NSGYQSWGPLEFGKLFFAFWVIVHLYPFLKGLMGRQNRTPPTIVIVWAVLLASIFSLWVRI
SEQ ID NO:22 -----
SEQ ID NO:23 NSGYQSWGPLEFGKLFFALWVIAHLYPFLKGLLGRQNRTPPTIVIVWSVLLASIFSLWVRI
SEQ ID NO:24 NNGYDSWGPLEFGKLFFALWVIVHLYPFLKGLMGRQNRTPPTIVIVWSVLLASIFSLWVRI
SEQ ID NO:25 NNGYGSWGPLEFGKLFFAFWVILHLYPFLKGLMGRQNRTPPTIVVLSILLASIFSLWVRI
SEQ ID NO:26 YQAVPQWSKLIGGAFFSFWVLAHLYPFAKGLMGRGKTPTIVFVWAGLIAITISLLWTAI
SEQ ID NO:27 NNGYQSWGPLEFGKLFFSFWVIVHLYPFLKGLMGRQNRTPPTIVIVWSVLLASIFSLWVRI
SEQ ID NO:28 NSGYQSWGPLEFGKLFFAFWVILHLYPFLKGLMGRQNRTPPTIVVWSILLASIFSLWVRI
SEQ ID NO:29 NNGYDSWGPLEFGKLFFALWVILHLYPFLKGLLGRQNRTPPTIVVWSILLASIFSLWVRI

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1261 1284
SEQ ID NO:2 DPFISDTQKAVAM-GQCGVNC---
SEQ ID NO:4 DPFLAKDDGPILL--EECGLDCN--
SEQ ID NO:6 DPFIPKAKGPILKP-C-GVEC---
SEQ ID NO:8 DPFLAKSNGPILL--EECGLDCN--
SEQ ID NO:10 DPFLAKDDGPILL--EECGLDCN--
SEQ ID NO:12 KQMVILSGPR-----
SEQ ID NO:14 DPFVLKTKGPDTKL--CGINC---
SEQ ID NO:16 DPFTSDSNKLT--NGQCGINC---
SEQ ID NO:18 NPFVSRD-GPVL--EICGLNCDES
SEQ ID NO:20 DPFTTRLAGPNI--QTCGINC---
SEQ ID NO:22 -----
SEQ ID NO:23 NPFVDANPNANNFNKGKGVF----
SEQ ID NO:24 NPFVAKG-GPVL--EICGLNCGN-
SEQ ID NO:25 DPFLPKQTGPVLKQ-C-GVEC---
SEQ ID NO:26 NP---NTGPAAAAEGVGGGGFQFP
SEQ ID NO:27 DPFVLKTKGPDTSK--CGINC---
SEQ ID NO:28 DPFTTRVTGPDV--EQCGINC---
SEQ ID NO:29 NPFVAKG-GPIL--EICGLDC--L

```

DECLARATION and POWER OF ATTORNEY

As a below-named inventor, I hereby declare that:

My residence, post office address and citizenship are as stated below next to my name.

I believe I am the original, first and sole inventor (if only one name is listed below) or an original, first and joint inventor (if plural names are listed below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

PLANT CELLULOSE SYNTHASES

the specification of which is attached hereto unless the following box is checked:

☒ was filed on **13 JULY 1999** as U.S. Application No. _____ or PCT International Application No. _____
PCT/US99/15871 and was amended on _____ (if applicable).

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is known to me to be material to patentability as defined in 37 CFR § 1.56.

I hereby claim foreign priority benefits under 35 U.S.C. § 119(a)-(d) or § 365(b) of any foreign application(s) for patent or inventor's certificate, or § 365(a) of any PCT International application which designated at least one country other than the United States, listed below and have also identified below, by checking the box, any foreign application for patent or inventor's certificate, or PCT International application having a filing date before that of the application on which priority is claimed.

Application No.	Country	Filing Date	Priority Claimed (Yes/No)
-----------------	---------	-------------	---------------------------

I hereby claim the benefit under 35 U.S.C. § 119(e) of any United States Provisional Application(s) listed below.

U.S. Provisional Application No.**U.S. Filing Date**

60/092,844

14 JULY 1998

I hereby claim the benefit under 35 U.S.C. § 120 of any United States application(s), or § 365(c) of any PCT International Application designating the United States, listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States application or PCT International Application in the manner provided by the first paragraph of 35 U.S.C. § 112, I acknowledge the duty to disclose information which is known to me to be material to patentability as defined in 37 CFR § 1.56 which became available between the filing date of the prior application and the national or PCT International filing date of this application.

Application No.	Filing Date	Status (patented, pending or abandoned)
-----------------	-------------	---

POWER OF ATTORNEY: I hereby appoint the following attorney(s) and/or agent(s) the power to prosecute this application and transact all business in the Patent and Trademark Office connected therewith:Name: **WILLIAM R. MAJARIAN**Registration No.: **41,173**

Send correspondence and direct telephone calls to:

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Wilmington, DE 19898, U.S.A.**Tel. No.****(302) 992-4926****Fax No.****(302) 892-7949**

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issuing thereon.

INVENTOR(S)

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Signature (please sign full name): <i>[Signature]</i>			Date: 8/30/99
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Signature (please sign full name): <i>[Signature]</i>			Date: 9/1/99
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Residence & Citizenship	City ARDEN	State or Foreign Country DELAWARE	Country of Citizenship U.S.A.
Post Office Address	Post Office Address 1902 MILLERS ROAD	City ARDEN	State or Country DELAWARE Zip Code 19810

☒ Additional Inventors are being named on separately numbered sheets attached hereto.

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	Signature (please sign full name): <i>Anthony Kinney</i>		Date: <i>3 Sept 99</i>
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Full Name of Inventor	Last Name LIGHTNER	First Name JONATHAN	Middle Name E.
	Signature (please sign full name): <i>Jonathan Lightner</i>		Date: <i>16 Sept 99</i>
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Full Name of Inventor	Last Name MIAO	First Name GUO-HUA	Middle Name
	Signature (please sign full name): <i>Guo Hua Miao</i>		Date: <i>9-9-1999</i>
Residence & Citizenship	City HOCKESSIN	State or Foreign Country DELAWARE <i>DE</i>	Country of Citizenship CN
Post Office Address	Post Office Address 202 CHERRY BLOSSOM PLACE	City HOCKESSIN	State or Country DELAWARE Zip Code 19707
Full Name of Inventor	Last Name RAFALSKI	First Name J.	Middle Name ANTONI
	Signature (please sign full name): <i>J. Antoni Rafalski</i>		Date: <i>5-8-1999</i>
Residence & Citizenship	City WILMINGTON	State or Foreign Country DELAWARE <i>DE</i>	Country of Citizenship U.S.A.
Post Office Address	Post Office Address 2028 LONGCOME DRIVE	City WILMINGTON	State or Country DELAWARE Zip Code 19810
Full Name of Inventor	Last Name THORPE	First Name CATHERINE	Middle Name J.
	Signature (please sign full name): <i>Catherine S. Thorpe</i>		Date: <i>21st September 1999</i>
Residence & Citizenship	City CAMBRIDGE	State or Foreign Country UNITED KINGDOM <i>GBX</i>	Country of Citizenship GB
Post Office Address	Post Office Address 20 THE BEECHES, WOODHEAD DRIVE	City CAMBRIDGE	State or Country UNITED KINGDOM Zip Code CR4 1FY

GENERAL POWER OF ATTORNEY
(Concerning Several International Patent Applications)

The undersigned, Vernon R. Rice, Vice President and Assistant General Counsel of E. I. DU PONT DE NEMOURS AND COMPANY, 1007 Market Street, Wilmington, Delaware 19898 USA ("DuPont"), hereby confirms that the power to sign for DuPont has been granted to various individuals (as set forth in the attached excerpt from DuPont's Patent Board Rules of Procedure (January 1988), Appendix Section III.A.4), including the Chairman, Vice-Chairman, and those individuals who are Assistant Secretaries of the Patent Board. Currently these Assistant Secretaries are:

Roger A. Bowman
Linda J. Davis
John E. Griffiths

Miriam D. McConahey
Dorothy W. Shafer
Deborah A. Meginniss

In addition, the authority to act on behalf of DuPont before the competent International Authorities in connection with any and all international patent applications filed by it with the United States as Receiving Office and to make or receive payments on its behalf is hereby granted to:

Beardell, Lori Y.	34,293	Katz, Elliott A.	26,396
Belopolsky, Ima	43,319	Kelly, Patricia L.	39,247
Benjamin, Steven C.	36,087	King, Karen K.	34,850
Birch, Linda D.	38,719	Kueller, Mark D.	31,925
Bowen, Jr., Alanson G.	24,027	Krukziel, Charles E.	27,344
Christenbury, Lynn M.	30,971	Jarnholm, Arne R.	30,396
Cotreau, William J.	36,490	Langworthy, John A.	32,255
Deitch, Gerald E.	30,457	Lerman, Bart E.	31,897
Deshmukh, Sudhir	33,677	Levitt, Cary A.	31,848
Dobson, Kevin S.	40,296	Magee, Thomas H.	27,355
Duffy, Roseanne R.	33,869	Mayer, Nancy S.	29,190
Edwards, Mark A.	39,542	Medwick, George M.	27,456
Estrin, Barry	26,452	Morrissey, Bruce W.	30,663
Evans, Craig H.	31,825	Santopietro, Lois A.	36,264
Fair, Tamara L.	35,867	Schaeffer, Andrew L.	33,605
Feltham, S. Neil	36,506	Sebree, Chytrea J.	45,348
Floyd, Linda Axamethy	33,692	Shafer, Robert J.	24,437
Frank, George A.	27,636	Shay, Lucas K.	34,724
Golian, Andrew G.	25,293	Shipley, James E.	32,003
Gorman, Thomas W.	31,959	Siegeff, Barbara C.	30,684
Gould, David J.	25,338	Sinnot, Jessica M.	34,015
Griffiths, John E.	32,647	Steinberg, Thomas W.	37,013
Hamby, Jane O.	32,872	Stevenson, Robert B.	26,039
Hamby, William H.	31,521	Strickland, Frederick D.	39,041
Heiser, David E.	31,366	Tessari, Joseph A.	32,177
Hendrickson, John S.	30,847	Tulloch, Rebecca W.	36,297
Jones, Brian C.	37,857	Walker, P. Michael	32,602
Joung, J. Kenneth	41,881	Wang, Chen	38,650

The undersigned ratifies fully all actions already taken by the above-named individuals in accordance with the authority granted hereby.

E. I. DU PONT DE NEMOURS AND COMPANY

By: 

Vernon R. Rice

Vice President and Assistant General Counsel

Date: 5/11/2010

SEQUENCE LISTING

<110> E. I. du Pont de Nemours and Company

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Gly Gly Arg Leu Lys Trp Leu Glu Arg Phe Ala Tyr Thr Asn Thr Ile
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Val Tyr Pro Phe Thr Ser Ile Pro Leu Leu Ala Tyr Cys Thr Ile Pro
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 Glu Gln Asn Gly Gln Val Cys Gln Ile Cys Gly Asp Asp Val Gly Leu
 35 40 45
 Ala Pro Gly Gly Asp Pro Phe Val Ala Cys Asn Glu Cys Ala Phe Pro
 50 55 60
 Val Cys Arg Asp Cys Tyr Glu Tyr Glu Arg Arg Glu Gly Thr Gln Asn
 65 70 75 80
 Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Leu Lys Gly Cys Gln Arg
 85 90 95
 Val Thr Gly Asp Glu Glu Glu Asp Gly Val Asp Asp Leu Asp Asn Glu
 100 105 110
 Phe Asn Trp Asp Gly His Asp Ser Gln Ser Val Ala Glu Ser Met Leu
 115 120 125
 Tyr Gly His Met Ser Tyr Gly Arg Gly Gly Asp Pro Asn Gly Ala Pro
 130 135 140
 Gln Ala Phe Gln Leu Asn Pro Asn Val Pro Leu Leu Thr Asn Gly Gln
 145 150 155 160

Met Val Asp Asp Ile Pro Pro Glu Gln His Ala Leu Val Pro Ser Phe
165 170 175

Met Gly Gly Gly Gly Lys Arg Ile His Pro Leu Pro Tyr Ala Asp Pro
180 185 190

Ser Leu Pro Val Gln Pro Arg Ser Met Asp Pro Ser Lys Asp Leu Ala
195 200 205

Ala Tyr Gly Tyr Gly Ser Val Ala Trp Lys Glu Arg Met Glu Asn Trp
210 215 220

Lys Gln Arg Gln Glu Arg Met His Gln Thr Gly Asn Asp Gly Gly Gly
225 230 235 240

Asp Asp Gly Asp Asp Ala Asp Leu Pro Leu Met Asp Glu Ala Arg Gln
245 250 255

Gln Leu Ser Arg Lys Ile Pro Leu Pro Ser Ser Gln Ile Asn Pro Tyr
260 265 270

Arg Met Ile Ile Ile Ile Arg Leu Val Val Leu Gly Phe Phe Phe His
275 280 285

Tyr Arg Val Met His Pro Val Asn Asp Ala Phe Ala Leu Trp Leu Ile
290 295 300

Ser Val Ile Cys Glu Ile Trp Phe Ala Met Ser Trp Ile Leu Asp Gln
305 310 315 320

Phe Pro Lys Trp Phe Pro Ile Glu Arg Glu Thr Tyr Leu Asp Arg Leu
325 330 335

Ser Leu Arg Phe Asp Lys Glu Gly Gln Pro Ser Gln Leu Ala Pro Ile
340 345 350

Asp Phe Phe Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro Leu Val
355 360 365

Thr Thr Asn Thr Val Leu Ser Ile Leu Ser Val Asp Tyr Pro Val Asp
370 375 380

Lys Val Ser Cys Tyr Val Ser Asp Asp Gly Ala Ala Met Leu Thr Phe
385 390 395 400

Glu Ala Leu Ser Glu Thr Ser Glu Phe Ala Lys Lys Trp Val Pro Phe
405 410 415

s Lys Arg Tyr Asn Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe Gln
420 425 430

G. s Ile Asp Tyr Leu Lys Asp Lys Val Ala Ala Asn Phe Val Arg
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Glu A r Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile
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Asn Ala Le l Ala Lys Ala Gln Lys Val Pro Glu Glu Gly Trp Thr
465 470 475 480

Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Val Arg Asp His Pro
485 490 495

Gly Met Ile Gln Val Phe Leu Gly Gln Ser Gly Gly Leu Asp Cys Glu
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Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro
515 520 525

Gly Tyr Asn His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val Arg
530 535 540

Val Ser Ala Val Leu Thr Asn Ala Pro Tyr Leu Leu Asn Leu Asp Cys
545 550 555 560

Asp His Tyr Ile Asn Asn Ser Lys Ala Ile Lys Glu Ala Met Cys Phe
565 570 575

Met Met Asp Pro Leu Leu Gly Lys Lys Val Cys Tyr Val Gln Phe Pro
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Gln Arg Phe Asp Gly Ile Asp Arg His Asp Arg Tyr Ala Asn Arg Asn
595 600 605

Val Val Phe Phe Asp Ile Asn Met Lys Gly Leu Asp Gly Ile Gln Gly
610 615 620

Pro Ile Tyr Val Gly Thr Gly Cys Val Phe Arg Arg Gln Ala Leu Tyr
625 630 635 640

Gly Tyr Asp Ala Pro Lys Thr Lys Lys Pro Pro Ser Arg Thr Cys Asn
645 650 655

Cys Trp Pro Lys Trp Cys Phe Cys Cys Cys Cys Phe Gly Asn Arg Lys
660 665 670

Gln Lys Lys Thr Thr Lys Pro Lys Thr Glu Lys Lys Lys Leu Leu Phe
675 680 685

Phe Lys Lys Glu Glu Asn Gln Ser Pro Ala Tyr Ala Leu Gly Glu Ile
690 695 700

Asp Glu Ala Ala Pro Gly Ala Glu Asn Glu Lys Ala Gly Ile Val Asn
705 710 715 720

Gln Gln Lys Leu Glu Lys Lys Phe Gly Gln Ser Ser Val Phe Val Thr
725 730 735

Ser Thr Leu Leu Glu Asn Gly Gly Thr Leu Lys Ser Ala Ser Pro Ala
740 745 750

Ser Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp
755 760 765

Lys Thr Asp Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr
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Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys His Gly Trp Arg Ser
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18

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<212> PRT
<213> Oryza sativa

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35 40 45

Ala Gly Ala Glu Val Thr Arg Asn Ala Val Val Glu Ala Ala Arg Phe
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Ala Ala Leu Trp Val Ser Phe Cys Arg Lys His Gly Val Glu Pro Arg
65 70 75 80

Asn Leu Glu Ala Tyr Phe Asn Ala Gly Glu Gly Gly Gly Gly Lys Ala
85 90 95

Lys Val Val Ala Arg Gly Ser Tyr Arg Gly Met Ala Trp Pro Glu Leu
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Val Arg Asp Arg Arg Arg Val Arg Arg Glu Tyr Glu Glu Met Arg Leu
115 120 125

Arg Ile Asp Ala Leu Gln Ala Ala Asp Ala Arg Arg Arg Arg Arg Gly
130 135 140

Ala Ala Asp Asp His Ala Gly Val Val Gln Val Leu Ile Asp Phe Ala
145 150 155 160

Gly Ser Val Pro Gln Leu Gly Val Ala Asn Gly Ser Lys Leu Ile Asp
165 170 175

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Val Ala Ser Val Asp Val Cys Leu Pro Ala Leu Val Tyr Val Cys Arg
 180 185 190

Glu Lys Arg Arg Gly His Ala His His Arg Lys Ala Gly Ala Met Asn
 195 200 205

Ala Pro Phe Ile Leu Asp Leu Asp Cys Asp Tyr Tyr Val Asn Asn Ser
 210 215 220

Gln Ala Leu Arg Ala Gly Ile Cys Phe Met Ile Glu Arg Gly Gly Gly
 225 230 235 240

Gly Ala Ala Glu Asp Ala Gly Ala Val Ala Phe Val Gln Phe Pro Gln
 245 250 255

Arg Val Asp Gly Val Asp Pro Gly Asp Arg Tyr Ala Asn His Asn Arg
 260 265 270

Val Leu Phe Asp Cys Thr Glu Leu Gly Leu Asp Gly Leu Gln Gly Pro
 275 280 285

Ile Tyr Val Gly Thr Gly Cys Leu Phe Arg Arg Val Ala Leu Tyr Ser
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Val Asp Leu Pro Arg Trp Arg Pro Arg Arg Ser Leu Gly Cys Arg Leu
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Leu Ser Gly Pro Arg
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 <213> Glycine max

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 35 40 45
 Asp Gly Asp Leu Phe Val Ala Cys Asn Glu Cys Gly Phe Pro Val Cys
 50 55 60

Arg Pro Cys Tyr Glu Tyr Glu Arg Arg Glu Gly Ser His Leu Cys Pro
 65 70 75 80
 Gln Cys Lys Thr Arg Tyr Lys Arg Leu Lys Gly Ser Pro Arg Val Glu
 85 90 95
 Gly Asp Asp Asp Glu Glu Asp Val Asp Asp Ile Glu His Glu Phe Asn
 100 105 110
 Ile Asp Glu Gln Lys Asn Lys His Gly Gln Val Ala Glu Ala Met Leu
 115 120 125
 His Gly Arg Met Ser Tyr Gly Arg Gly Pro Glu Asp Asp Asp Asn Ser
 130 135 140
 Gln Phe Pro Thr Pro Val Ile Ala Gly Gly Arg Ser Arg Pro Val Ser
 145 150 155 160
 Gly Glu Phe Pro Ile Ser Ser Asn Ala Tyr Gly Asp Gln Met Leu Ser
 165 170 175
 Ser Ser Leu His Lys Arg Val His Pro Tyr Pro Val Ser Glu Pro Gly
 180 185 190
 Ser Ala Arg Trp Asp Glu Lys Lys Xaa Asp Gly Trp Lys Asp Arg Met
 195 200 205
 Asp Asp Trp Lys Leu Gln Gln Gly Asn Leu Gly Pro Glu Pro Asp Glu
 210 215 220
 Asp Pro Asp Ala Ala Met Leu Asp Glu Ala Arg Gln Pro Leu Ser Arg
 225 230 235 240
 Lys Val Pro Ile Ala Ser Ser Lys Ile Asn Pro Tyr Arg Met Val Ile
 245 250 255
 Val Ala Arg Leu Val Ile Leu Ala Phe Phe Leu Arg Tyr Arg Leu Met
 260 265 270
 Asn Pro Val His Asp Ala Leu Gly Leu Trp Leu Thr Ser Ile Ile Cys
 275 280 285
 Glu Ile Trp Phe Ala Phe Ser Trp Ile Leu Asp Gln Phe Pro Lys Trp
 290 295 300
 Phe Pro Ile Asp Arg Glu Thr Tyr Leu Asp Arg Leu Ser Ile Arg Tyr
 305 310 315 320
 Glu Arg Glu Gly Glu Pro Asn Met Leu Ala Pro Val Asp Val Phe Val
 325 330 335
 Ser Thr Val Asp Pro Met Lys Glu Pro Pro Leu Val Thr Ala Asn Thr
 340 345 350
 Val Leu Ser Ile Leu Ala Met Asp Tyr Pro Val Asp Lys Ile Ser Cys
 355 360 365
 Tyr Ile Ser Asp Asp Gly Ala Ser Met Cys Thr Phe Glu Ser Leu Ser
 370 375 380

Glu Thr Ala Glu Phe Ala Arg Lys Trp Val Pro Phe Cys Lys Lys Phe
 385 390 395 400
 Ser Ile Glu Pro Arg Ala Pro Glu Met Tyr Phe Ser Glu Lys Ile Asp
 405 410 415
 Tyr Leu Lys Asp Lys Val Gln Pro Thr Phe Val Lys Glu Arg Arg Ala
 420 425 430
 Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala Leu Val
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 Thr Pro Trp Pro Gly Asn Asn Thr Lys Asp His Pro Gly Met Ile Gln
 465 470 475 480
 Val Phe Leu Gly Ser Ser Gly Gly Leu Asp Thr Glu Gly Asn Gln Leu
 485 490 495
 Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Phe Gln His
 500 505 510
 His Lys Lys Ala Gly Ala Met Asn Ala Leu Val Arg Val Ser Ala Val
 515 520 525
 Leu Thr Asn Ala Pro Phe Met Leu Asn Leu Asp Cys Asp His Tyr Val
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 565 570 575
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 595 600 605
 Gly Thr Gly Cys Val Phe Arg Arg Gln Ala Leu Tyr Gly Tyr Asn Pro
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 690 695 700

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 Ser Phe Trp Val Ile Val His Leu Tyr Pro Phe Leu Lys Gly Leu Met
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 Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Val Ile Trp Ser Val Leu
 995 1000 1005
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<210> 16
 <211> 610
 <212> PRT
 <213> Glycine max

<400> 16
 Ala Lys Ala Gln Lys Met Pro Glu Glu Gly Trp Thr Met Gln Asp Gly
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 Thr Pro Trp Pro Gly Asn Asn Pro Arg Asp His Pro Gly Met Ile Gln
 20 25 30

Val Phe Leu Gly His Ser Gly Gly Leu Asp Thr Asp Gly Asn Glu Leu
 35 40 45
 Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Phe Gln His
 50 55 60
 His Lys Lys Ala Gly Ala Met Asn Ala Leu Ile Arg Val Ser Ala Val
 65 70 75 80
 Leu Thr Asn Gly Ala Tyr Leu Leu Asn Val Asp Cys Asp His Tyr Phe
 85 90 95
 Asn Asn Ser Lys Ala Leu Lys Glu Ala Met Cys Phe Met Met Asp Pro
 100 105 110
 Val Leu Gly Lys Lys Thr Cys Tyr Val Gln Phe Pro Gln Arg Phe Asp
 115 120 125
 Gly Ile Asp Leu His Asp Arg Tyr Ala Asn Arg Asn Ile Val Phe Phe
 130 135 140
 Asp Ile Asn Met Lys Gly Gln Asp Gly Val Gln Gly Pro Val Tyr Val
 145 150 155 160
 Gly Thr Gly Cys Cys Phe Asn Arg Gln Ala Leu Tyr Gly Tyr Asp Pro
 165 170 175
 Val Leu Thr Glu Glu Asp Leu Glu Pro Asn Ile Ile Val Lys Ser Cys
 180 185 190
 Cys Gly Ser Arg Lys Lys Gly Lys Gly Gly Asn Lys Lys Tyr Ser Asp
 195 200 205
 Lys Lys Lys Ala Met Gly Arg Thr Glu Ser Thr Val Pro Ile Phe Asn
 210 215 220
 Met Glu Asp Ile Glu Glu Gly Val Glu Gly Tyr Asp Asp Glu Arg Thr
 225 230 235 240
 Leu Leu Met Ser Gln Lys Ser Leu Glu Lys Arg Phe Gly Gln Ser Pro
 245 250 255
 Val Phe Ile Ala Ala Thr Phe Met Glu Gln Gly Gly Ile Pro Pro Ser
 260 265 270
 Thr Asn Pro Ala Thr Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys
 275 280 285
 Gly Tyr Glu Asp Lys Thr Glu Trp Gly Lys Glu Ile Gly Trp Ile Tyr
 290 295 300
 Gly Ser Val Thr Glu Asp Ile Leu Thr Gly Phe Lys Met His Ala Arg
 305 310 315 320
 Gly Trp Ile Ser Ile Tyr Cys Met Pro Pro Arg Pro Ala Phe Lys Gly
 325 330 335
 Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu Asn Gln Val Leu Arg Trp
 340 345 350

Ala Leu Gly Ser Ile Glu Ile Phe Leu Ser Arg His Cys Pro Leu Trp
 355 360 365

Tyr Gly Tyr Asn Gly Lys Leu Lys Pro Leu Met Arg Leu Ala Tyr Ile
 370 375 380

Asn Thr Ile Val Tyr Pro Phe Thr Ser Ile Pro Leu Ile Ala Tyr Cys
 385 390 395 400

Thr Leu Pro Ala Phe Cys Leu Leu Thr Asn Lys Phe Ile Ile Pro Glu
 405 410 415

Ile Ser Asn Phe Ala Ser Met Trp Phe Ile Leu Leu Phe Val Ser Ile
 420 425 430

Phe Thr Thr Ser Ile Leu Glu Leu Arg Trp Ser Gly Val Ser Ile Glu
 435 440 445

Asp Trp Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly Thr Ser Ala
 450 455 460

His Leu Phe Ala Val Phe Gln Gly Leu Leu Lys Val Leu Ala Gly Ile
 465 470 475 480

Asp Thr Asn Phe Thr Val Thr Ser Lys Ala Ser Asp Glu Asp Gly Asp
 485 490 495

Phe Ala Glu Leu Tyr Val Phe Lys Trp Thr Ser Leu Leu Ile Pro Pro
 500 505 510

Thr Thr Val Leu Ile Val Asn Leu Val Gly Ile Val Ala Gly Val Ser
 515 520 525

Tyr Ala Ile Asn Ser Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys
 530 535 540

Leu Phe Phe Ala Ile Trp Val Ile Ala His Leu Tyr Pro Phe Leu Lys
 545 550 555 560

Gly Leu Leu Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Ile Val Trp
 565 570 575

Ser Val Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val Arg Ile Asp
 580 585 590

Pro Phe Thr Ser Asp Ser Asn Lys Leu Thr Asn Gly Gln Cys Gly Ile
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Asn Cys
 610

<210> 17
 <211> 2890
 <212> DNA
 <213> Glycine max

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 agaaacatac cttgatogtc tgtcactcag gtatgaaaaa gaagggaagc catctgagtt 180

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tgatgttgaa ggaaatgagc taccccgctt ggtctacgtt tctagagaaa agaggccagg 720
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aaaaaaaaaa 2890

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<210> 18
<211> 793
<212> PRT
<213> Glycine max

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Val Ile Cys Glu Ile Trp Phe Ala Val Ser Trp Ile Met Asp Gln Phe
          20             25             30

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Pro Lys Trp Tyr Pro Ile Gln Arg Glu Thr Tyr Leu Asp Arg Leu Ser
 35 40 45
 Leu Arg Tyr Glu Lys Glu Gly Lys Pro Ser Glu Leu Ser Ser Val Asp
 50 55 60
 Val Phe Val Ser Thr Val Asp Pro Met Lys Glu Pro Pro Leu Ile Thr
 65 70 75 80
 Ala Asn Thr Val Leu Ser Ile Leu Ala Val Asp Tyr Pro Val Asp Lys
 85 90 95
 Val Ala Cys Tyr Val Ser Asp Asp Gly Ala Ala Met Leu Thr Phe Glu
 100 105 110
 Ala Leu Ser Glu Thr Ser Glu Phe Ala Arg Arg Trp Val Pro Phe Cys
 115 120 125
 Lys Lys Tyr Asn Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe Gly Gln
 130 135 140
 Lys Met Asp Tyr Leu Lys Asn Lys Val His Pro Ala Phe Val Arg Glu
 145 150 155 160
 Arg Arg Ala Met Lys Arg Asp Tyr Glu Glu Phe Lys Val Arg Ile Asn
 165 170 175
 Ser Leu Val Ala Thr Ala Gln Lys Val Pro Glu Asp Gly Trp Thr Met
 180 185 190
 Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Val Arg Asp His Pro Gly
 195 200 205
 Met Ile Gln Val Phe Leu Gly Gln Asp Gly Val Arg Asp Val Glu Gly
 210 215 220
 Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro Gly
 225 230 235 240
 Phe Asp His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val Arg Ala
 245 250 255
 Ser Ala Ile Ile Thr Asn Ala Pro Tyr Leu Leu Asn Val Asp Cys Asp
 260 265 270
 His Tyr Ile Asn Asn Ser Lys Ala Leu Arg Glu Ala Met Cys Phe Met
 275 280 285
 Met Asp Pro Gln Leu Gly Lys Lys Val Cys Tyr Val Gln Phe Pro Gln
 290 295 300
 Arg Phe Asp Gly Ile Asp Arg His Asp Arg Tyr Ser Asn Arg Asn Val
 305 310 315 320
 Val Phe Phe Asp Ile Asn Met Lys Gly Leu Asp Gly Ile Gln Gly Pro
 325 330 335
 Ile Tyr Val Gly Thr Gly Cys Val Phe Arg Arg Tyr Ala Leu Tyr Gly
 340 345 350

Tyr Asp Ala Pro Ala Lys Lys Lys Pro Pro Ser Lys Thr Cys Asn Cys
 355 360 365
 Trp Pro Lys Trp Cys Cys Leu Cys Cys Gly Ser Arg Lys Lys Lys Asn
 370 375 380
 Ala Asn Ser Lys Lys Glu Lys Lys Arg Lys Val Lys His Ser Glu Ala
 385 390 395 400
 Ser Lys Gln Ile His Ala Leu Glu Asn Ile Glu Ala Gly Asn Glu Gly
 405 410 415
 Thr Asn Asn Glu Lys Thr Ser Asn Leu Thr Gln Thr Lys Leu Glu Lys
 420 425 430
 Arg Phe Gly Gln Ser Pro Val Phe Val Ala Ser Thr Leu Leu Asp Asp
 435 440 445
 Gly Gly Val Pro His Gly Val Ser Pro Ala Ser Leu Leu Lys Glu Ala
 450 455 460
 Ile Gln Val Ile Ser Cys Gly Tyr Glu Asp Lys Thr Glu Trp Gly Lys
 465 470 475 480
 Glu Val Gly Trp Ile Tyr Gly Ser Val Thr Glu Asp Ile Leu Thr Gly
 485 490 495
 Phe Lys Met His Cys His Gly Trp Arg Ser Val Tyr Cys Ile Pro Lys
 500 505 510
 Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu
 515 520 525
 His Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Phe Phe Ser
 530 535 540
 Arg His Cys Pro Ile Trp Tyr Gly Tyr Gly Gly Gly Leu Lys Leu Leu
 545 550 555 560
 Glu Arg Phe Ser Tyr Ile Asn Ser Val Val Tyr Pro Trp Thr Ser Leu
 565 570 575
 Pro Leu Leu Val Tyr Cys Thr Leu Pro Ala Ile Cys Leu Leu Thr Gly
 580 585 590
 Lys Phe Ile Val Pro Glu Ile Ser Asn Tyr Ala Ser Leu Val Phe Met
 595 600 605
 Ala Leu Phe Ile Ser Ile Ala Ala Thr Gly Ile Leu Glu Met Gln Trp
 610 615 620
 Gly Gly Val Ser Ile Asp Asp Trp Trp Arg Asn Glu Gln Phe Trp Val
 625 630 635 640
 Ile Gly Gly Val Ser Ser His Leu Phe Ala Leu Phe Gln Gly Leu Leu
 645 650 655
 Lys Val Leu Ala Gly Val Asn Thr Asn Phe Thr Val Thr Ser Lys Ala
 660 665 670

Ala Asp Asp Gly Glu Phe Ser Glu Leu Tyr Ile Phe Lys Trp Thr Ser
675 680 685

Leu Leu Ile Pro Pro Met Thr Leu Leu Ile Met Asn Ile Val Gly Val
690 695 700

Val Val Gly Ile Ser Asp Ala Ile Asn Asn Gly Tyr Asp Ser Trp Gly
705 710 715 720

Pro Leu Phe Gly Arg Leu Phe Phe Ala Leu Trp Val Ile Leu His Leu
725 730 735

Tyr Pro Phe Leu Lys Gly Leu Leu Gly Lys Gln Asp Arg Met Pro Thr
740 745 750

Ile Ile Leu Val Trp Ser Ile Leu Leu Ala Ser Ile Leu Thr Leu Met
755 760 765

Trp Val Arg Ile Asn Pro Phe Val Ser Arg Asp Gly Pro Val Leu Glu
770 775 780

Ile Cys Gly Leu Asn Cys Asp Glu Ser
785 790

<210> 19
<211> 1742
<212> DNA
<213> Triticum aestivum

<220>
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<222> (9)

<220>
<221> unsure
<222> (271)

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<210> 20
<211> 506
<212> PRT
<213> Triticum aestivum

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<220>
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<400> 20

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Glu Ala Met Cys Phe Leu Met Asp Pro Asn Leu Gly Pro Gln Val Cys
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Tyr Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp Arg Asn Asp Arg
          20             25             30

```

```

Tyr Ala Asn Arg Asn Thr Val Phe Phe Asp Ile Asn Leu Arg Gly Leu
      35             40             45

```

```

Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe Asn
      50             55             60

```

```

Arg Thr Ala Ile Tyr Gly Tyr Glu Pro Pro Ile Lys Ala Lys Lys Pro
      65             70             75             80

```

```

Gly Phe Leu Ala Ser Leu Cys Xaa Gly Lys Lys Lys Ala Ser Lys Ser
          85             90             95

```

```

Lys Lys Arg Ser Ser Asp Lys Lys Lys Ser Asn Lys His Val Asp Ser
      100            105            110

```

```

Ser Val Pro Val Phe Asn Leu Glu Asp Ile Glu Glu Gly Val Glu Gly
      115            120            125

```

```

Ala Gly Phe Asp Asp Glu Lys Ser Val Leu Met Ser Gln Met Ser Leu
      130            135            140

```

```

Glu Lys Arg Phe Gly Gln Ser Ala Ala Phe Val Ala Ser Thr Leu Met
      145            150            155            160

```

```

Glu Tyr Gly Gly Val Pro Gln Ser Ser Thr Pro Glu Ser Leu Leu Lys
      165            170            175

```

```

Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp Lys Ser Glu Trp
      180            185            190

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Gly Thr Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr Glu Asp Ile Leu
      195            200            205

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Thr Gly Phe Lys Met His Ala Arg Gly Trp Arg Ser Ile Tyr Cys Met
      210            215            220

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Pro Lys Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp
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 Arg Leu Asn Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Leu
 245 250 255
 Phe Ser Arg His Cys Pro Leu Trp Tyr Gly Tyr Gly Gly Arg Leu Lys
 260 265 270
 Phe Leu Glu Arg Phe Ala Tyr Ile Asn Thr Thr Ile Tyr Pro Leu Thr
 275 280 285
 Ser Leu Pro Leu Leu Val Tyr Cys Ile Leu Pro Ala Ile Cys Leu Leu
 290 295 300
 Thr Gly Lys Phe Ile Met Pro Glu Ile Ser Asn Leu Ala Ser Ile Trp
 305 310 315 320
 Phe Ile Ala Leu Phe Leu Ser Ile Phe Ala Thr Gly Ile Leu Glu Met
 325 330 335
 Arg Trp Ser Gly Val Gly Ile Asp Glu Trp Trp Arg Asn Glu Gln Phe
 340 345 350
 Trp Val Ile Gly Gly Ile Ser Ala His Leu Phe Ala Val Phe Gln Gly
 355 360 365
 Leu Leu Lys Val Leu Ala Gly Ile Asp Thr Asn Phe Thr Val Thr Ser
 370 375 380
 Lys Ala Asn Asp Glu Glu Gly Asp Phe Ala Glu Leu Tyr Met Phe Lys
 385 390 395 400
 Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Ile Leu Ile Ile Asn Met
 405 410 415
 Val Gly Val Val Ala Gly Thr Ser Tyr Ala Ile Asn Ser Gly Tyr Gln
 420 425 430
 Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe Ala Phe Trp Val Ile
 435 440 445
 Val His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg Gln Asn Arg
 450 455 460
 Thr Pro Thr Ile Val Ile Val Trp Ala Val Leu Leu Ala Ser Ile Phe
 465 470 475 480
 Ser Leu Leu Trp Val Arg Val Asp Pro Phe Thr Thr Arg Leu Ala Gly
 485 490 495
 Pro Asn Ile Gln Thr Cys Gly Ile Asn Cys
 500 505

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 <211> 1029
 <212> DNA
 <213> Triticum aestivum

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gcctggaaa 1029

<210> 22
<211> 340
<212> PRT
<213> Triticum aestivum

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Arg Ile His Pro Leu Pro Phe Ala Asp Pro Asn Leu Pro Val Gln Pro
35 40 45
Arg Ser Met Asp Pro Ser Lys Asp Leu Ala Ala Tyr Gly Tyr Gly Ser
50 55 60
Val Ala Trp Lys Glu Arg Met Glu Gly Trp Lys Gln Lys Gln Glu Arg
65 70 75 80
Leu Gln His Val Arg Ser Glu Gly Gly Gly Asp Trp Asp Gly Asp Asp
85 90 95
Ala Asp Leu Pro Leu Met Asp Glu Ala Arg Gln Pro Leu Ser Arg Lys
100 105 110
Val Pro Ile Ser Ser Ser Arg Ile Asn Pro Tyr Arg Met Ile Ile Val
115 120 125
Ile Arg Leu Val Val Leu Gly Phe Phe Phe His Tyr Arg Val Met His
130 135 140
Pro Ala Lys Asp Ala Phe Ala Leu Trp Leu Ile Ser Val Ile Cys Glu
145 150 155 160
Ile Trp Phe Ala Met Ser Cys Ile Leu Asp Gln Phe Pro Lys Trp Phe
165 170 175

Pro Ile Glu Arg Glu Thr Tyr Leu Asp Arg Leu Ser Leu Arg Phe Asp
 180 185 190

Lys Glu Gly Gln Pro Ser Gln Leu Ala Pro Ile Asp Phe Phe Val Ser
 195 200 205

Thr Val Asp Pro Thr Lys Glu Pro Pro Leu Val Thr Ala Asn Thr Val
 210 215 220

Leu Ser Ile Leu Ser Val Asp Tyr Pro Val Glu Lys Val Ser Cys Tyr
 225 230 235 240

Val Ser Asp Asp Gly Ala Ala Met Leu Thr Phe Glu Ala Leu Ser Glu
 245 250 255

Thr Ser Glu Phe Ala Lys Lys Trp Val Pro Phe Ser Lys Lys Phe Asn
 260 265 270

Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe Gln Gln Lys Ile Asp Tyr
 275 280 285

Leu Lys Asp Lys Val Ala Ala Ser Phe Val Arg Glu Arg Arg Ala Met
 290 295 300

Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala Leu Val Ala
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Lys Ala Gln Lys Val Pro Glu Glu Gly Trp Thr Met Gln Asp Gly Ser
 325 330 335

Pro Trp Pro Gly
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 <211> 1081
 <212> PRT
 <213> Arabidopsis thaliana

<400> 23
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Leu Val Arg Ile Arg His Glu Ser Asp Gly Gly Thr Lys Pro Leu Lys
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Asn Met Asn Gly Gln Ile Cys Gln Ile Cys Gly Asp Asp Val Gly Leu
 35 40 45

Ala Glu Thr Gly Asp Val Phe Val Ala Cys Asn Glu Cys Ala Phe Pro
 50 55 60

Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Lys Asp Gly Thr Gln Cys
 65 70 75 80

Cys Pro Gln Cys Lys Thr Arg Phe Arg Arg His Arg Gly Ser Pro Arg
 85 90 95

Val Glu Gly Asp Glu Asp Glu Asp Asp Val Asp Asp Ile Glu Asn Glu
 100 105 110

Phe Asn Tyr Ala Gln Gly Ala Asn Lys Ala Arg His Gln Arg His Gly
 115 120 125
 Glu Glu Phe Ser Ser Ser Ser Arg His Glu Ser Gln Pro Ile Pro Leu
 130 135 140
 Leu Thr His Gly His Thr Val Ser Gly Glu Ile Arg Thr Pro Asp Thr
 145 150 155 160
 Gln Ser Val Arg Thr Thr Ser Gly Pro Leu Gly Pro Ser Asp Arg Asn
 165 170 175
 Ala Ile Ser Ser Pro Tyr Ile Asp Pro Arg Gln Pro Val Pro Val Arg
 180 185 190
 Ile Val Asp Pro Ser Lys Asp Leu Asn Ser Tyr Gly Leu Gly Asn Val
 195 200 205
 Asp Trp Lys Glu Arg Val Glu Gly Trp Lys Leu Lys Gln Glu Lys Asn
 210 215 220
 Met Leu Gln Met Thr Gly Lys Tyr His Glu Gly Lys Gly Gly Glu Ile
 225 230 235 240
 Glu Gly Thr Gly Ser Asn Gly Glu Glu Leu Gln Met Ala Asp Asp Thr
 245 250 255
 Arg Leu Pro Met Ser Arg Val Val Pro Ile Pro Ser Ser Arg Leu Thr
 260 265 270
 Pro Tyr Arg Val Val Ile Ile Leu Arg Leu Ile Ile Leu Cys Phe Phe
 275 280 285
 Leu Gln Tyr Arg Thr Thr His Pro Val Lys Asn Ala Tyr Pro Leu Trp
 290 295 300
 Leu Thr Ser Val Ile Cys Glu Ile Trp Phe Ala Phe Ser Trp Leu Leu
 305 310 315 320
 Asp Gln Phe Pro Lys Trp Tyr Pro Ile Asn Arg Glu Thr Tyr Leu Asp
 325 330 335
 Arg Leu Ala Ile Arg Tyr Asp Arg Asp Gly Glu Pro Ser Gln Leu Val
 340 345 350
 Pro Val Asp Val Phe Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro
 355 360 365
 Leu Val Thr Ala Asn Thr Val Leu Ser Ile Leu Ser Val Asp Tyr Pro
 370 375 380
 Val Asp Lys Val Ala Cys Tyr Val Ser Asp Asp Gly Ser Ala Met Leu
 385 390 395 400
 Thr Phe Glu Ser Leu Ser Glu Thr Ala Glu Phe Ala Lys Lys Trp Val
 405 410 415
 Pro Phe Cys Lys Lys Phe Asn Ile Glu Pro Arg Ala Pro Glu Phe Tyr
 420 425 430

Phe Ala Gln Lys Ile Asp Tyr Leu Lys Asp Lys Ile Gln Pro Ser Phe
 435 440 445
 Val Lys Glu Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val
 450 455 460
 Arg Ile Asn Ala Leu Val Ala Lys Ala Gln Lys Ile Pro Glu Glu Gly
 465 470 475 480
 Trp Thr Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Thr Arg Asp
 485 490 495
 His Pro Gly Met Ile Gln Val Phe Leu Gly His Ser Gly Gly Leu Asp
 500 505 510
 Thr Asp Gly Asn Glu Leu Pro Arg Leu Ile Tyr Val Ser Arg Glu Lys
 515 520 525
 Arg Pro Gly Phe Gln His His Lys Lys Ala Gly Ala Met Asn Ala Leu
 530 535 540
 Ile Arg Val Ser Ala Val Leu Thr Asn Gly Ala Tyr Leu Leu Asn Val
 545 550 555 560
 Asp Cys Asp His Tyr Phe Asn Asn Ser Lys Ala Ile Lys Glu Ala Met
 565 570 575
 Cys Phe Met Met Asp Pro Ala Ile Gly Lys Lys Cys Cys Tyr Val Gln
 580 585 590
 Phe Pro Gln Arg Phe Asp Gly Ile Asp Leu His Asp Arg Tyr Ala Asn
 595 600 605
 Arg Asn Ile Val Phe Phe Asp Ile Asn Met Lys Gly Leu Asp Gly Ile
 610 615 620
 Gln Gly Pro Val Tyr Val Gly Thr Gly Cys Cys Phe Asn Arg Gln Ala
 625 630 635 640
 Leu Tyr Gly Tyr Asp Pro Val Leu Thr Glu Glu Asp Leu Glu Pro Asn
 645 650 655
 Ile Ile Val Lys Ser Cys Cys Gly Ser Arg Lys Lys Gly Lys Ser Ser
 660 665 670
 Lys Lys Tyr Asn Tyr Glu Lys Arg Arg Gly Ile Asn Arg Ser Asp Ser
 675 680 685
 Asn Ala Pro Leu Phe Asn Met Glu Asp Ile Asp Glu Gly Phe Glu Gly
 690 695 700
 Tyr Asp Asp Glu Arg Ser Ile Leu Met Ser Gln Arg Ser Val Glu Lys
 705 710 715 720
 Arg Phe Gly Gln Ser Pro Val Phe Ile Ala Ala Thr Phe Met Glu Gln
 725 730 735
 Gly Gly Ile Pro Pro Thr Thr Asn Pro Ala Thr Leu Leu Lys Glu Ala
 740 745 750

Ile His Val Ile Ser Cys Gly Tyr Glu Asp Lys Thr Glu Trp Gly Lys
 755 760 765
 Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr Glu Asp Ile Leu Thr Gly
 770 775 780
 Phe Lys Met His Ala Arg Gly Trp Ile Ser Ile Tyr Cys Asn Pro Pro
 785 790 795 800
 Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu
 805 810 815
 Asn Gln Val Leu Arg Trp Ala Leu Gly Ser Ile Glu Ile Leu Leu Ser
 820 825 830
 Arg His Cys Pro Ile Trp Tyr Gly Tyr His Gly Arg Leu Arg Leu Leu
 835 840 845
 Glu Arg Ile Ala Tyr Ile Asn Thr Ile Val Tyr Pro Ile Thr Ser Ile
 850 855 860
 Pro Leu Ile Ala Tyr Cys Ile Leu Pro Ala Phe Cys Leu Ile Thr Asp
 865 870 875 880
 Arg Phe Ile Ile Pro Glu Ile Ser Asn Tyr Ala Ser Ile Trp Phe Ile
 885 890 895
 Leu Leu Phe Ile Ser Ile Ala Val Thr Gly Ile Leu Glu Leu Arg Trp
 900 905 910
 Ser Gly Val Ser Ile Glu Asp Trp Trp Arg Asn Glu Gln Phe Trp Val
 915 920 925
 Ile Gly Gly Thr Ser Ala His Leu Phe Ala Val Phe Gln Gly Leu Leu
 930 935 940
 Lys Val Leu Ala Gly Ile Asp Thr Asn Phe Thr Val Thr Ser Lys Ala
 945 950 955 960
 Thr Asp Glu Asp Gly Asp Phe Ala Glu Leu Tyr Ile Phe Lys Trp Thr
 965 970 975
 Ala Leu Leu Ile Pro Pro Thr Thr Val Leu Leu Val Asn Leu Ile Gly
 980 985 990
 Ile Val Ala Gly Val Ser Tyr Ala Val Asn Ser Gly Tyr Gln Ser Trp
 995 1000 1005
 Gly Pro Leu Phe Gly Lys Leu Phe Phe Ala Leu Trp Val Ile Ala His
 1010 1015 1020
 Leu Tyr Pro Phe Leu Lys Gly Leu Leu Gly Arg Gln Asn Arg Thr Pro
 1025 1030 1035 1040
 Thr Ile Val Ile Val Trp Ser Val Leu Leu Ala Ser Ile Phe Ser Leu
 1045 1050 1055
 Leu Trp Val Arg Ile Asn Pro Phe Val Asp Ala Asn Pro Asn Ala Asn
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Asn Phe Asn Gly Lys Gly Gly Val Phe
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<211> 1084

<212> PRT

<213> Arabidopsis thaliana

<400> 24

Met Asn Thr Gly Gly Arg Leu Ile Ala Gly Ser His Asn Arg Asn Glu
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Phe Val Leu Ile Asn Ala Asp Glu Ser Ala Arg Ile Arg Ser Val Gln
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Glu Leu Ser Gly Gln Thr Cys Gln Ile Cys Gly Asp Glu Ile Glu Leu
35 40 45

Thr Val Ser Ser Glu Leu Phe Val Ala Cys Asn Glu Cys Ala Phe Pro
50 55 60

Val Cys Arg Pro Cys Tyr Glu Tyr Glu Arg Arg Glu Gly Asn Gln Ala
65 70 75 80

Cys Pro Gln Cys Lys Thr Arg Tyr Lys Arg Ile Lys Gly Ser Pro Arg
85 90 95

Val Asp Gly Asp Asp Glu Glu Glu Glu Asp Ile Asp Asp Leu Glu Tyr
100 105 110

Glu Phe Asp His Gly Met Asp Pro Glu His Ala Ala Glu Ala Ala Leu
115 120 125

Ser Ser Arg Leu Asn Thr Gly Arg Gly Gly Leu Asp Ser Ala Pro Pro
130 135 140

Gly Ser Gln Ile Pro Leu Leu Thr Tyr Cys Asp Glu Asp Ala Asp Met
145 150 155 160

Tyr Ser Asp Arg His Ala Leu Ile Val Pro Pro Ser Thr Gly Tyr Gly
165 170 175

Asn Arg Val Tyr Pro Ala Pro Phe Thr Asp Ser Ser Ala Pro Pro Gln
180 185 190

Ala Arg Ser Met Val Pro Gln Lys Asp Ile Ala Glu Tyr Gly Tyr Gly
195 200 205

Ser Val Ala Trp Lys Asp Arg Met Glu Val Trp Lys Arg Arg Gln Gly
210 215 220

Glu Lys Leu Gln Val Ile Lys His Glu Gly Gly Asn Asn Gly Arg Gly
225 230 235 240

Ser Asn Asp Asp Asp Glu Leu Asp Asp Pro Asp Met Pro Met Met Asp
245 250 255

Glu Gly Arg Gln Pro Leu Ser Arg Lys Leu Pro Ile Arg Ser Ser Arg
260 265 270

Ile Asn Pro Tyr Arg Met Leu Ile Leu Cys Arg Leu Ala Ile Leu Gly
 275 280 285
 Leu Phe Phe His Tyr Arg Ile Leu His Pro Val Asn Asp Ala Tyr Gly
 290 295 300
 Leu Trp Leu Thr Ser Val Ile Cys Glu Ile Trp Phe Ala Val Ser Trp
 305 310 315 320
 Ile Leu Asp Gln Phe Pro Lys Trp Tyr Pro Ile Glu Arg Glu Thr Tyr
 325 330 335
 Leu Asp Arg Leu Ser Leu Arg Tyr Glu Lys Glu Gly Lys Pro Ser Gly
 340 345 350
 Leu Ala Pro Val Asp Val Phe Val Ser Thr Val Asp Pro Leu Lys Glu
 355 360 365
 Pro Pro Leu Ile Thr Ala Asn Thr Val Leu Ser Ile Leu Ala Val Asp
 370 375 380
 Tyr Pro Val Asp Lys Val Ala Cys Tyr Val Ser Asp Asp Gly Ala Ala
 385 390 395 400
 Met Leu Thr Phe Glu Ala Leu Ser Asp Thr Ala Glu Phe Ala Arg Lys
 405 410 415
 Trp Val Pro Phe Cys Lys Lys Phe Asn Ile Glu Pro Arg Ala Pro Glu
 420 425 430
 Trp Tyr Phe Ser Gln Lys Met Asp Tyr Leu Lys Asn Lys Val His Pro
 435 440 445
 Ala Phe Val Arg Glu Arg Arg Ala Met Lys Arg Asp Tyr Glu Glu Phe
 450 455 460
 Lys Val Lys Ile Asn Ala Leu Val Ala Thr Ala Gln Lys Val Pro Glu
 465 470 475 480
 Glu Gly Trp Thr Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn Val
 485 490 495
 Arg Asp His Pro Gly Met Ile Gln Val Phe Leu Gly His Ser Gly Val
 500 505 510
 Arg Asp Thr Asp Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg
 515 520 525
 Glu Lys Arg Pro Gly Phe Asp His His Lys Lys Ala Gly Ala Met Asn
 530 535 540
 Ser Leu Ile Arg Val Ser Ala Val Leu Ser Asn Ala Pro Tyr Leu Leu
 545 550 555 560
 Asn Val Asp Cys Asp His Tyr Ile Asn Asn Ser Lys Ala Ile Arg Glu
 565 570 575
 Ser Met Cys Phe Met Met Asp Pro Gln Ser Gly Lys Lys Val Cys Tyr
 580 585 590

Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp Arg His Asp Arg Tyr
 595 600 605
 Ser Asn Arg Asn Val Val Phe Phe Asp Ile Asn Met Lys Gly Leu Asp
 610 615 620
 Gly Ile Gln Gly Pro Ile Tyr Val Gly Thr Gly Cys Val Phe Arg Arg
 625 630 635 640
 Gln Ala Leu Tyr Gly Phe Asp Ala Pro Lys Lys Lys Lys Pro Pro Gly
 645 650 655
 Lys Thr Cys Asn Cys Trp Pro Lys Trp Cys Cys Leu Cys Cys Gly Leu
 660 665 670
 Arg Lys Lys Ser Lys Thr Lys Ala Lys Asp Lys Lys Thr Asn Thr Lys
 675 680 685
 Glu Thr Ser Lys Gln Ile His Ala Leu Glu Asn Val Asp Glu Gly Val
 690 695 700
 Ile Val Pro Val Ser Asn Val Glu Lys Arg Ser Glu Ala Thr Gln Leu
 705 710 715 720
 Lys Leu Glu Lys Lys Phe Gly Gln Ser Pro Val Phe Val Ala Ser Ala
 725 730 735
 Val Leu Gln Asn Gly Gly Val Pro Arg Asn Ala Ser Pro Ala Cys Leu
 740 745 750
 Leu Arg Glu Ala Ile Gln Val Ile Ser Cys Gly Tyr Glu Asp Lys Thr
 755 760 765
 Glu Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr Glu Asp
 770 775 780
 Ile Leu Thr Gly Phe Lys Met His Cys His Gly Trp Arg Ser Val Tyr
 785 790 795 800
 Cys Met Pro Lys Arg Ala Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu
 805 810 815
 Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu
 820 825 830
 Ile Phe Leu Ser Arg His Cys Pro Ile Trp Tyr Gly Tyr Gly Gly Gly
 835 840 845
 Leu Lys Trp Leu Glu Arg Phe Ser Tyr Ile Asn Ser Val Val Tyr Pro
 850 855 860
 Trp Thr Ser Leu Pro Leu Ile Val Tyr Cys Ser Leu Pro Ala Val Cys
 865 870 875 880
 Leu Leu Thr Gly Lys Phe Ile Val Pro Glu Ile Ser Asn Tyr Ala Gly
 885 890 895
 Ile Leu Phe Met Leu Met Phe Ile Ser Ile Ala Val Thr Gly Ile Leu
 900 905 910

Glu Met Gln Trp Gly Gly Val Gly Ile Asp Asp Trp Trp Arg Asn Glu
 915 920 925
 Gln Phe Trp Val Ile Gly Gly Ala Ser Ser His Leu Phe Ala Leu Phe
 930 935 940
 Gln Gly Leu Leu Lys Val Leu Ala Gly Val Asn Thr Asn Phe Thr Val
 945 950 955 960
 Thr Ser Lys Ala Ala Asp Asp Gly Ala Phe Ser Glu Leu Tyr Ile Phe
 965 970 975
 Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Leu Leu Ile Ile Asn
 980 985 990
 Ile Ile Gly Val Ile Val Gly Val Ser Asp Ala Ile Ser Asn Gly Tyr
 995 1000 1005
 Asp Ser Trp Gly Pro Leu Phe Gly Arg Leu Phe Phe Ala Leu Trp Val
 1010 1015 1020
 Ile Val His Leu Tyr Pro Phe Leu Lys Gly Met Leu Gly Lys Gln Asp
 1025 1030 1035 1040
 Lys Met Pro Thr Ile Ile Val Val Trp Ser Ile Leu Leu Ala Ser Ile
 1045 1050 1055
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 20 25 30
 His Pro Ser Phe Val Lys Glu Arg Arg Ala Met Lys Arg Glu Tyr Glu
 35 40 45
 Glu Phe Lys Val Arg Ile Asn Ala Leu Val Ala Lys Ala Gln Lys Lys
 50 55 60
 Pro Glu Glu Gly Trp Val Met Gln Asp Gly Thr Pro Trp Pro Gly Asn
 65 70 75 80
 Asn Thr Arg Asp His Pro Gly Met Ile Gln Val Tyr Leu Gly Ser Ala
 85 90 95
 Gly Ala Leu Asp Val Asp Gly Lys Glu Leu Pro Arg Leu Val Tyr Val
 100 105 110

Ser Arg Glu Lys Arg Pro Gly Tyr Gln His His Lys Lys Ala Gly Ala
 115 120 125
 Glu Asn Ala Leu Val Arg Val Ser Ala Val Leu Thr Asn Ala Pro Phe
 130 135 140
 Ile Leu Asn Leu Asp Cys Asp His Tyr Ile Asn Asn Ser Lys Ala Met
 145 150 155 160
 Arg Glu Ala Met Cys Phe Leu Met Asp Pro Gln Phe Gly Lys Lys Leu
 165 170 175
 Cys Tyr Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp Arg His Asp
 180 185 190
 Arg Tyr Ala Asn Arg Asn Val Val Phe Phe Asp Ile Asn Met Leu Gly
 195 200 205
 Leu Asp Gly Leu Gln Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe
 210 215 220
 Asn Arg Gln Ala Leu Tyr Gly Tyr Asp Pro Pro Val Ser Glu Lys Arg
 225 230 235 240
 Pro Lys Met Thr Cys Asp Cys Trp Pro Ser Trp Cys Cys Cys Cys
 245 250 255
 Gly Gly Ser Arg Lys Lys Ser Lys Lys Lys Gly Glu Lys Lys Gly Leu
 260 265 270
 Leu Gly Gly Leu Leu Tyr Gly Lys Lys Lys Lys Met Met Gly Lys Asn
 275 280 285
 Tyr Val Lys Lys Gly Ser Ala Pro Val Phe Asp Leu Glu Glu Ile Glu
 290 295 300
 Glu Gly Leu Glu Gly Tyr Glu Glu Leu Glu Lys Ser Thr Leu Met Ser
 305 310 315 320
 Gln Lys Asn Phe Glu Lys Arg Phe Gly Gln Ser Pro Val Phe Ile Ala
 325 330 335
 Ser Thr Leu Met Glu Asn Gly Gly Leu Pro Glu Gly Thr Asn Ser Thr
 340 345 350
 Ser Leu Ile Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Glu
 355 360 365
 Lys Thr Glu Trp Gly Lys Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr
 370 375 380
 Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg Gly Trp Lys Ser
 385 390 395 400
 Val Tyr Cys Val Pro Lys Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile
 405 410 415
 Asn Leu Ser Asp Arg Leu His Gln Val Leu Arg Trp Ala Leu Gly Ser
 420 425 430

Val Glu Ile Phe Leu Ser Arg His Cys Pro Leu Trp Tyr Gly Tyr Gly
 435 440 445
 Gly Lys Leu Lys Trp Leu Glu Arg Leu Ala Tyr Ile Asn Thr Ile Val
 450 455 460
 Tyr Pro Phe Thr Ser Ile Pro Leu Leu Ala Tyr Cys Thr Ile Pro Ala
 465 470 475 480
 Val Cys Leu Leu Thr Gly Lys Phe Ile Ile Pro Thr Leu Ser Asn Leu
 485 490 495
 Thr Ser Val Trp Phe Leu Ala Leu Phe Leu Ser Ile Ile Ala Thr Gly
 500 505 510
 Val Leu Glu Leu Arg Trp Ser Gly Val Ser Ile Gln Asp Trp Trp Arg
 515 520 525
 Asn Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ala His Leu Phe Ala
 530 535 540
 Val Phe Gln Gly Leu Leu Lys Val Leu Ala Gly Val Asp Thr Asn Phe
 545 550 555 560
 Thr Val Thr Ala Lys Ala Ala Asp Asp Thr Glu Phe Gly Glu Leu Tyr
 565 570 575
 Leu Phe Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Leu Ile Ile
 580 585 590
 Leu Asn Met Val Gly Val Val Ala Gly Val Ser Asp Ala Ile Asn Asn
 595 600 605
 Gly Tyr Gly Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe Ala Phe
 610 615 620
 Trp Val Ile Leu His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg
 625 630 635 640
 Gln Asn Arg Thr Pro Thr Ile Val Val Leu Trp Ser Ile Leu Leu Ala
 645 650 655
 Ser Ile Phe Ser Leu Val Trp Val Arg Ile Asp Pro Phe Leu Pro Lys
 660 665 670
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 675 680 685
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 <213> Arabidopsis thaliana
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 20 25 30

Tyr Val Ser Leu Ser Arg Asp Asn Ile Glu Leu Ser Gly Glu Leu Ser
 35 40 45
 Gly Asp Tyr Ser Asn Tyr Thr Val His Ile Pro Pro Thr Pro Asp Asn
 50 55 60
 Gln Pro Met Ala Thr Lys Ala Glu Glu Gln Tyr Val Ser Asn Ser Leu
 65 70 75 80
 Phe Thr Gly Gly Phe Asn Ser Val Thr Arg Ala His Leu Met Asp Lys
 85 90 95
 Val Ile Asp Ser Asp Val Thr His Pro Gln Met Ala Gly Ala Lys Gly
 100 105 110
 Ser Ser Cys Ala Met Pro Ala Cys Asp Gly Asn Val Met Lys Asp Glu
 115 120 125
 Arg Gly Lys Asp Val Met Pro Cys Glu Cys Arg Phe Lys Ile Cys Arg
 130 135 140
 Asp Cys Phe Met Asp Ala Gln Lys Glu Thr Gly Leu Cys Pro Gly Cys
 145 150 155 160
 Lys Glu Gln Tyr Lys Ile Gly Asp Leu Asp Asp Asp Thr Pro Asp Tyr
 165 170 175
 Ser Ser Gly Ala Leu Pro Leu Pro Ala Pro Gly Lys Asp Gln Arg Gly
 180 185 190
 Asn Asn Asn Asn Met Ser Met Met Lys Arg Asn Gln Asn Gly Glu Phe
 195 200 205
 Asp His Asn Arg Trp Leu Phe Glu Thr Gln Gly Thr Tyr Gly Tyr Gly
 210 215 220
 Asn Ala Tyr Trp Pro Gln Asp Glu Met Tyr Gly Asp Asp Met Asp Glu
 225 230 235 240
 Gly Met Arg Gly Gly Met Val Glu Thr Ala Asp Lys Pro Trp Arg Pro
 245 250 255
 Leu Ser Arg Arg Ile Pro Ile Pro Ala Ala Ile Ile Ser Pro Tyr Arg
 260 265 270
 Leu Leu Ile Val Ile Arg Phe Val Val Leu Cys Phe Phe Leu Thr Trp
 275 280 285
 Arg Ile Arg Asn Pro Asn Glu Asp Ala Ile Trp Leu Trp Leu Met Ser
 290 295 300
 Ile Ile Cys Glu Leu Trp Phe Gly Phe Ser Trp Ile Leu Asp Gln Ile
 305 310 315 320
 Pro Lys Leu Cys Pro Ile Asn Arg Ser Thr Asp Leu Glu Val Leu Arg
 325 330 335
 Asp Lys Phe Asp Met Pro Ser Pro Ser Asn Pro Thr Gly Arg Ser Asp
 340 345 350

Leu Pro Gly Ile Asp Leu Phe Val Ser Thr Ala Asp Pro Glu Lys Glu
 355 360 365
 Pro Pro Leu Val Thr Ala Asn Thr Ile Leu Ser Ile Leu Ala Val Asp
 370 375 380
 Tyr Pro Val Glu Lys Val Ser Cys Tyr Leu Ser Asp Asp Gly Gly Ala
 385 390 395 400
 Leu Leu Ser Phe Glu Ala Met Ala Glu Ala Ala Ser Phe Ala Asp Leu
 405 410 415
 Trp Val Pro Phe Cys Arg Lys His Asn Ile Glu Pro Arg Asn Pro Asp
 420 425 430
 Ser Tyr Phe Ser Leu Lys Ile Asp Pro Thr Lys Asn Lys Ser Arg Ile
 435 440 445
 Asp Phe Val Lys Asp Arg Arg Lys Ile Lys Arg Glu Tyr Asp Glu Phe
 450 455 460
 Lys Val Arg Ile Asn Gly Leu Pro Asp Ser Ile Arg Arg Arg Ser Asp
 465 470 475 480
 Ala Phe Asn Ala Arg Glu Glu Met Lys Ala Leu Lys Gln Met Arg Glu
 485 490 495
 Ser Gly Gly Asp Pro Thr Glu Pro Val Lys Val Pro Lys Ala Thr Trp
 500 505 510
 Met Ala Asp Gly Thr His Trp Pro Gly Thr Trp Ala Ala Ser Thr Arg
 515 520 525
 Glu His Ser Lys Gly Asp His Ala Gly Ile Leu Gln Val Met Leu Lys
 530 535 540
 Pro Pro Ser Ser Asp Pro Leu Ile Gly Asn Ser Asp Asp Lys Val Ile
 545 550 555 560
 Asp Phe Ser Asp Thr Asp Thr Arg Leu Pro Met Phe Val Tyr Val Ser
 565 570 575
 Arg Glu Lys Arg Pro Gly Tyr Asp His Asn Lys Lys Ala Gly Ala Met
 580 585 590
 Asn Ala Leu Val Arg Ala Ser Ala Ile Leu Ser Asn Gly Pro Phe Ile
 595 600 605
 Leu Asn Leu Asp Cys Asp His Tyr Ile Tyr Asn Cys Lys Ala Val Arg
 610 615 620
 Glu Gly Met Cys Phe Met Met Asp Arg Gly Gly Glu Asp Ile Cys Tyr
 625 630 635 640
 Ile Gln Phe Pro Gln Arg Phe Glu Gly Ile Asp Pro Ser Asp Arg Tyr
 645 650 655
 Ala Asn Asn Asn Thr Val Phe Phe Asp Gly Asn Met Arg Ala Leu Asp
 660 665 670

Gly Val Gln Gly Pro Val Tyr Val Gly Thr Gly Thr Met Phe Arg Arg
 675 680 685
 Phe Ala Leu Tyr Gly Phe Asp Pro Pro Asn Pro Asp Lys Leu Leu Glu
 690 695 700
 Lys Lys Glu Ser Glu Thr Glu Ala Leu Thr Thr Ser Asp Phe Asp Pro
 705 710 715 720
 Asp Leu Asp Val Thr Gln Leu Pro Lys Arg Phe Gly Asn Ser Thr Leu
 725 730 735
 Leu Ala Glu Ser Ile Pro Ile Ala Glu Phe Gln Gly Arg Pro Leu Ala
 740 745 750
 Asp His Pro Ala Val Lys Tyr Gly Arg Pro Pro Gly Ala Leu Arg Val
 755 760 765
 Pro Arg Asp Pro Leu Asp Ala Thr Thr Val Ala Glu Ser Val Ser Val
 770 775 780
 Ile Ser Cys Trp Tyr Glu Asp Lys Thr Glu Trp Gly Asp Arg Val Gly
 785 790 795 800
 Trp Ile Tyr Gly Ser Val Thr Glu Asp Val Val Thr Gly Tyr Arg Met
 805 810 815
 His Asn Arg Gly Trp Arg Ser Val Tyr Cys Ile Thr Lys Arg Asp Ser
 820 825 830
 Phe Arg Gly Ser Ala Pro Ile Asn Leu Thr Asp Arg Leu His Gln Val
 835 840 845
 Leu Arg Trp Ala Thr Gly Ser Val Glu Ile Phe Phe Ser Arg Asn Asn
 850 855 860
 Ala Ile Leu Ala Ser Lys Arg Leu Lys Phe Leu Gln Arg Leu Ala Tyr
 865 870 875 880
 Leu Asn Val Gly Ile Tyr Pro Phe Thr Ser Leu Phe Leu Ile Leu Tyr
 885 890 895
 Cys Phe Leu Pro Ala Phe Ser Leu Phe Ser Gly Gln Phe Ile Val Arg
 900 905 910
 Thr Leu Ser Ile Ser Phe Leu Val Tyr Leu Leu Met Ile Thr Ile Cys
 915 920 925
 Leu Ile Gly Leu Ala Val Leu Glu Val Lys Trp Ser Gly Ile Gly Leu
 930 935 940
 Glu Glu Trp Trp Arg Asn Glu Gln Trp Trp Leu Ile Ser Gly Thr Ser
 945 950 955 960
 Ser His Leu Tyr Ala Val Val Gln Gly Val Leu Lys Val Ile Ala Gly
 965 970 975
 Ile Glu Ile Ser Phe Thr Leu Thr Thr Lys Ser Gly Gly Asp Asp Asn
 980 985 990

Glu Asp Ile Tyr Ala Asp Leu Tyr Ile Val Lys Trp Ser Ser Leu Met
 995 1000 1005
 Ile Pro Pro Ile Val Ile Ala Met Val Asn Ile Ile Ala Ile Val Val
 1010 1015 1020
 Ala Phe Ile Arg Thr Ile Tyr Gln Ala Val Pro Gln Trp Ser Lys Leu
 1025 1030 1035 1040
 Ile Gly Gly Ala Phe Phe Ser Phe Trp Val Leu Ala His Leu Tyr Pro
 1045 1050 1055
 Phe Ala Lys Gly Leu Met Gly Arg Arg Gly Lys Thr Pro Thr Ile Val
 1060 1065 1070
 Phe Val Trp Ala Gly Leu Ile Ala Ile Thr Ile Ser Leu Leu Trp Thr
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 Gly Gly Gly Phe Gln Phe Pro
 1105 1110
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 Asp Gly Gln Phe Cys Glu Ile Cys Gly Asp Gln Ile Gly Leu Thr Val
 35 40 45
 Glu Gly Asp Leu Phe Val Ala Cys Asn Glu Cys Gly Phe Pro Ala Cys
 50 55 60
 Arg Pro Cys Tyr Glu Tyr Glu Arg Arg Glu Gly Thr Gln Asn Cys Pro
 65 70 75 80
 Gln Cys Lys Thr Arg Tyr Lys Arg Leu Arg Gly Ser Pro Arg Val Glu
 85 90 95
 Gly Asp Glu Asp Glu Glu Asp Ile Asp Asp Ile Glu Tyr Glu Phe Asn
 100 105 110
 Ile Glu His Glu Gln Asp Lys His Lys His Ser Ala Glu Ala Met Leu
 115 120 125
 Tyr Gly Lys Met Ser Tyr Gly Arg Gly Pro Glu Asp Asp Glu Asn Gly
 130 135 140
 Arg Phe Pro Pro Val Ile Ala Gly Gly His Ser Gly Glu Phe Pro Val
 145 150 155 160

Gly Gly Gly Tyr Gly Asn Gly Glu His Gly Leu His Lys Arg Val His
 165 170 175
 Pro Tyr Pro Ser Ser Glu Ala Gly Ser Glu Gly Gly Trp Arg Glu Arg
 180 185 190
 Met Asp Asp Trp Lys Leu Gln His Gly Asn Leu Gly Pro Glu Pro Asp
 195 200 205
 Asp Asp Pro Glu Met Gly Leu Ile Asp Glu Ala Arg Gln Pro Leu Ser
 210 215 220
 Arg Lys Val Pro Ile Ala Ser Ser Lys Ile Asn Pro Tyr Arg Met Val
 225 230 235 240
 Ile Val Ala Arg Leu Val Ile Leu Ala Val Phe Leu Arg Tyr Arg Leu
 245 250 255
 Leu Asn Pro Val His Asp Ala Leu Gly Leu Trp Leu Thr Ser Val Ile
 260 265 270
 Cys Glu Ile Trp Phe Ala Val Ser Trp Ile Leu Asp Gln Phe Pro Lys
 275 280 285
 Trp Phe Pro Ile Glu Arg Glu Thr Tyr Leu Asp Arg Leu Ser Leu Arg
 290 295 300
 Tyr Glu Arg Glu Gly Glu Pro Asn Met Leu Ala Pro Val Asp Val Phe
 305 310 315 320
 Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro Leu Val Thr Ser Asn
 325 330 335
 Thr Val Leu Ser Ile Leu Ala Met Asp Tyr Pro Val Glu Lys Ile Ser
 340 345 350
 Cys Tyr Val Ser Asp Asp Gly Ala Ser Met Leu Thr Phe Glu Ser Leu
 355 360 365
 Ser Glu Thr Ala Glu Phe Ala Arg Lys Trp Val Pro Phe Cys Lys Lys
 370 375 380
 Phe Ser Ile Glu Pro Arg Ala Pro Glu Met Tyr Phe Thr Leu Lys Val
 385 390 395 400
 Asp Tyr Leu Gln Asp Lys Val His Pro Thr Phe Val Lys Glu Arg Arg
 405 410 415
 Ala Met Lys Arg Glu Tyr Glu Glu Phe Lys Val Arg Ile Asn Ala Gln
 420 425 430
 Val Ala Lys Ala Ser Lys Val Pro Leu Glu Gly Trp Ile Met Gln Asp
 435 440 445
 Gly Thr Pro Trp Pro Gly Asn Asn Thr Lys Asp His Pro Gly Met Ile
 450 455 460
 Gln Val Phe Leu Gly His Ser Gly Gly Phe Asp Val Glu Gly His Glu
 465 470 475 480

Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro Gly Phe Gln
 485 490 495
 His His Lys Lys Ala Gly Ala Met Asn Ala Leu Val Arg Val Ala Gly
 500 505 510
 Val Leu Thr Asn Ala Pro Phe Met Leu Asn Leu Asp Cys Asp His Tyr
 515 520 525
 Val Asn Asn Ser Lys Ala Val Arg Glu Ala Met Cys Phe Leu Met Asp
 530 535 540
 Pro Gln Ile Gly Lys Lys Val Cys Tyr Val Gln Phe Pro Gln Arg Phe
 545 550 555 560
 Asp Gly Ile Asp Thr Asn Asp Arg Tyr Ala Asn Arg Asn Thr Val Phe
 565 570 575
 Phe Asp Ile Asn Met Lys Gly Leu Asp Gly Ile Gln Gly Pro Val Tyr
 580 585 590
 Val Gly Thr Gly Cys Val Phe Lys Arg Gln Ala Leu Tyr Gly Tyr Glu
 595 600 605
 Pro Pro Lys Gly Pro Lys Arg Pro Lys Met Ile Ser Cys Gly Cys Cys
 610 615 620
 Pro Cys Phe Gly Arg Arg Arg Lys Asn Lys Lys Phe Ser Lys Asn Asp
 625 630 635 640
 Met Asn Gly Asp Val Ala Ala Leu Gly Gly Ala Glu Gly Asp Lys Glu
 645 650 655
 His Leu Met Phe Glu Met Asn Phe Glu Lys Thr Phe Gly Gln Ser Ser
 660 665 670
 Ile Phe Val Thr Ser Thr Leu Met Glu Glu Gly Gly Val Pro Pro Ser
 675 680 685
 Ser Ser Pro Ala Val Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys
 690 695 700
 Gly Tyr Glu Asp Lys Thr Glu Trp Gly Thr Glu Leu Gly Trp Ile Tyr
 705 710 715 720
 Gly Ser Ile Thr Glu Asp Ile Leu Thr Gly Phe Lys Met His Cys Arg
 725 730 735
 Gly Trp Arg Ser Ile Tyr Cys Met Pro Lys Arg Pro Ala Phe Lys Gly
 740 745 750
 Ser Ala Pro Ile Asn Leu Ser Asp Arg Leu Asn Gln Val Leu Arg Trp
 755 760 765
 Ala Leu Gly Ser Val Glu Ile Phe Phe Ser Arg His Ser Pro Leu Trp
 770 775 780
 Tyr Gly Tyr Lys Gly Gly Lys Leu Lys Trp Leu Glu Arg Phe Ala Tyr
 785 790 795 800

Ala Asn Thr Thr Ile Tyr Pro Phe Thr Ser Ile Pro Leu Leu Ala Tyr
805 810 815

Cys Ile Leu Pro Ala Ile Cys Leu Leu Thr Asp Lys Phe Ile Met Pro
820 825 830

Pro Ile Ser Thr Phe Ala Ser Leu Phe Phe Ile Ser Leu Phe Met Ser
835 840 845

Ile Ile Val Thr Gly Ile Leu Glu Leu Arg Trp Ser Gly Val Ser Ile
850 855 860

Glu Glu Trp Trp Arg Asn Glu Gln Phe Trp Val Ile Gly Gly Ile Ser
865 870 875 880

Ala His Leu Phe Ala Val Val Gln Gly Leu Leu Lys Ile Leu Ala Gly
885 890 895

Ile Asp Thr Asn Phe Thr Val Thr Ser Lys Ala Thr Asp Asp Asp Asp
900 905 910

Phe Gly Glu Leu Tyr Ala Phe Lys Trp Thr Thr Leu Leu Ile Pro Pro
915 920 925

Thr Thr Val Leu Ile Ile Asn Ile Val Gly Val Val Ala Gly Ile Ser
930 935 940

Asp Ala Ile Asn Asn Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys
945 950 955 960

Leu Phe Phe Ser Phe Trp Val Ile Val His Leu Tyr Pro Phe Leu Lys
965 970 975

Gly Leu Met Gly Arg Gln Asn Arg Thr Pro Thr Ile Val Val Ile Trp
980 985 990

Ser Val Leu Leu Ala Ser Ile Phe Ser Leu Leu Trp Val Arg Ile Asp
995 1000 1005

Pro Phe Val Leu Lys Thr Lys Gly Pro Asp Thr Ser Lys Cys Gly Ile
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Asn Cys
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<212> PRT
<213> Gossypium hirsutum

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35 40 45

Glu Trp Tyr Phe Ala Gln Lys Ile Asp Tyr Leu Lys Asp Lys Val Gln
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 Thr Ser Phe Val Lys Glu Arg Arg Ala Met Lys Arg Glu Tyr Glu Glu
 65 70 75 80
 Phe Lys Val Arg Val Asn Gly Leu Val Ala Lys Ala Gln Lys Val Pro
 85 90 95
 Glu Glu Gly Trp Ile Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Asn
 100 105 110
 Thr Arg Asp His Pro Gly Met Ile Gln Val Phe Leu Gly Gln Ser Gly
 115 120 125
 Gly Leu Asp Ala Glu Gly Asn Glu Leu Pro Arg Leu Val Tyr Val Ser
 130 135 140
 Arg Glu Lys Arg Pro Gly Phe Gln His His Lys Lys Ala Gly Ala Met
 145 150 155 160
 Asn Ala Leu Val Arg Val Ser Ala Val Leu Thr Asn Gly Ala Phe Leu
 165 170 175
 Leu Asn Leu Asp Cys Asp His Tyr Ile Asn Asn Ser Lys Ala Leu Arg
 180 185 190
 Glu Ala Met Cys Phe Leu Met Asp Pro Asn Leu Gly Lys Gln Val Cys
 195 200 205
 Tyr Val Gln Phe Pro Gln Arg Phe Asp Gly Ile Asp Arg Asn Asp Arg
 210 215 220
 Tyr Ala Asn Arg Asn Thr Val Phe Phe Asp Ile Asn Leu Arg Gly Leu
 225 230 235 240
 Asp Gly Ile Gln Gly Pro Val Tyr Val Gly Thr Gly Cys Val Phe Asn
 245 250 255
 Arg Thr Ala Leu Tyr Gly Tyr Glu Pro Pro Leu Lys Pro Lys His Arg
 260 265 270
 Lys Thr Gly Ile Leu Ser Ser Leu Cys Gly Gly Ser Arg Lys Lys Ser
 275 280 285
 Ser Lys Ser Ser Lys Lys Gly Ser Asp Lys Lys Lys Ser Gly Lys His
 290 295 300
 Val Asp Ser Thr Val Pro Val Phe Asn Leu Glu Asp Ile Glu Glu Gly
 305 310 315 320
 Val Glu Gly Ala Gly Phe Asp Asp Glu Lys Ser Leu Leu Met Ser Gln
 325 330 335
 Met Ser Leu Glu Lys Arg Phe Gly Gln Ser Ala Val Phe Val Ala Ser
 340 345 350
 Thr Leu Met Glu Asn Gly Gly Val Pro Gln Ser Ala Thr Pro Glu Thr
 355 360 365

Leu Leu Lys Glu Ala Ile His Val Ile Ser Cys Gly Tyr Glu Asp Lys
 370 375 380
 Thr Asp Trp Gly Ser Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr Glu
 385 390 395 400
 Asp Ile Leu Thr Gly Phe Lys Met His Ala Arg Gly Trp Arg Ser Ile
 405 410 415
 Tyr Cys Met Pro Lys Arg Pro Ala Phe Lys Gly Ser Ala Pro Ile Asn
 420 425 430
 Leu Ser Asp Arg Leu Asn Gln Val Leu Arg Trp Ala Leu Gly Ser Val
 435 440 445
 Glu Ile Leu Phe Ser Arg His Cys Pro Ile Trp Tyr Gly Tyr Ser Gly
 450 455 460
 Arg Leu Lys Trp Leu Glu Arg Phe Ala Tyr Val Asn Thr Thr Ile Tyr
 465 470 475 480
 Pro Val Thr Ala Ile Pro Leu Leu Met Tyr Cys Thr Leu Pro Ala Val
 485 490 495
 Cys Leu Leu Thr Asn Lys Phe Ile Ile Pro Gln Ile Ser Asn Leu Ala
 500 505 510
 Ser Ile Trp Phe Ile Ser Leu Phe Leu Ser Ile Phe Ala Thr Gly Ile
 515 520 525
 Leu Lys Met Lys Trp Asn Gly Val Gly Ile Asp Gln Trp Trp Arg Asn
 530 535 540
 Glu Gln Phe Trp Val Ile Gly Gly Val Ser Ala His Leu Phe Ala Val
 545 550 555 560
 Phe Gln Gly Leu Leu Lys Val Leu Ala Gly Ile Asp Thr Asn Phe Thr
 565 570 575
 Val Thr Ser Lys Ala Ser Asp Glu Asp Gly Asp Phe Ala Glu Leu Tyr
 580 585 590
 Met Phe Lys Trp Thr Thr Leu Leu Ile Pro Pro Thr Thr Leu Leu Ile
 595 600 605
 Ile Asn Leu Val Gly Val Val Ala Gly Ile Ser Tyr Val Ile Asn Ser
 610 615 620
 Gly Tyr Gln Ser Trp Gly Pro Leu Phe Gly Lys Leu Phe Phe Ala Phe
 625 630 635 640
 Trp Val Ile Ile His Leu Tyr Pro Phe Leu Lys Gly Leu Met Gly Arg
 645 650 655
 Gln Asn Arg Thr Pro Thr Ile Val Val Val Trp Ser Ile Leu Leu Ala
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 Ser Ile Phe Ser Leu Leu Trp Val Arg Ile Asp Pro Phe Thr Thr Arg
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Val Thr Gly Pro Asp Val Glu Gln Cys Gly Ile Asn Cys
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<211> 1081

<212> PRT

<213> Arabidopsis thaliana

<400> 29

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Gly Gln Thr Cys Gln Ile Cys Arg Asp Glu Ile Glu Leu Thr Val Asp
 35 40 45

Gly Glu Pro Phe Val Ala Cys Asn Glu Cys Ala Phe Pro Val Cys Arg
 50 55 60

Pro Cys Tyr Glu Tyr Glu Arg Arg Glu Gly Asn Gln Ala Cys Pro Gln
 65 70 75 80

Cys Lys Thr Arg Phe Lys Arg Leu Lys Gly Ser Pro Arg Val Glu Gly
 85 90 95

Asp Glu Glu Glu Asp Asp Ile Asp Asp Leu Asp Asn Glu Phe Glu Tyr
 100 105 110

Gly Asn Asn Gly Ile Gly Phe Asp Gln Val Ser Glu Gly Met Ser Ile
 115 120 125

Ser Arg Arg Asn Ser Gly Phe Pro Gln Ser Asp Leu Asp Ser Ala Pro
 130 135 140

Pro Gly Ser Gln Ile Pro Leu Leu Thr Tyr Gly Asp Glu Asp Val Glu
 145 150 155 160

Ile Ser Ser Asp Arg His Ala Leu Ile Val Pro Pro Ser Leu Gly Gly
 165 170 175

His Gly Asn Arg Val His Pro Val Ser Leu Ser Asp Pro Thr Val Ala
 180 185 190

Ala His Arg Arg Leu Met Val Pro Gln Lys Asp Leu Ala Val Tyr Gly
 195 200 205

Tyr Gly Ser Val Ala Trp Lys Asp Arg Met Glu Glu Trp Lys Arg Lys
 210 215 220

Gln Asn Glu Lys Leu Gln Val Val Arg His Glu Gly Asp Pro Asp Phe
 225 230 235 240

Glu Asp Gly Asp Asp Ala Asp Phe Pro Met Met Asp Glu Gly Arg Gln
 245 250 255

Pro Leu Ser Met Lys Ile Pro Ile Lys Ser Ser Lys Ile Asn Pro Tyr
 260 265 270

Arg Met Leu Ile Val Leu Arg Leu Val Ile Leu Gly Leu Phe Phe His
 275 280 285
 Tyr Arg Ile Leu His Pro Val Lys Asp Ala Tyr Ala Leu Trp Leu Ile
 290 295 300
 Ser Val Ile Cys Glu Ile Trp Phe Ala Val Ser Trp Val Leu Asp Gln
 305 310 315 320
 Phe Pro Lys Trp Tyr Pro Ile Glu Arg Glu Thr Tyr Leu Asp Arg Leu
 325 330 335
 Ser Leu Arg Tyr Glu Lys Glu Gly Lys Pro Ser Gly Leu Ser Pro Val
 340 345 350
 Asp Val Phe Val Ser Thr Val Asp Pro Leu Lys Glu Pro Pro Leu Ile
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 Thr Ala Asn Thr Val Leu Ser Ile Leu Ala Val Asp Tyr Pro Val Asp
 370 375 380
 Lys Val Ala Cys Tyr Val Ser Asp Asp Gly Ala Ala Met Leu Thr Phe
 385 390 395 400
 Glu Ala Leu Ser Glu Thr Ala Glu Phe Ala Arg Lys Trp Val Pro Phe
 405 410 415
 Cys Lys Lys Tyr Cys Ile Glu Pro Arg Ala Pro Glu Trp Tyr Phe Cys
 420 425 430
 His Lys Met Asp Tyr Leu Lys Asn Lys Val His Pro Ala Phe Val Arg
 435 440 445
 Glu Arg Arg Ala Met Lys Arg Asp Tyr Glu Glu Phe Lys Val Lys Ile
 450 455 460
 Asn Ala Leu Val Ala Thr Ala Gln Lys Val Pro Glu Asp Gly Trp Thr
 465 470 475 480
 Met Gln Asp Gly Thr Pro Trp Pro Gly Asn Ser Val Arg Asp His Pro
 485 490 495
 Gly Met Ile Gln Val Phe Leu Gly Ser Asp Gly Val Arg Asp Val Glu
 500 505 510
 Asn Asn Glu Leu Pro Arg Leu Val Tyr Val Ser Arg Glu Lys Arg Pro
 515 520 525
 Gly Phe Asp His His Lys Lys Ala Gly Ala Met Asn Ser Leu Ile Arg
 530 535 540
 Val Ser Gly Val Leu Ser Asn Ala Pro Tyr Leu Leu Asn Val Asp Cys
 545 550 555 560
 Asp His Tyr Ile Asn Asn Ser Lys Ala Leu Arg Glu Ala Met Cys Phe
 565 570 575
 Met Met Asp Pro Gln Ser Gly Lys Lys Ile Cys Tyr Val Gln Phe Pro
 580 585 590

Gln Arg Phe Asp Gly Ile Asp Arg His Asp Arg Tyr Ser Asn Arg Asn
 595 600 605
 Val Val Phe Phe Asp Ile Asn Met Lys Gly Leu Asp Gly Leu Gln Gly
 610 615 620
 Pro Ile Tyr Val Gly Thr Gly Cys Val Phe Arg Arg Gln Ala Leu Tyr
 625 630 635 640
 Gly Phe Asp Ala Pro Lys Lys Lys Lys Gly Pro Arg Lys Thr Cys Asn
 645 650 655
 Cys Trp Pro Lys Trp Cys Leu Leu Cys Phe Gly Ser Arg Lys Asn Arg
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 Lys Ala Lys Thr Val Ala Ala Asp Lys Lys Lys Lys Asn Arg Glu Ala
 675 680 685
 Ser Lys Gln Ile His Ala Leu Glu Asn Ile Glu Glu Gly Arg Gly His
 690 695 700
 Lys Val Leu Asn Val Glu Gln Ser Thr Glu Ala Met Gln Met Lys Leu
 705 710 715 720
 Gln Lys Lys Tyr Gly Gln Ser Pro Val Phe Val Ala Ser Ala Arg Leu
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 Glu Asn Gly Gly Met Ala Arg Asn Ala Ser Pro Ala Cys Leu Leu Lys
 740 745 750
 Glu Ala Ile Gln Val Ile Ser Arg Gly Tyr Glu Asp Lys Thr Glu Trp
 755 760 765
 Gly Lys Glu Ile Gly Trp Ile Tyr Gly Ser Val Thr Glu Asp Ile Leu
 770 775 780
 Thr Gly Ser Lys Met His Ser His Gly Trp Arg His Val Tyr Cys Thr
 785 790 795 800
 Pro Lys Leu Ala Ala Phe Lys Gly Ser Ala Pro Ile Asn Leu Ser Asp
 805 810 815
 Arg Leu His Gln Val Leu Arg Trp Ala Leu Gly Ser Val Glu Ile Phe
 820 825 830
 Leu Ser Arg His Cys Pro Ile Trp Tyr Gly Tyr Gly Gly Gly Leu Lys
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 Trp Leu Glu Arg Leu Ser Tyr Ile Asn Ser Val Val Tyr Pro Trp Thr
 850 855 860
 Ser Leu Pro Leu Ile Val Tyr Cys Ser Leu Pro Ala Ile Cys Leu Leu
 865 870 875 880
 Thr Gly Lys Phe Ile Val Pro Glu Ile Ser Asn Tyr Ala Ser Ile Leu
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Gln Trp Gly Lys Val Gly Ile Asp Asp Trp Trp Arg Asn Glu Gln Phe
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 Leu Leu Lys Val Leu Ala Gly Val Asp Thr Asn Phe Thr Val Thr Ser
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 Lys Ala Ala Asp Asp Gly Glu Phe Ser Asp Leu Tyr Leu Phe Lys Trp
 965 970 975
 Thr Ser Leu Leu Ile Pro Pro Met Thr Leu Leu Ile Ile Asn Val Ile
 980 985 990
 Gly Val Ile Val Gly Val Ser Asp Ala Ile Ser Asn Gly Tyr Asp Ser
 995 1000 1005
 Trp Gly Pro Leu Phe Gly Arg Leu Phe Phe Ala Leu Trp Val Ile Ile
 1010 1015 1020
 His Leu Tyr Pro Phe Leu Lys Gly Leu Leu Gly Lys Gln Asp Arg Met
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 Pro Thr Ile Ile Val Val Trp Ser Ile Leu Leu Ala Ser Ile Leu Thr
 1045 1050 1055
 Leu Leu Trp Val Arg Val Asn Pro Phe Val Ala Lys Gly Gly Pro Ile
 1060 1065 1070
 Leu Glu Ile Cys Gly Leu Asp Cys Leu
 1075 1080